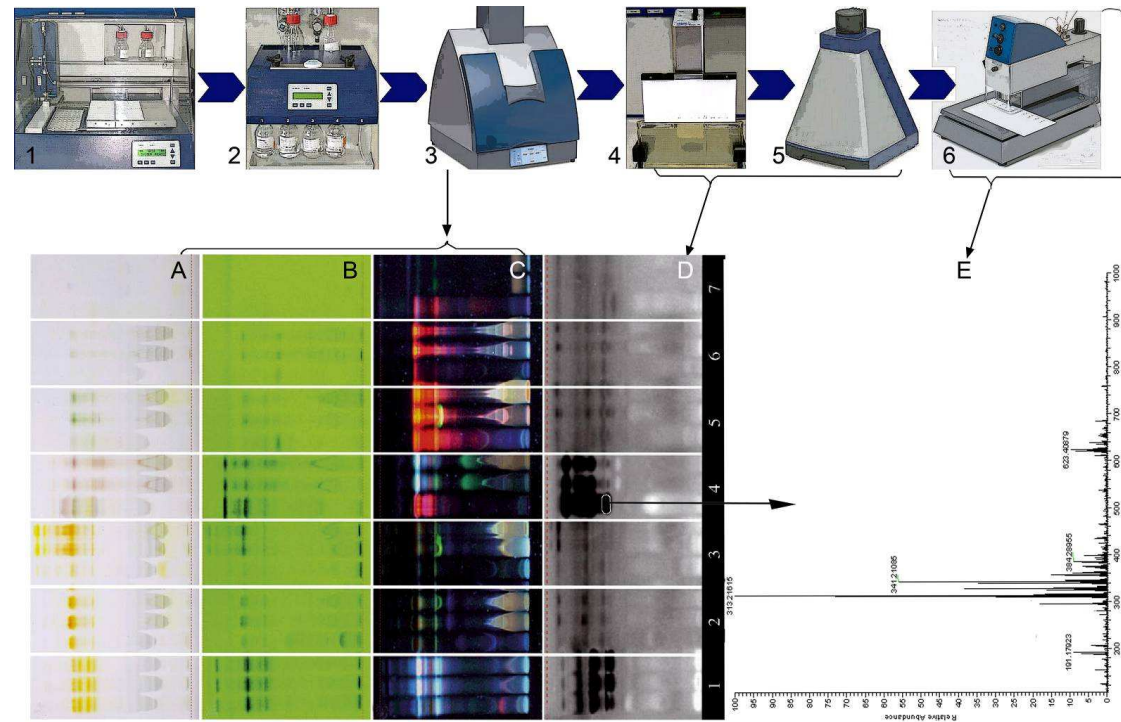


Hyphenated HPTLC methods for identification and structure elucidation



PD Dr. Gertrud Morlock,
 Institute of Food Chemistry
 University of Hohenheim, Stuttgart



Hyphenation

- 1980: term hyphenation by Hirschfeld
- comprises the different approaches to combine mainly spectrometers with chromatographic systems to get further information about the sample
- **hyphen** (-) symbolizes this attempt of combination, which did not reach its stage of full maturity so far
- **slash** (/) is found for hyphenated methods at a mature state
- 2007: term “hypernation“ (super-hyphenation) by Wilson and Brinkman
→ to place all of the required spectrometers into a single system
so that all of the spectroscopic information is obtained in a single run



Hyphenation

Problems associated with column-based hypernations are:

- Capital cost and strategies for dealing with the large amounts of data produced by such systems.
- Complexity of the instrumentation increases → difficult to operate in routine
- A single eluent (→ optimal for all detectors) is difficult to obtain.
- Differences in sensitivity are challenging.

All these problems are less challenging in HPTLC-based hypernations:

- Open system is highly adaptive to different sensitivities
- Cost-effective by modular instrumentation
- Generating less data due to targeted access to points-of-care on the plate
- Directly accessible for the respective optimal solvent

=> The main difference:

HPLC: sample in solvent; after separation → sample in the waste

HPTLC: solvent evaporated; after separation → still on the plate.



G Model

CHROMA-351018; No. of Pages 10

ARTICLE IN PRESS

Journal of Chromatography A, xxx (2010) xxx–xxx



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Contents lists available at ScienceDirect

Journal of Chromatography A

journal homepage: www.elsevier.com/locate/chroma



Review

Hyphenations in planar chromatography

Gertrud Morlock*, Wolfgang Schwack

University of Hohenheim, Institute of Food Chemistry, Garbenstrasse 28, 70599 Stuttgart, Germany

- HPTLC-UV/Vis/FLD-MS [13,14],
- HPTLC-UV/Vis/FLD-bioactivity-HRMS [15],
- HPTLC-UV-FTIR [16,17],
- HPTLC-UV/Vis/FLD-FTIR ATR [18],
- TLC-Vis-SERS [12].

ARTICLE INFO

Article history:

Available online xxx

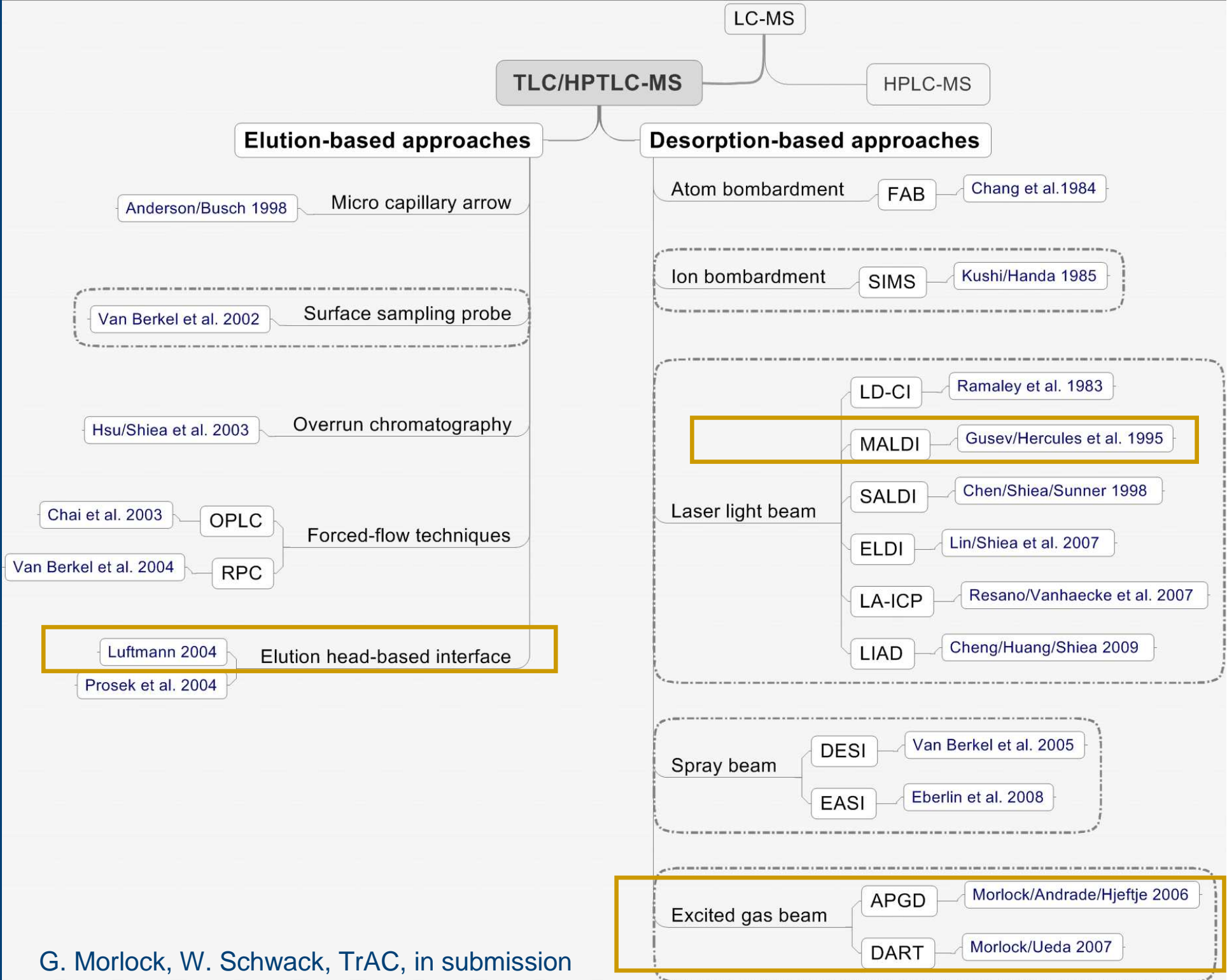
Keywords:

Mass spectrometry
High-performance thin-layer chromatography
Effect-directed analysis
Bioassays
Cost-effective analysis
High-throughput system

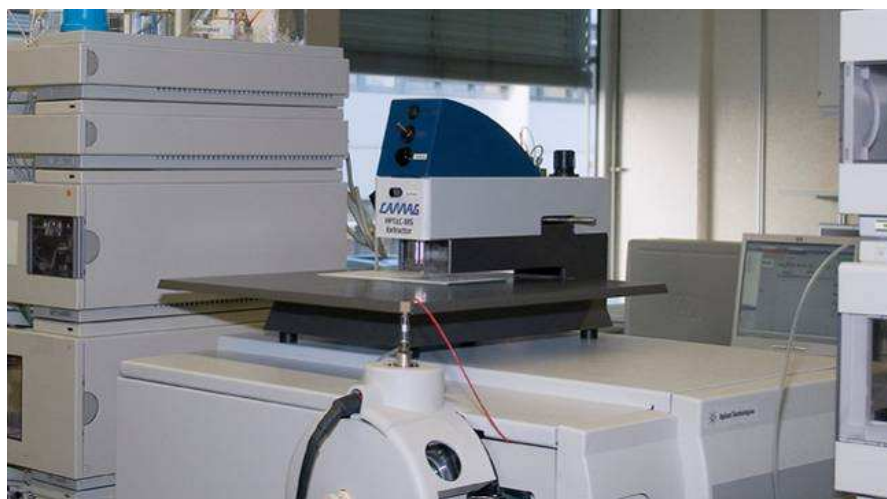
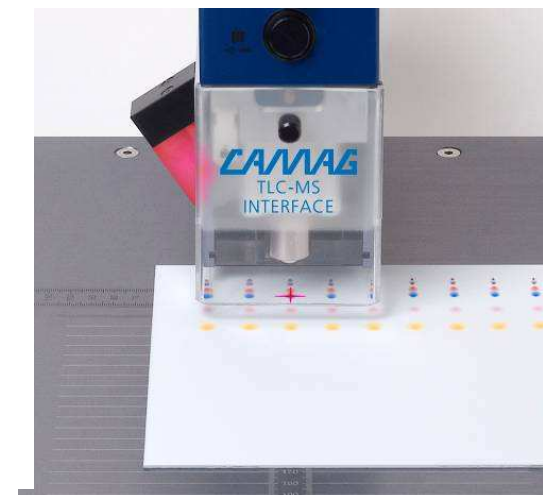
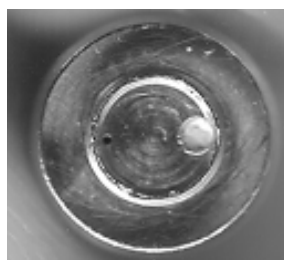
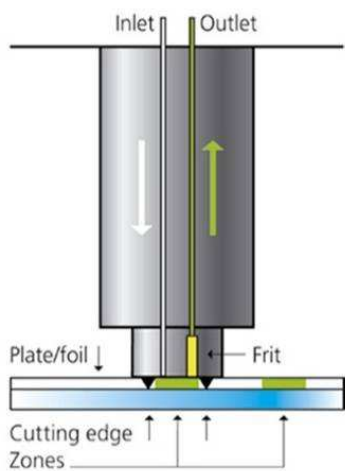
ABSTRACT

This review is focused on planar chromatography and especially on its most important subcategory high-performance thin-layer chromatography (HPTLC). The image-giving format of the open, planar stationary phase and the post-chromatographic evaporation of the mobile phase ease the performance of various kinds of hyphenations and even super-hyphenations. Examples in the field of natural product search, food and lipid analysis are demonstrated, which point out the hyphenation with effect-directed analysis (EDA) and mass spectrometry and illustrate the efficiency gain. Depending on the task at hand, hyphenations can readily be selected as required to reach the relevant information about the sample, and at the same time, information is obtained for many samples in parallel. The flexibility and the unrivalled features through the planar format valuably assist separation scientists.

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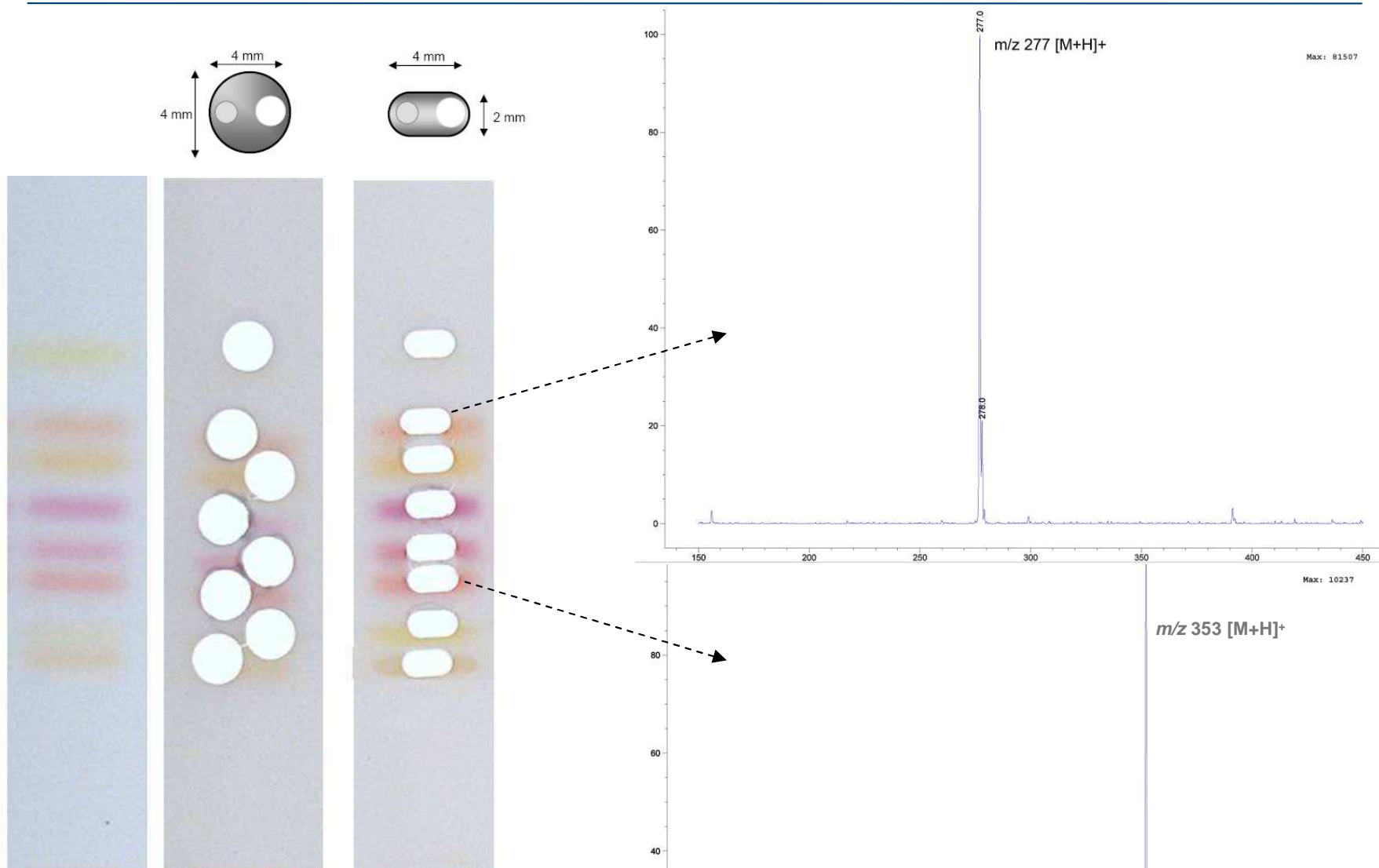
Elution head-based HPTLC-MS → TLC-MS Interface



H. Luftmann, Anal Bioanal Chem 378 (2004) 964-968

A. Alpmann, G. Morlock, Anal Bioanal Chem 386 (2006) 1543-1551

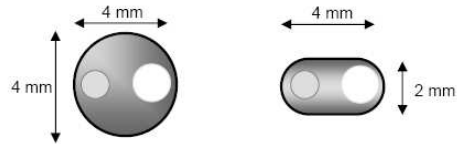
Elution head-based HPTLC-MS



G. Morlock , CBS 103 (2009) 16

Elution head

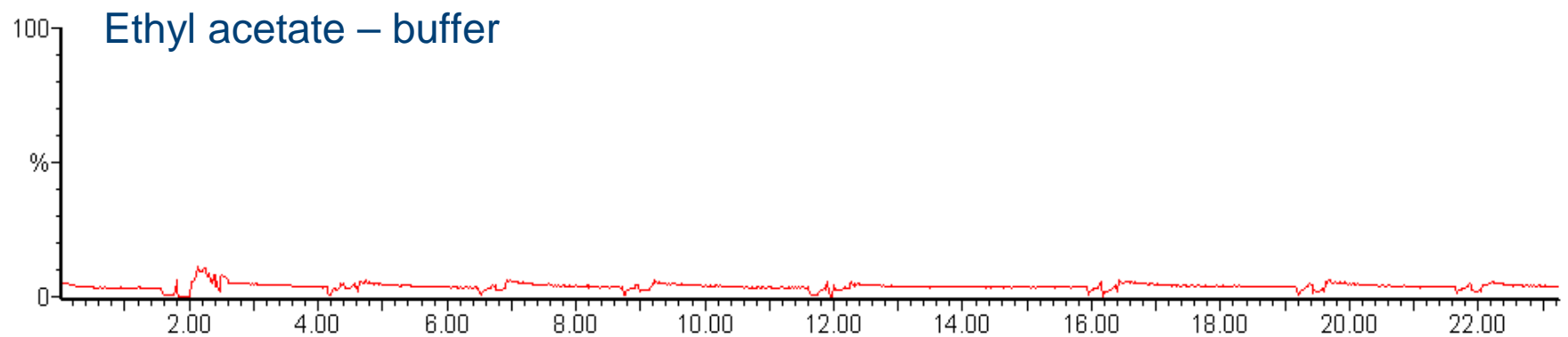
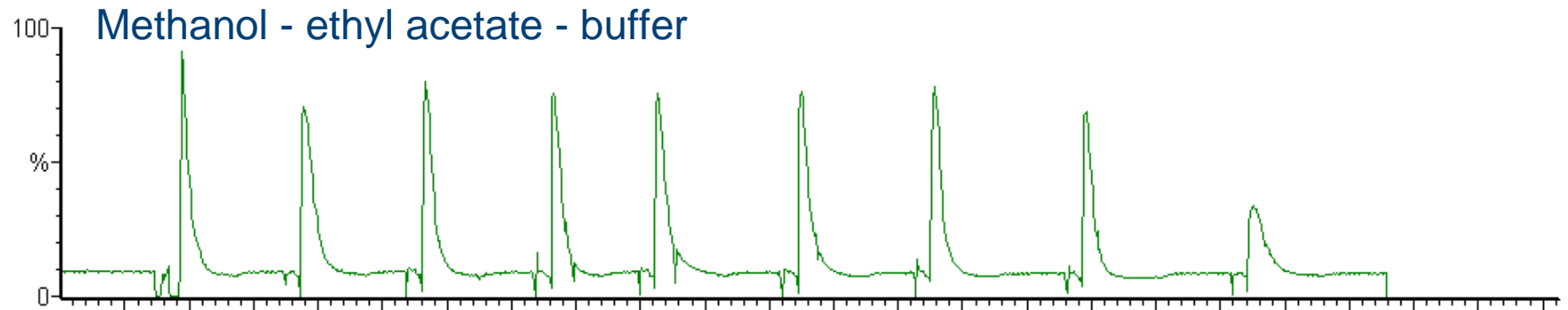
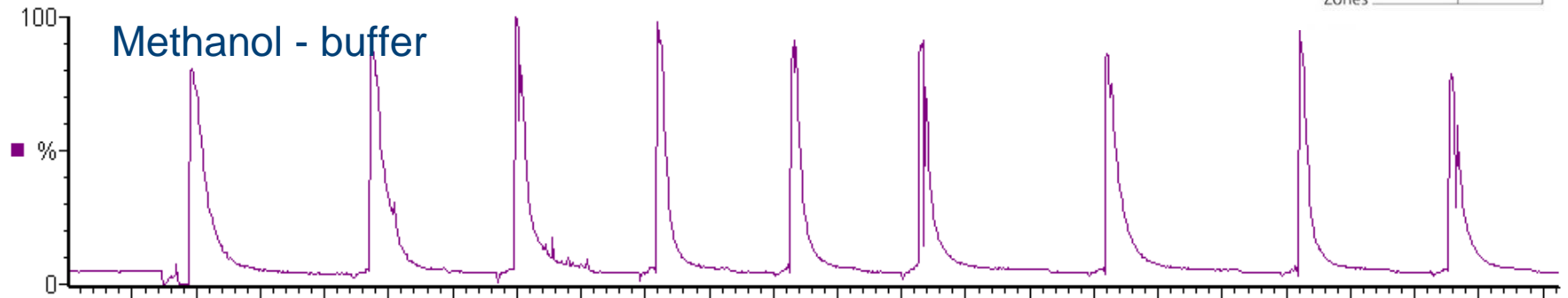
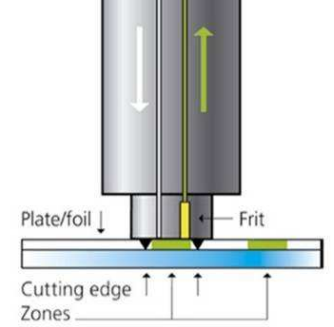
Cutting edge geometry → U. Jautz, G. Morlock, J Planar Chromatogr 21 (2008) 367



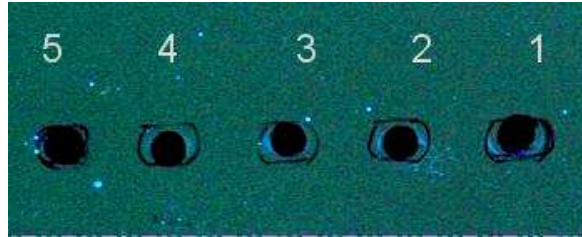
Cutting edge height

- 0.2 mm for standard layers → CAMAG Bibliography Service CBS 102 (2009)
- 0.1 mm for extra thin layers → U. Jautz, G. Morlock, Anal Bioanal Chem 387 (2007) 1083
- 0.5 mm for preparative layers → E. Dytkeiwitz, G. Morlock, J AOAC Int 91 (2008) 1237

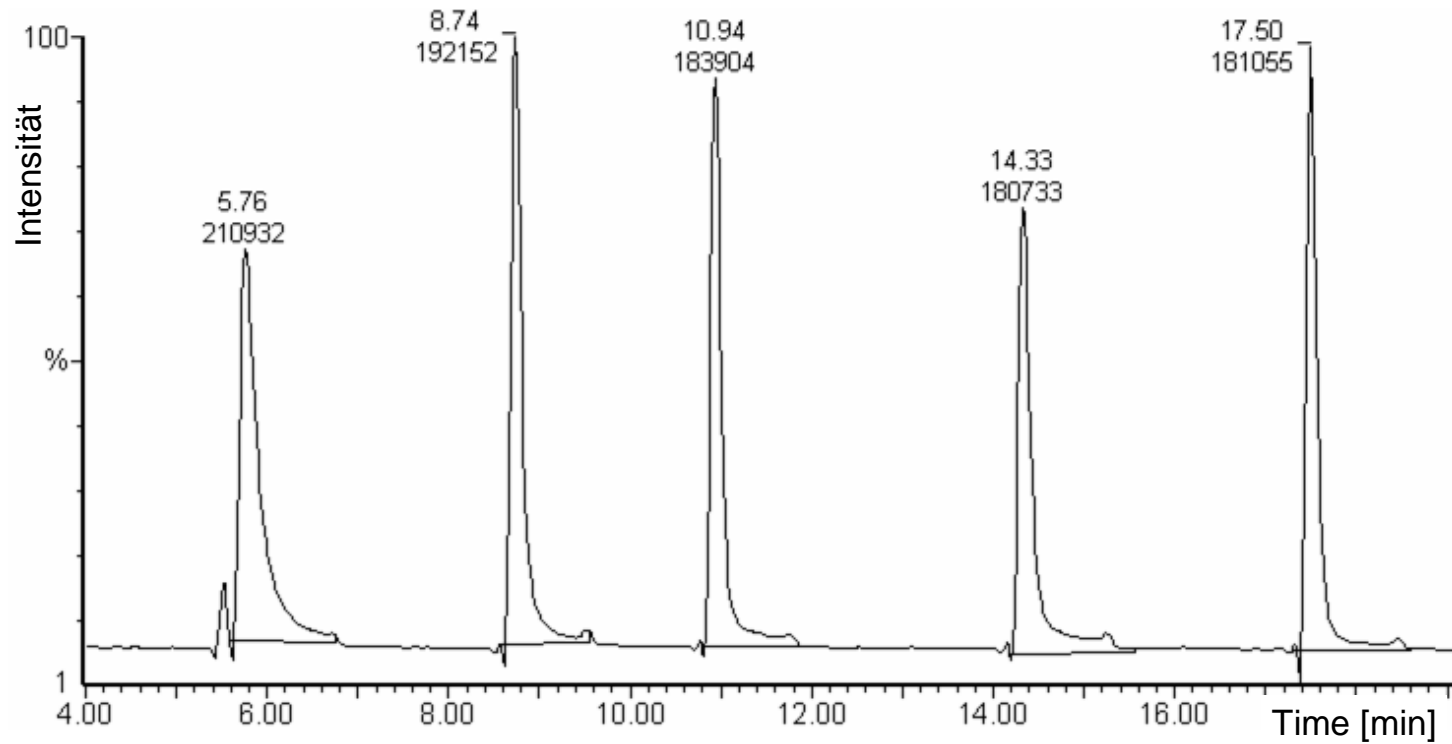
Elution profiles with different solvents



Repeatability of the extraction



→ %RSD = 6.7 % (6 ng/zone, n = 5)



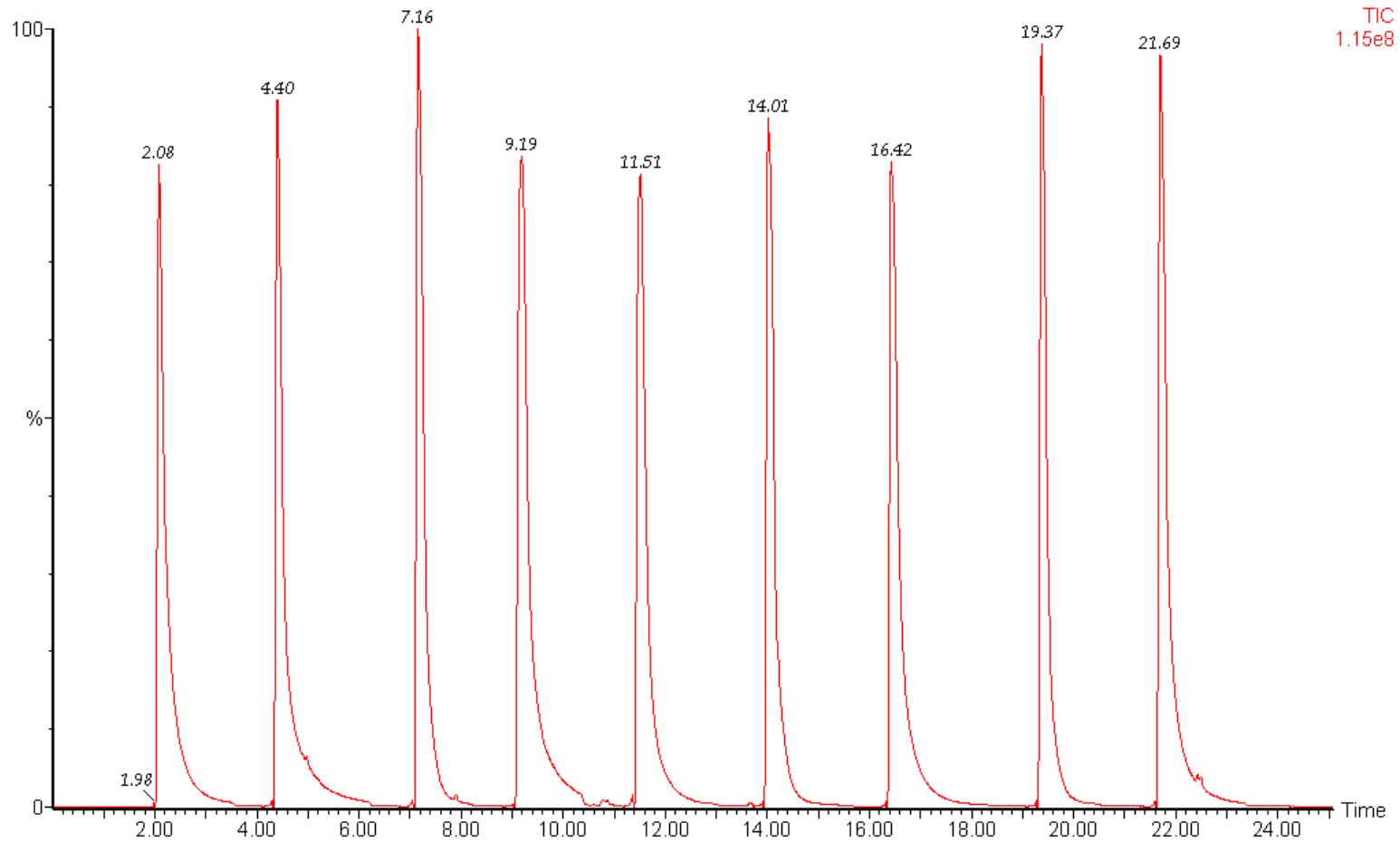
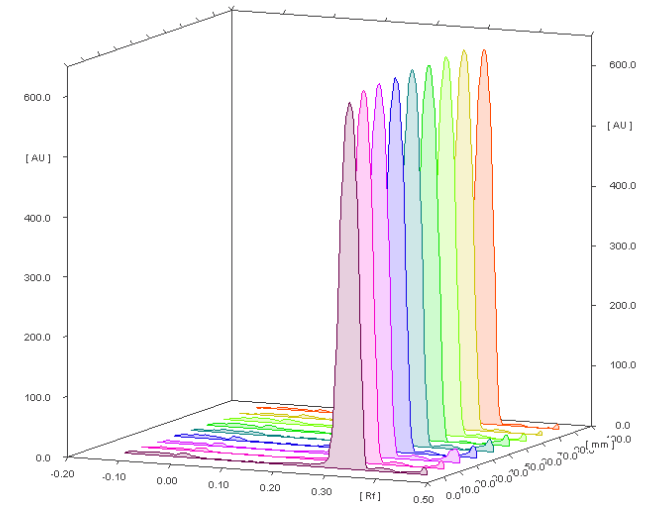
SIM elution profile of ITX @ m/z 255 [M+H]⁺ and 277 [M+Na]⁺



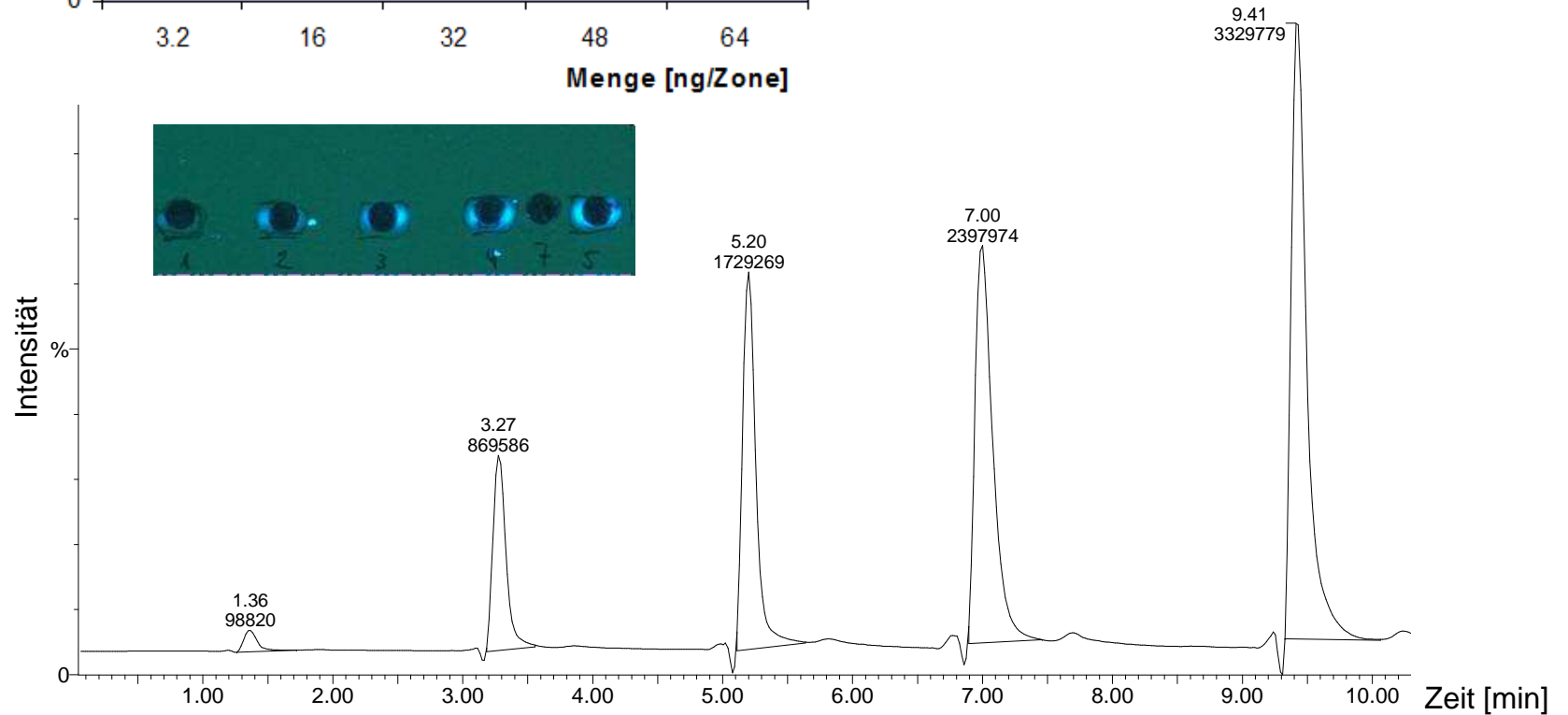
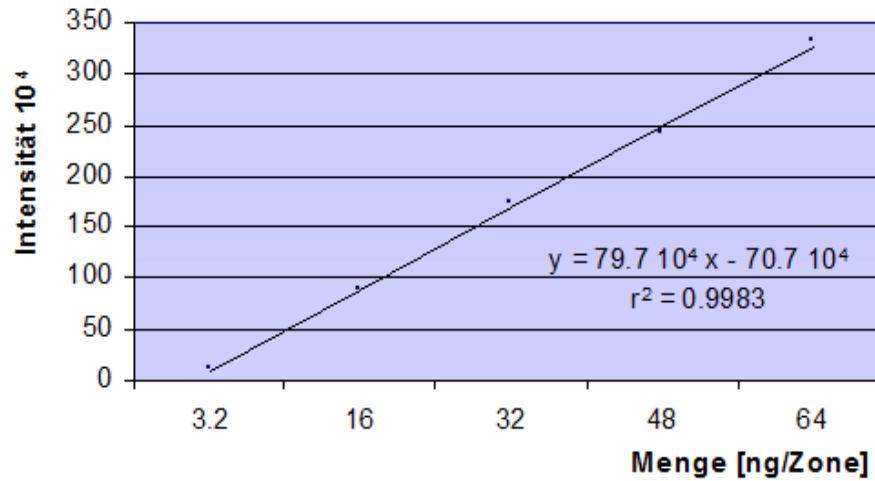
Repeatability of extraction

SIM @ m/z 329 $[M+Na]^+$

Repeatability (%RSD, $n = 9$): 7 %



Analytical response

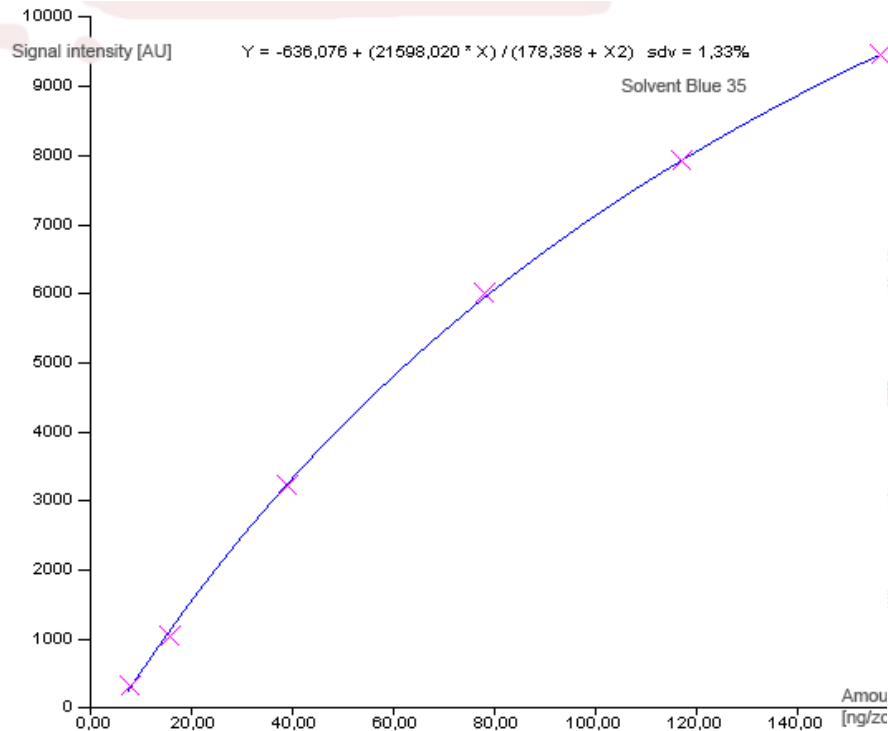


SIM elution profile of ITX @ m/z 255 $[M+H]^+$ and 277 $[M+Na]^+$

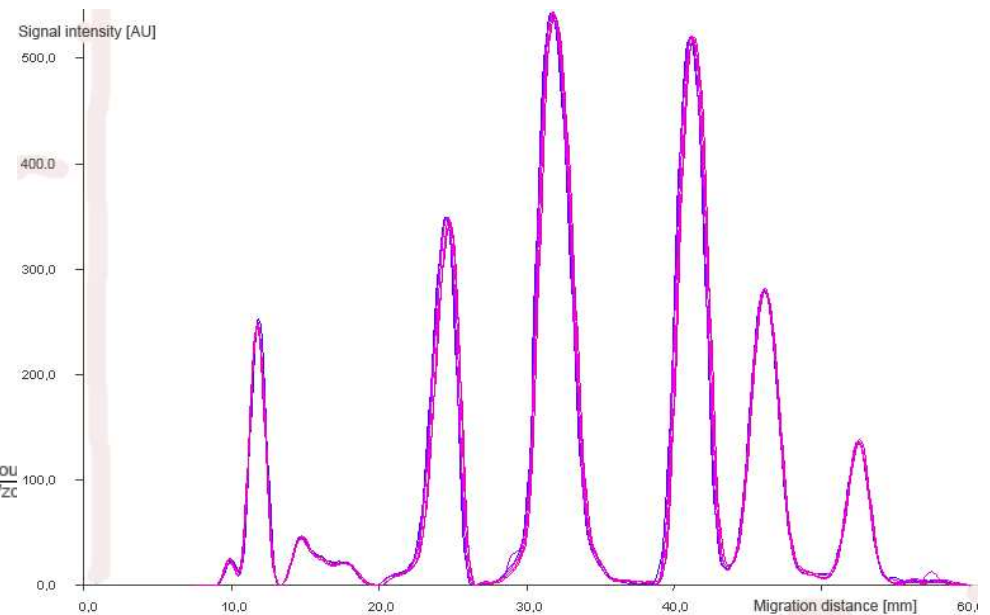
Performance data obtained with the TLC-MS interface

⇒ before: check of performance data by HPTLC-Vis

Calibration for Solvent Blue 35 (%RSD = 1.3%)



Overlay of 5 analog curves (%RSD ≤ 1.3%)

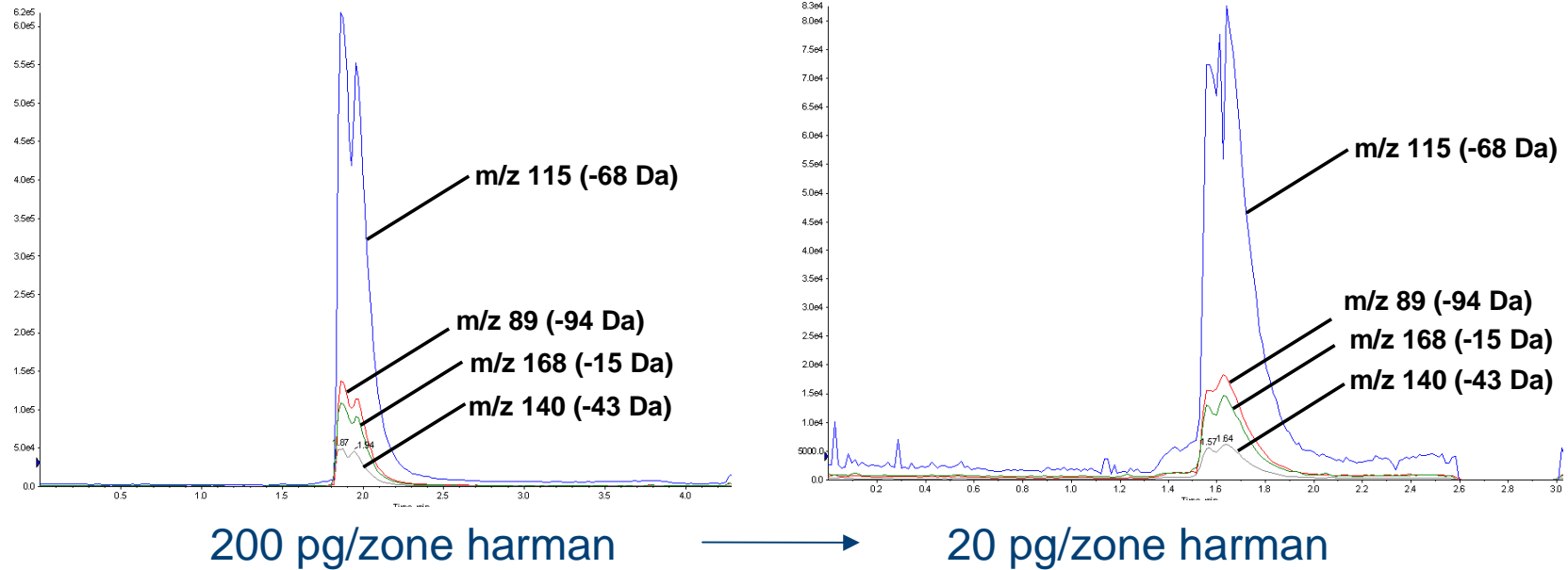




Performance data obtained with the TLC-MS interface

HPTLC-ESI-MS (SIM, peak area)	hR_F - value	Linearity		Precision	
		Calibration range (ng/band)	Determination coefficient	Conc. (ng/band)	n = 5, %RSD
Dimethyl Yellow	65	12 – 234	0.9943	1125	8.1
Oracet Red G	50	2 – 39	0.9950	189	11.0
Solvent Blue 35	41	10 – 52	0.9931	750	4.6
Sudan Red G	27	6 – 117	0.9984	564	8.8
Solvent Blue 22	17	21 – 78	0.9976	750	3.8
Oracet Violet 2R	4	8 – 156	0.9752	1500	11.6

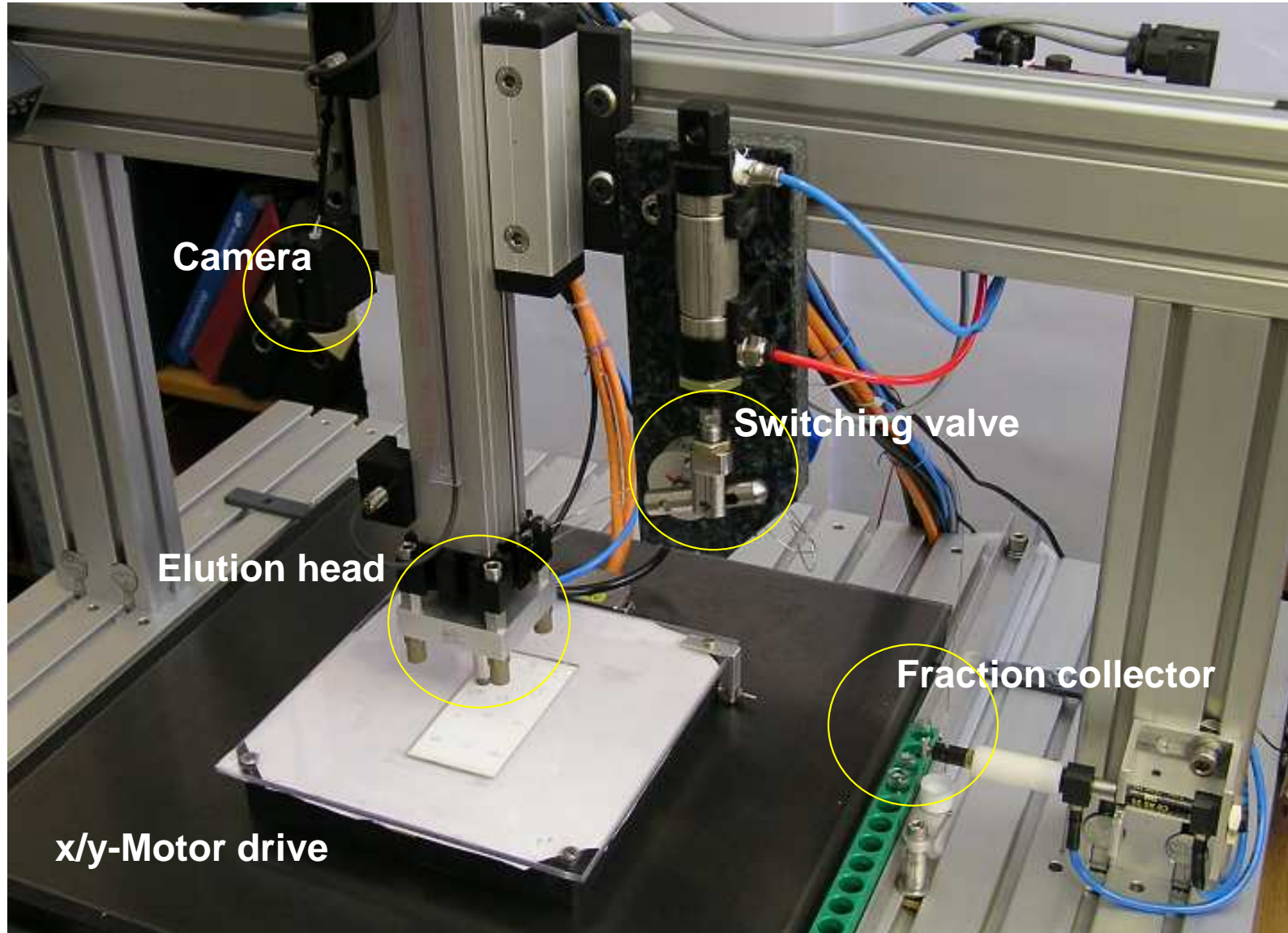
Detectability by HPTLC-ESI-MS/MS



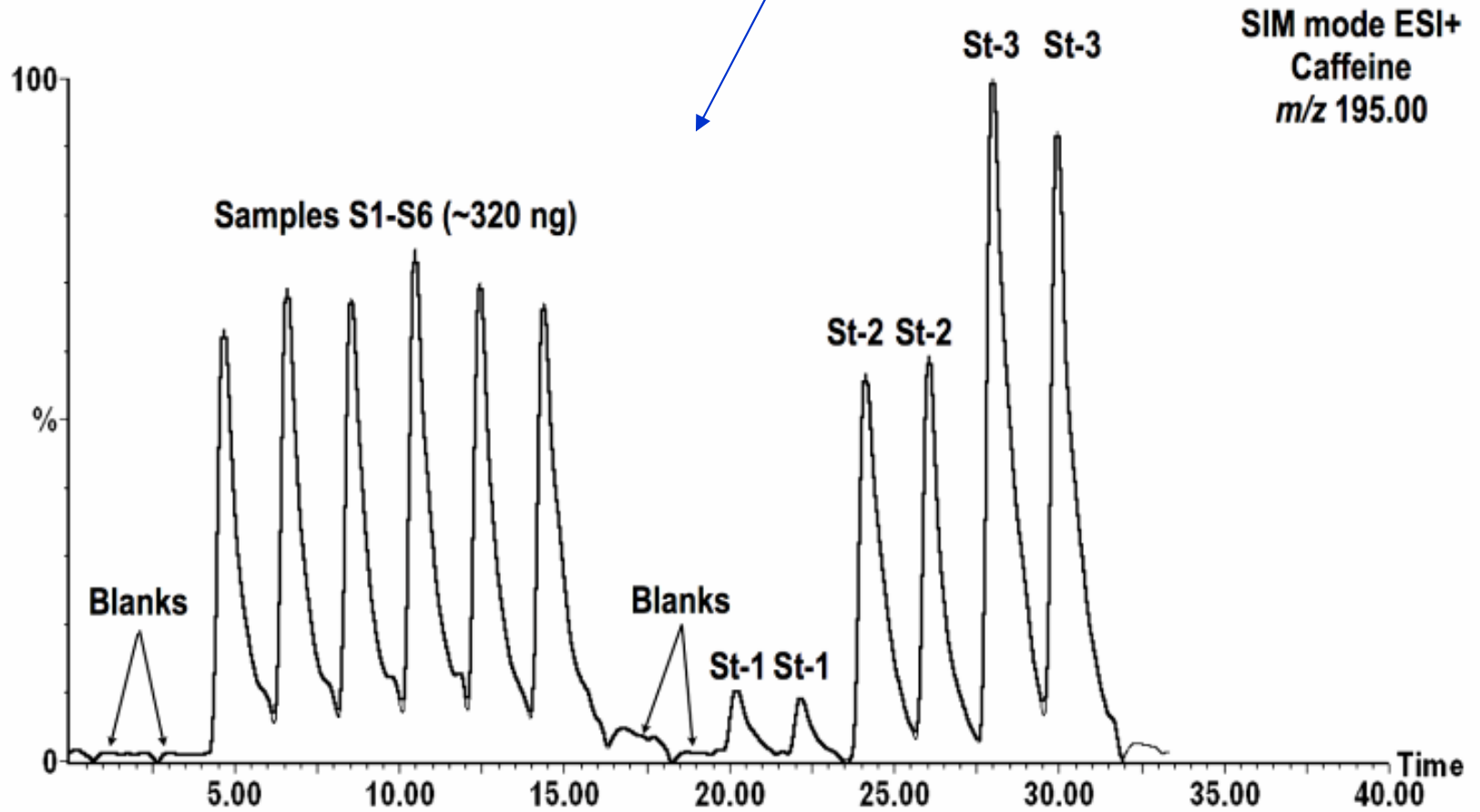
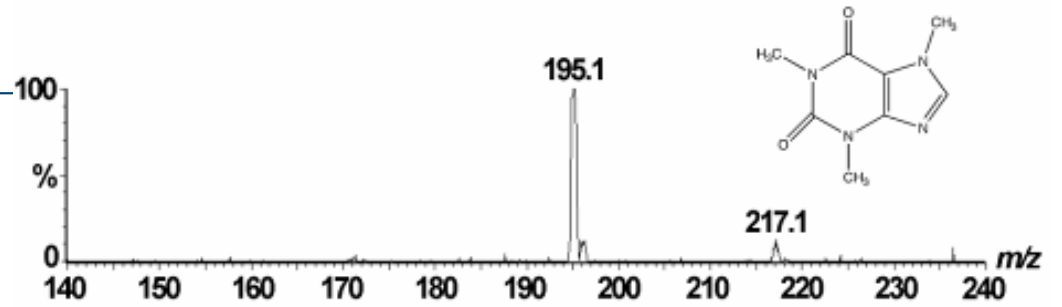
→ LOQ better than 20 pg/zone harman (S/N 20)

→ Detectability comparable to HPLC/MS

Hands-free interface called 'R3D3'



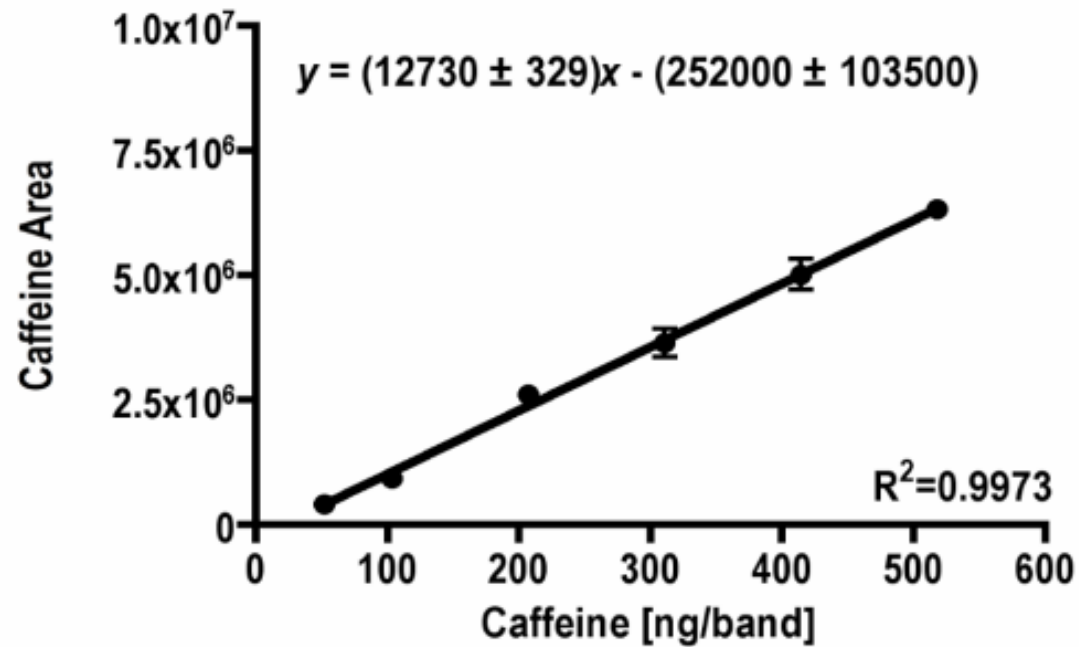
Elution profiles





Data of validation without IS

- Repeatability (%RSD, $n = 6$) in matrix: 5.6 %
- Linearity R^2 : 0.9973





Analysis of samples containing caffeine

Sample	Pharmaceutical mean \pm SD (mg/tablet)	Energy drink mean \pm SD (mg/100 mL)
HPTLC/ESI-MS RSD (% , n = 6)	102.09 \pm 5.76 (5.6)	32.91 \pm 1.60 (4.9)
HPTLC/UV RSD (% , n = 5)	101.98 \pm 2.30 (2.3)	33.71 \pm 0.96 (2.8)
Label	100	32

→ Comparable findings to validated HPTLC/UV methods (F-test, t-test)



Comparison of automated interfaces

Parameter	Precision %RSD	Linear Response r^2
-----------	-------------------	--------------------------

Quantification **without** internal standard

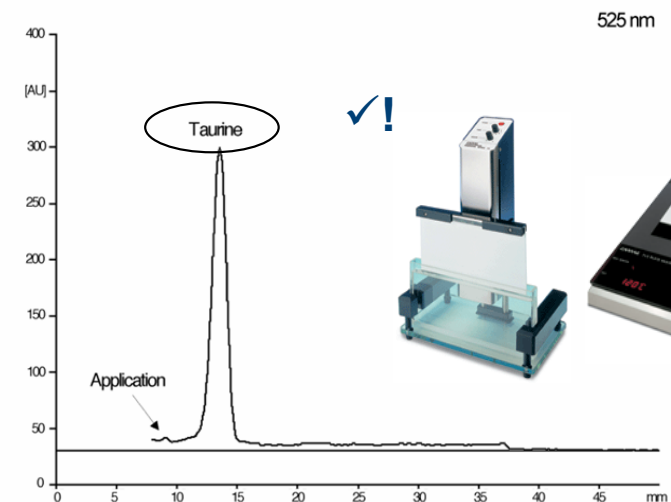
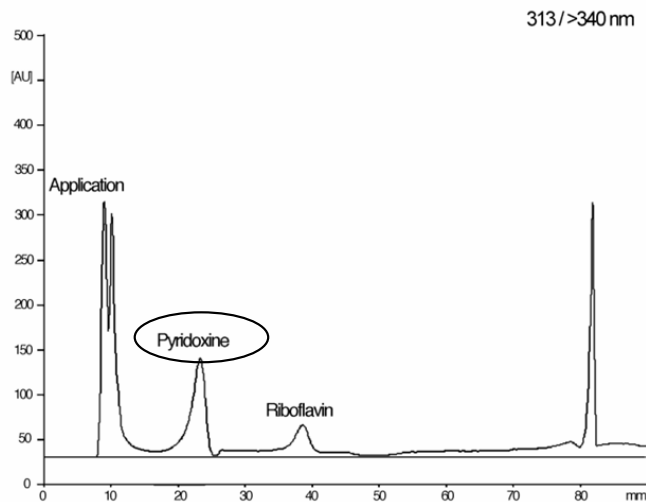
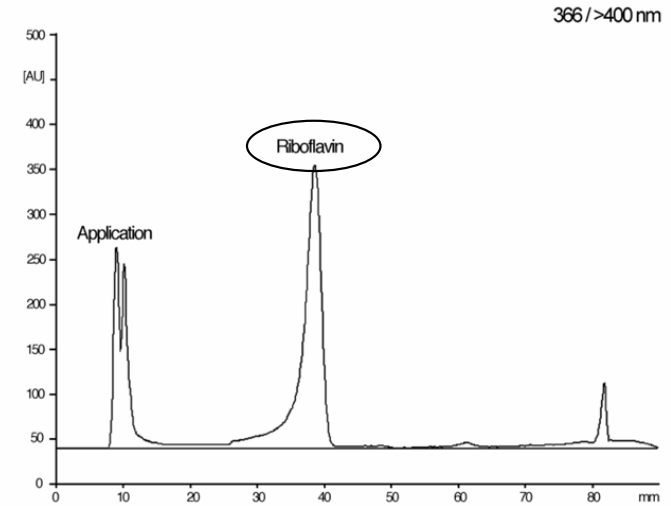
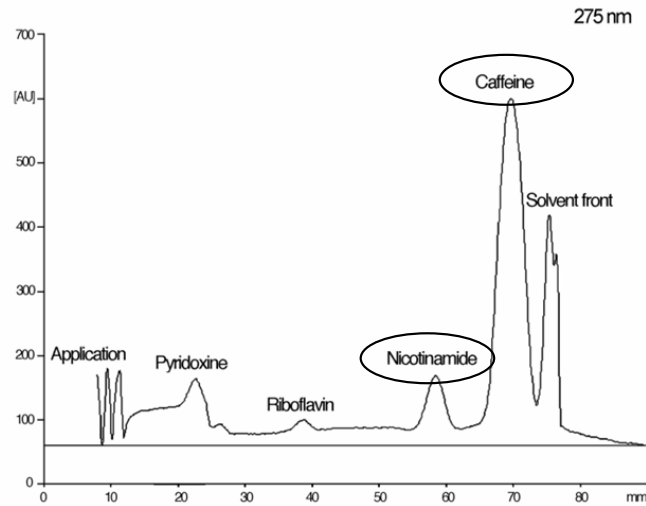
Parameter	Precision %RSD	Linear Response r^2
Elution head (autom.)	$\leq 5.6 \%$	0.9973
DESI	$\leq 16.8 \%$	0.95 - 0.98
MALDI	10 %	-
LA-ICP	17 – 41 %	≥ 0.90

Quantification **with** internal standard

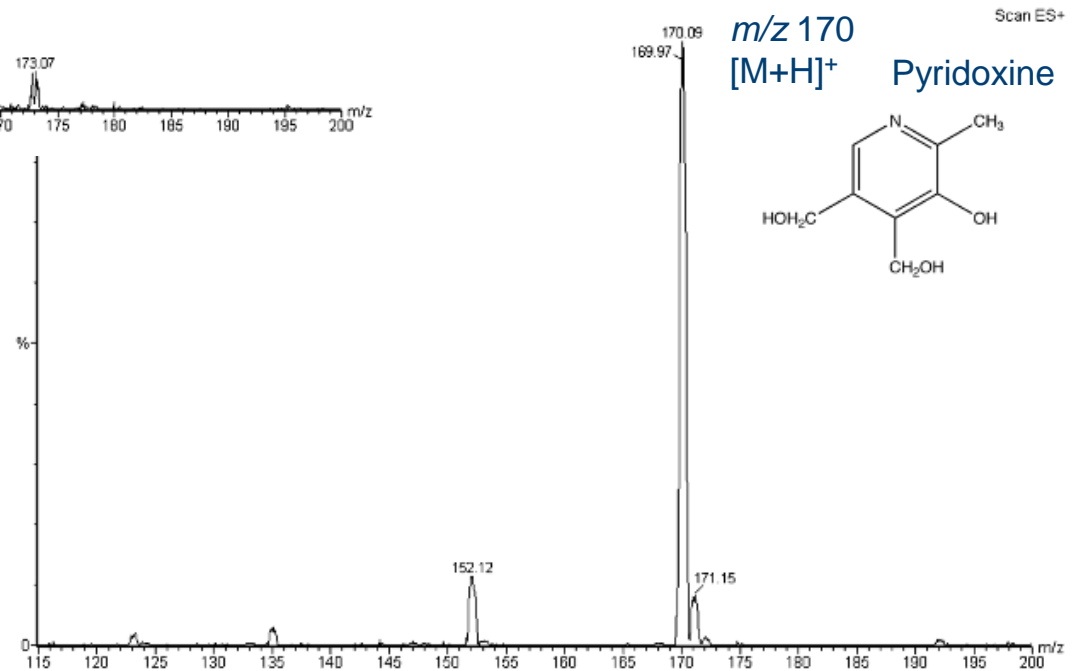
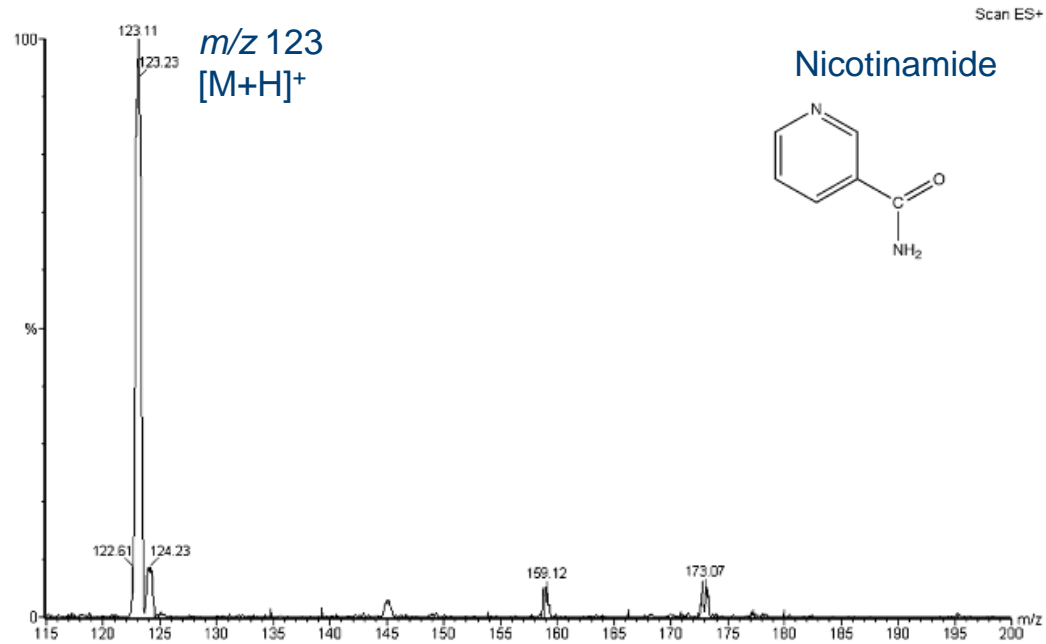
Parameter	Precision %RSD	Linear Response r^2
Micro-junction ESI	$\leq 4.4 \%$	0.9999
SALDI/APCI	7 %	0.9991
MALDI	$\leq 8.9 \%$	0.9969
LA-ICP	3 – 40 %	≥ 0.98

Active ingredients in energy drinks

Simultaneous determination by MWL scan (UV/FLD) → Derivatization → Vis

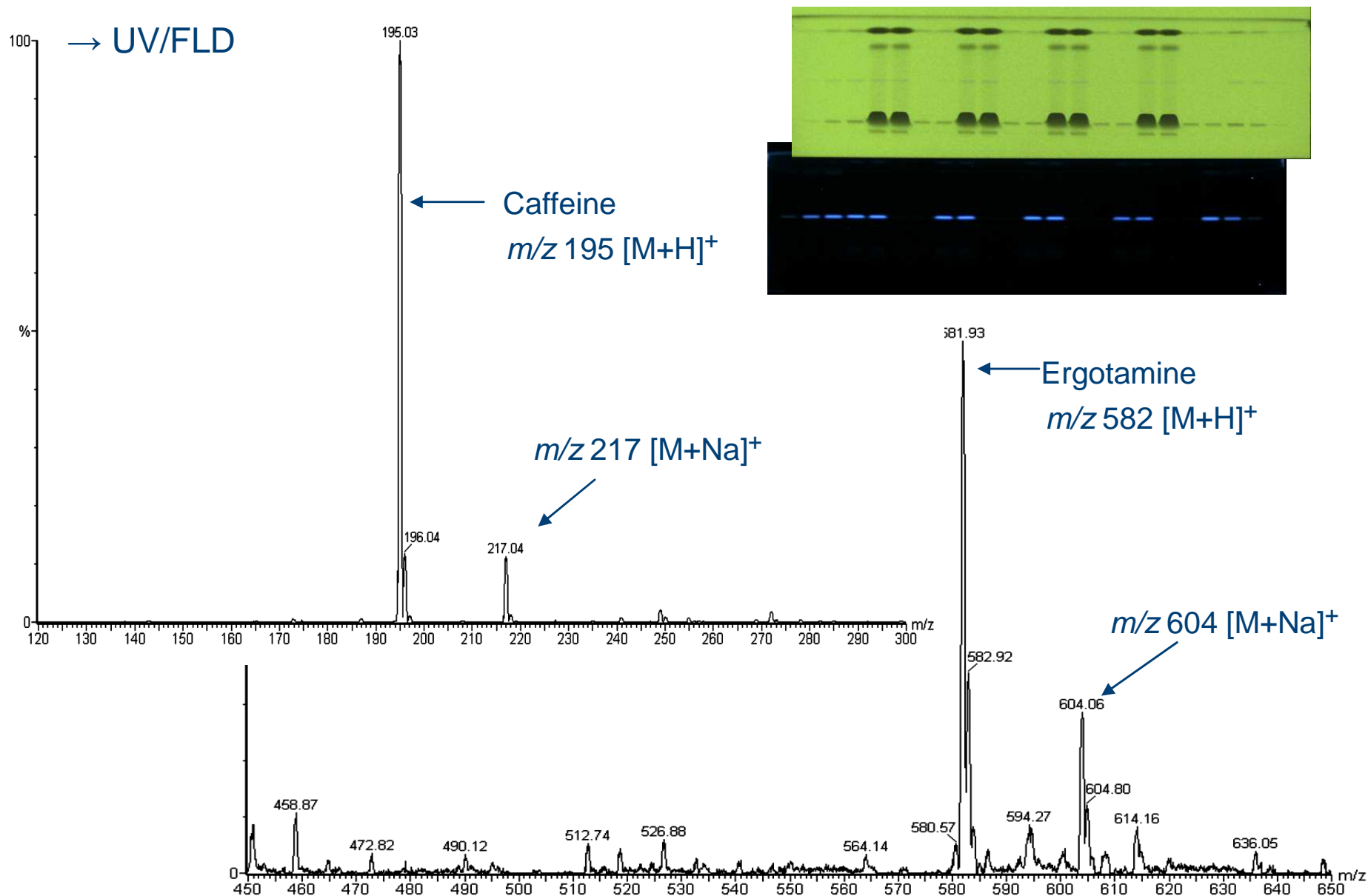


Confirmation by MS

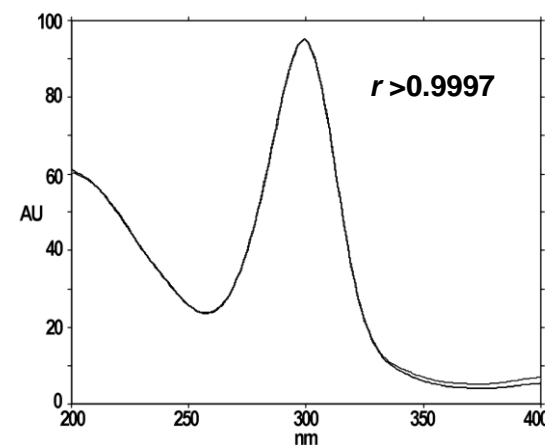
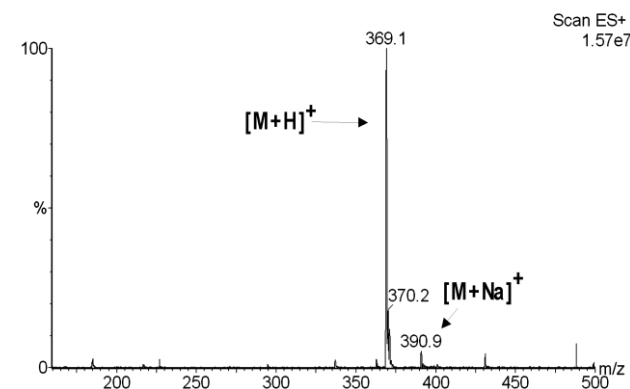
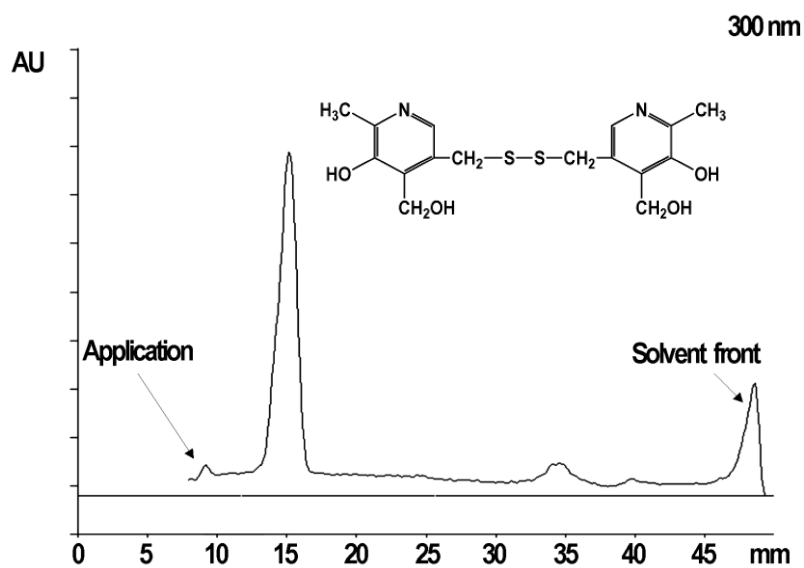
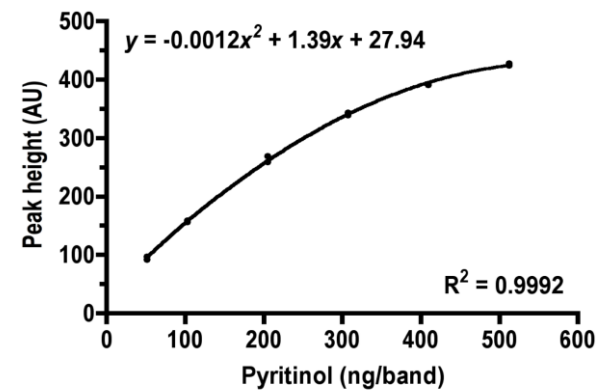
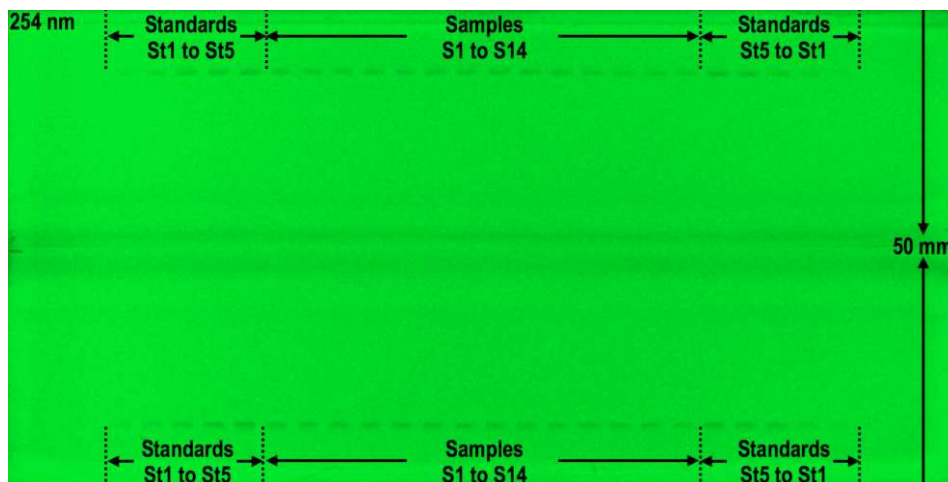




Caffeine, ergotamine and metamizol in tablets

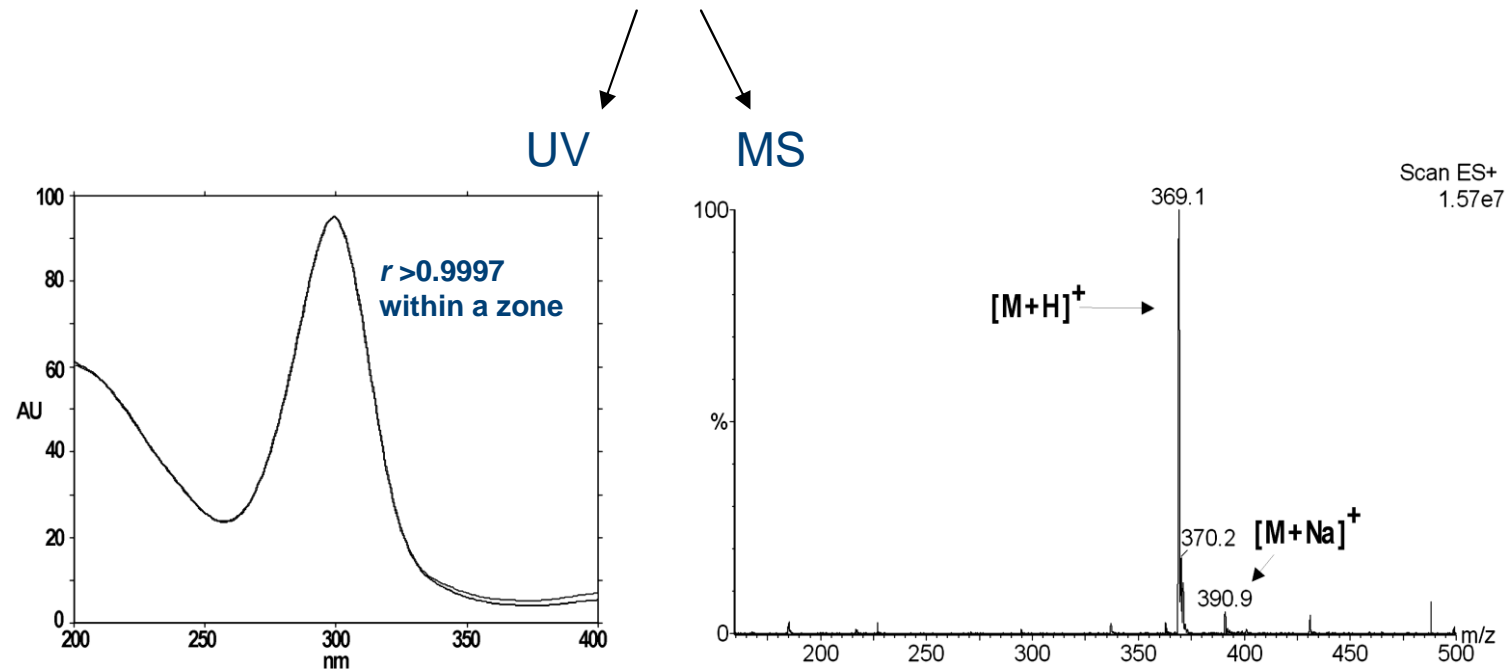


Pyridinol in tablets



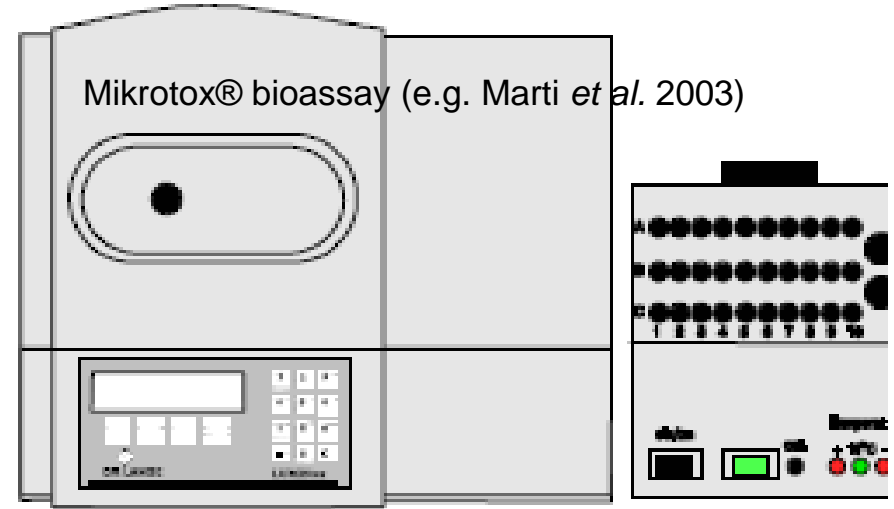
...no need for a higher separation power...

- Repeatability (%RSD, n = 6) in matrix: 0.4 %
- Intermediate precision (%RSD, n = 3) in matrix: 2.95 %
- Recoveries of spiked samples (three levels): 98.5 - 101.9 % (\pm 3.6 - 4.7%)
- LOD/LOQ: 0.6/2.0 $\mu\text{g/mL}$ (6/20 ng/band)
- Up to 17 times less mobile phase consumption
- Up to 8 times faster
- Selectivity proven by spectral purity

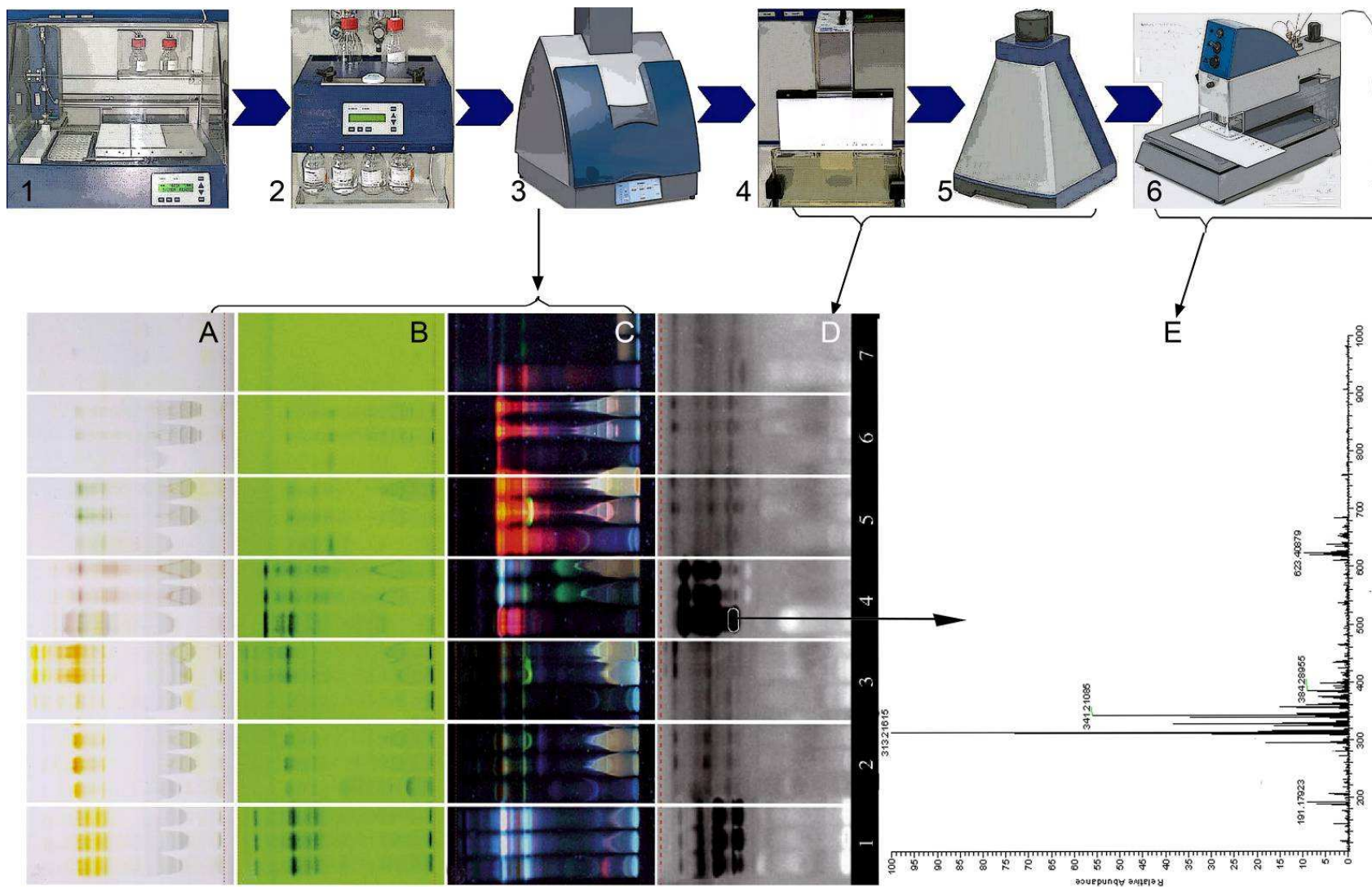


Effect-directed analysis → sum parameter!

DEUTSCHE NORM		Mai 2009
DIN EN ISO 11348-2		
ICS 13.060.70	Ersatz für DIN EN ISO 11348-2:1999-04	
Wasserbeschaffenheit – Bestimmung der Hemmwirkung von Wasserproben auf die Lichtemission von <i>Vibrio fischeri</i> (Leuchtbakterientest) – Teil 2: Verfahren mit flüssig getrockneten Bakterien (ISO 11348-2:2007); Deutsche Fassung EN ISO 11348-2:2008		



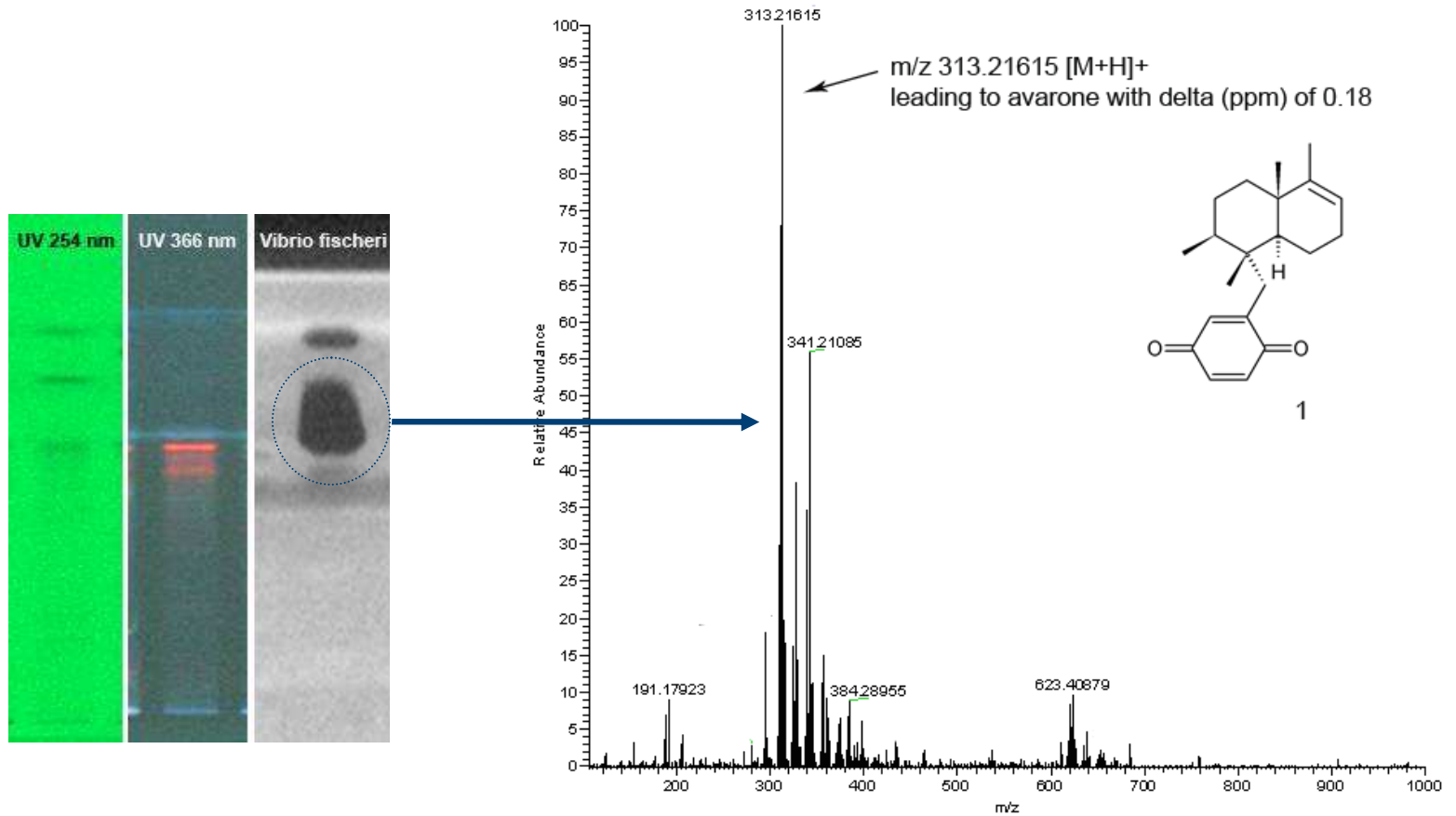
HPTLC-VIS/UV/FLD-EDA-HRMS



G. Morlock, W. Schwack LCGC Eur July (2008) 366-371

A. Klöppel, W. Grasse, F. Brümmer, G. Morlock, J Planar Chromatogr 21 (2008) 431-436

What is it? → HRMS



Case study: Structure elucidation

Correct substance assignment of a dye mixture?





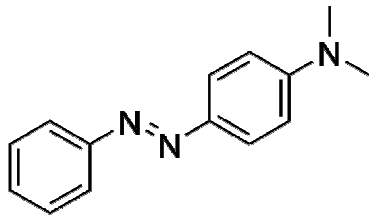
Improved mass assignment

... from a single quadrupole MS → MassWorks software (Cerno)

www.cernobioscience.com



Dye	hR_F -value	Monoisotopic mass measured	Theoretical monoisotopic mass of the proposed ion	Δ (ppm)	Spectral accuracy	Double bond equivalent	Protonated molecular formula	Assigned to
Dimethyl Yellow ✓	65	226.1402	226.1344	-25,5482	98.5341	8.5	$C_{14}H_{16}N_3$	$[M+H]^+$



Dimethyl Yellow



Tasks



Calibration



Analysis

Recent

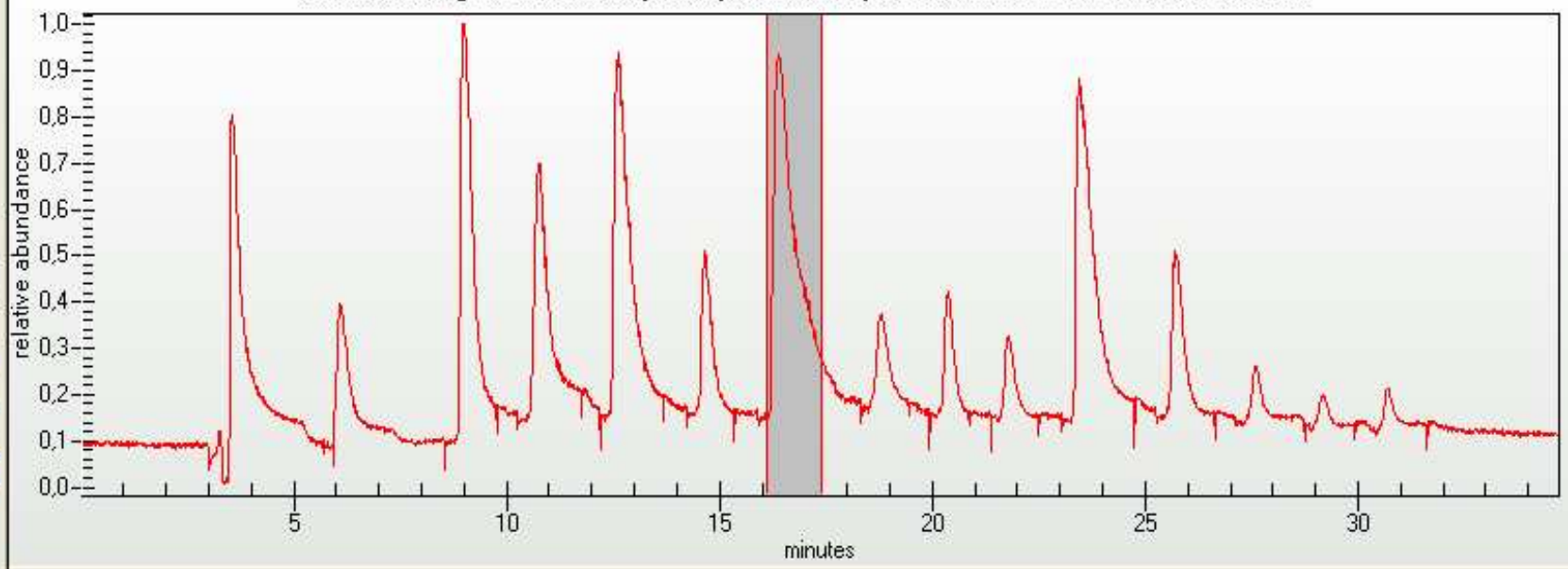


RT Window List

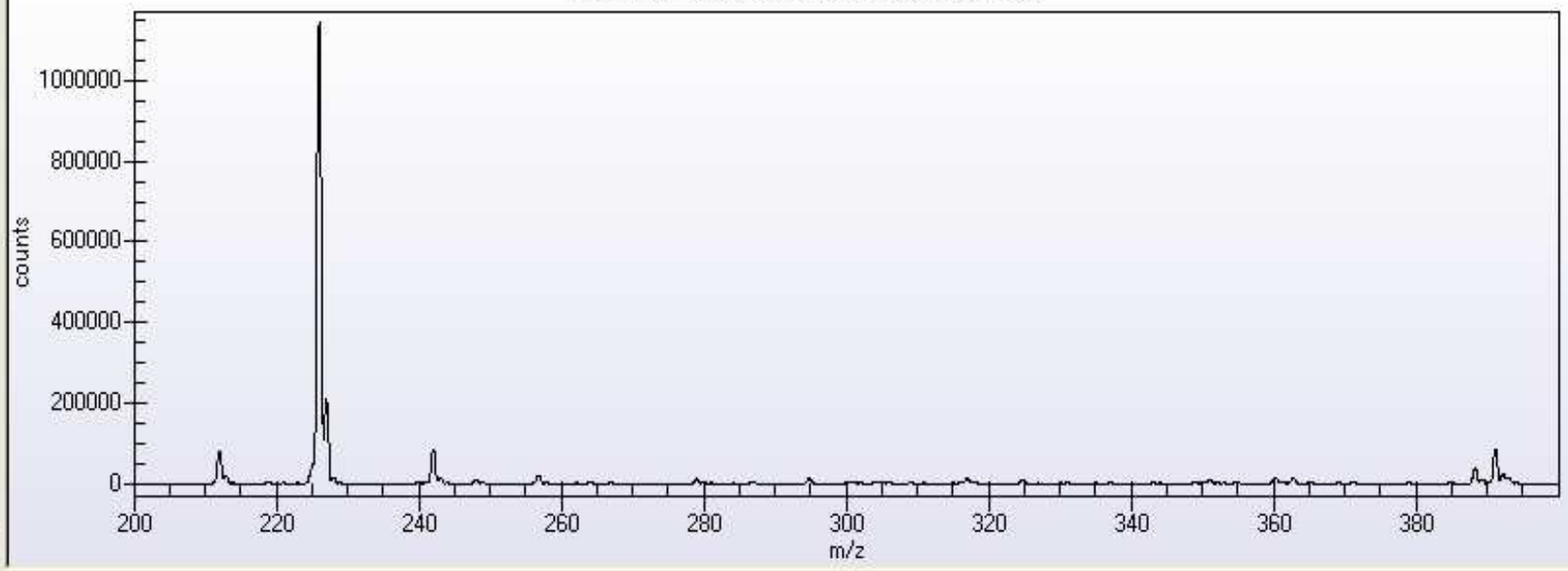
O..	Scan	Time	Name
<input type="checkbox"/>	12...	16.1...	Scans 1...

✓ Instrument Data ✗ Standard Ions ✗ Calibrate ✗ Review

I:\Veröffentlichung\2010\Neil Brett\Dyes study MS\Masses by Cerno\Cerno GM\130510000001.D\MSD1.MS



Average of Scans 1203 thru 1299 (16.13 to 17.42)





Tasks



Calibration



Analysis

Recent

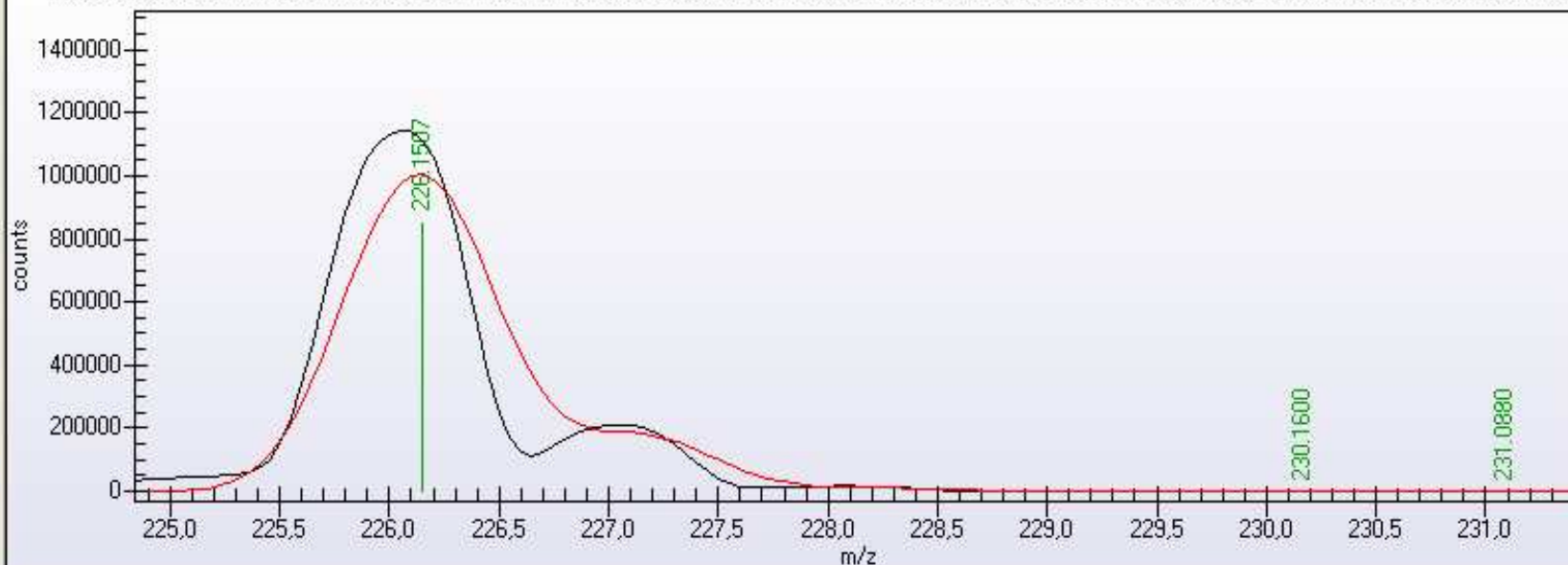


RT Window List

O..	Scan	Time	Name
<input type="checkbox"/>	12...	16.1...	Scans 1...

Instrument Data
 Standard Ions
 Calibrate
 Review

Average of Scans 1203 thru 1299 (16.13 to 17.42) - I:\Veröffentlichung\2010\Neil Brett\Dyes study MS\Masses by Cerno\Cerno GM\130510000001.D\MSD1.MS



	RT Window	Name	Net Formula	Mono Mass	Closest Centroid	ΔM (mDa)	ΔM (ppm)	Spectral Accuracy	RMSE
1	0: Scans 1203-1299		C ₁₄ H ₁₆ N ₃	226,1344	226,1507	16,3500	72,3	99,1	3,107,36



Tasks

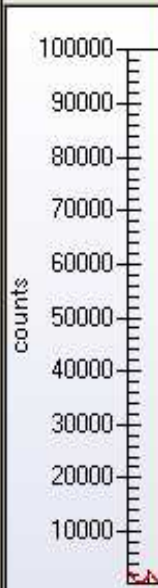
Calibration

Analysis

Recent

RT Window List

O.	Scan	Time	Name
<input type="checkbox"/>	4...	5.9...	Scans...



CLIPS Search

Accurate mass to search: Show All Results

Charge: Show Top Results

Formula Generation by Mass

Mass Tolerance

mDa

PPM

Electron State

Odd Even Both

Double Bond Equivalent Range

Min Max

	Element	Min	Max
1	C	0	20
2	H	0	100
3	O	0	10
4	N	0	10
5			

Formula Determination by Spectral Accuracy

Profile Mass Range (Da)

Start End

Mixture Search

Formula 2:

Formula 3:

Interference Rejection:

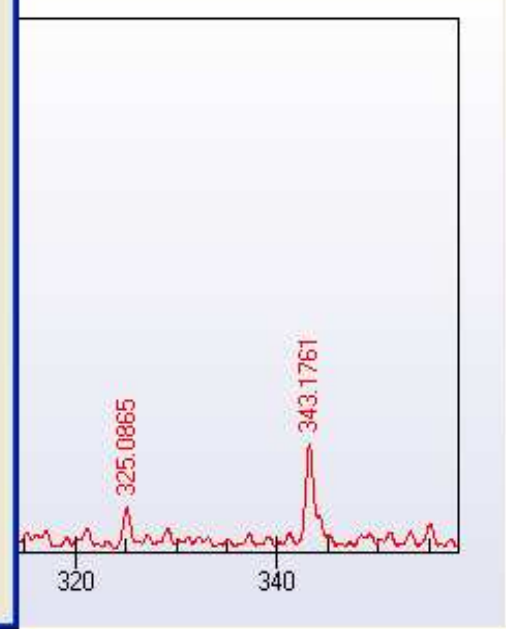
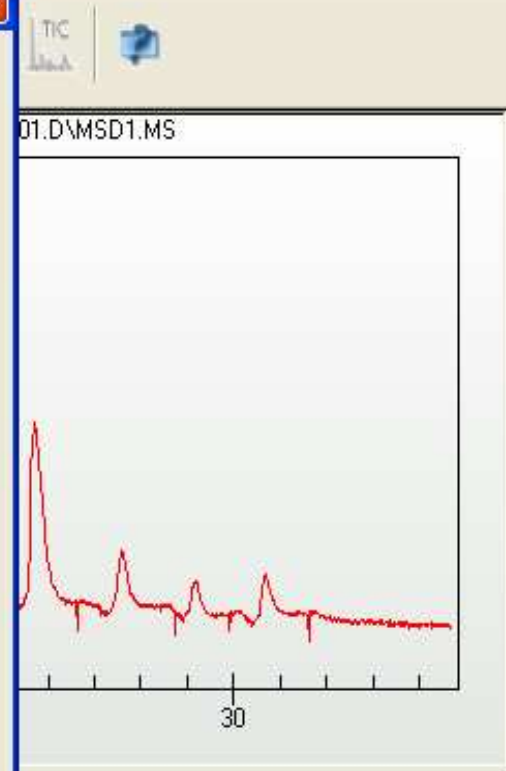
Show this dialog before each search

Reset to Factory Defaults

Reset to My Defaults

Save As My Defaults

Search Cancel





Tasks



Calibration



Analysis

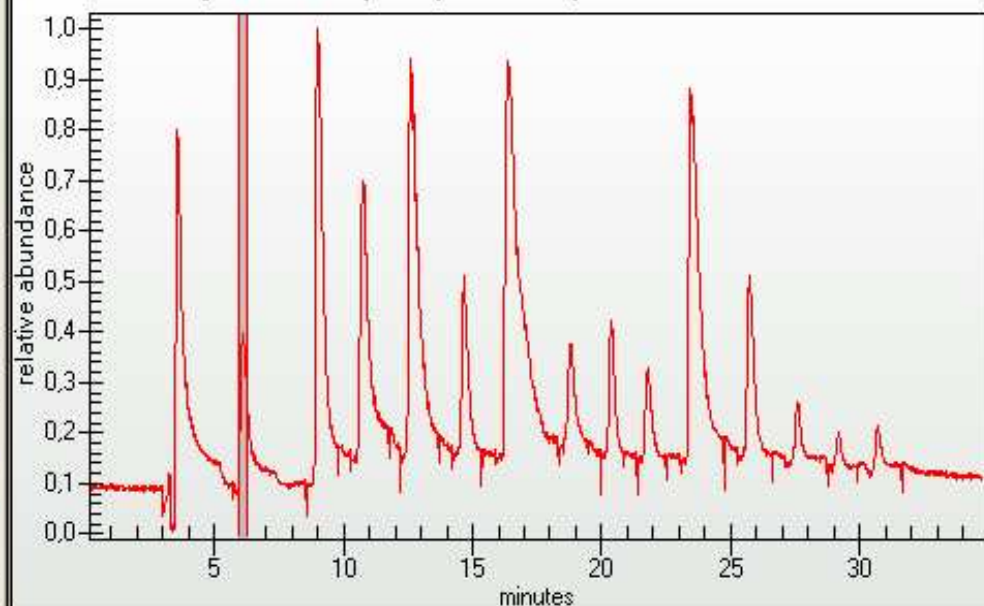
Recent



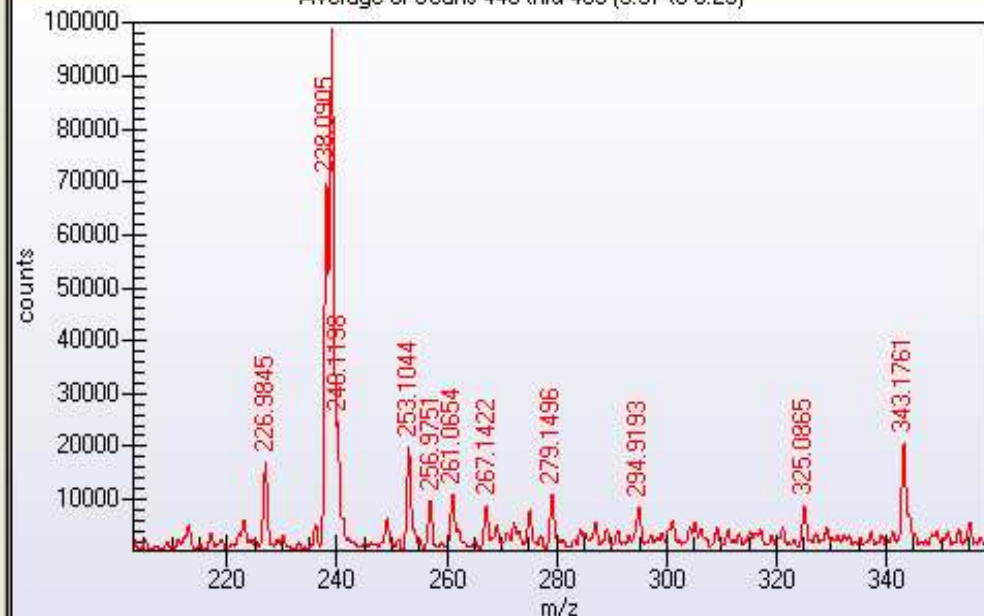
CLIPS Results

	Formula	Mono Isotope	Mass Error (mDa)	Mass Error (PPM)	Spectral Accuracy	RMSE	DBE
1	C15H12O2N	238,0868	-3,6964	-15,5250	45,5841	25.459	10,5
2	C14H12ON3	238,0980	7,5370	31,6561	45,7740	25.370	10,5
3	C10H12O4N3	238,0828	-7,7191	-32,4209	43,8702	26.261	6,5
4	C9H12O3N5	238,0940	3,5143	14,7603	44,1377	26.136	6,5
5	C8H16O7N	238,0927	2,1769	9,1431	43,3133	26.521	1,5
6	C3H16O9N3	238,0887	-1,8459	-7,7528	41,6101	27.318	-2,5
7	C2H16O8N5	238,0999	9,3875	39,4284	41,7678	27.245	-2,5
8	CH20O12N	238,0985	8,0501	33,8111	41,0559	27.578	-7,5

I:\Veröffentlichung\2010\Neil Brett\Dyes study MS\Masses by Cerno\Cerno GM130510000001.D\MSD1.MS



Average of Scans 446 thru 465 (5.97 to 6.23)





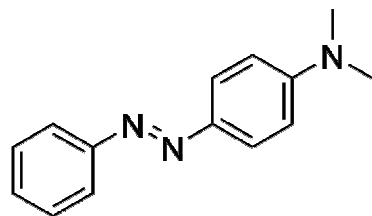
Improved mass assignment

... from a single quadrupole MS → MassWorks software (Cerno)

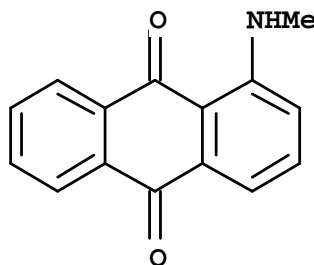
www.cernobioscience.com



Dye	hR_F -value	Monoisotopic mass measured	Theoretical monoisotopic mass of the proposed ion	Δ (ppm)	Spectral accuracy	Double bond equivalent	Protonated molecular formula	Assigned to
Dimethyl Yellow ✓	65	226.1402	226.1344	-25,5482	98.5341	8.5	$C_{14}H_{16}N_3$	$[M+H]^+$
Oracet Red G ✓	50	238.0893	238.0868	-10.4850	91.1892	10.5	$C_{15}H_{12}NO_2$	$[M+H]^+$



Dimethyl Yellow



Oracet Red G



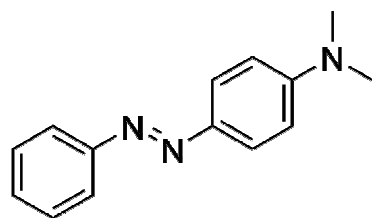
Improved mass assignment

... from a single quadrupole MS → MassWorks software (Cerno)

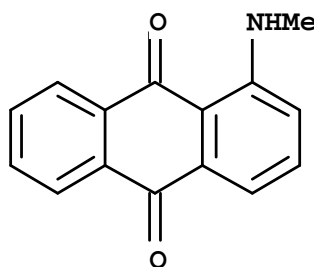
www.cernobioscience.com



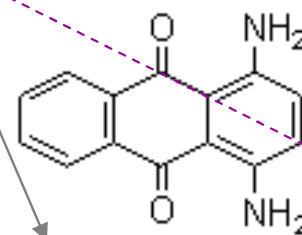
Dye	hR_F -value	Monoisotopic mass measured	Theoretical monoisotopic mass of the proposed ion	Δ (ppm)	Spectral accuracy	Double bond equivalent	Protonated molecular formula	Assigned to
Dimethyl Yellow ✓	65	226.1402	226.1344	-25,5482	98.5341	8.5	$C_{14}H_{16}N_3$	$[M+H]^+$
Oracet Red G ✓	50	238.0893	238.0868	-10.4850	91.1892	10.5	$C_{15}H_{12}NO_2$	$[M+H]^+$
Oracet Violet 2R ?	17	305.1610	305.1654	14.3803	98.0042	11.5	$C_{20}H_{21}N_2O$	$[M+H]^+$
Indophenol ?	4	239.0839	239.0821	7.7270	85.4156	10.5	$C_{14}H_{11}N_2O_2$	$[M+H]^+$



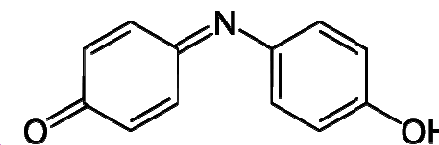
Dimethyl Yellow



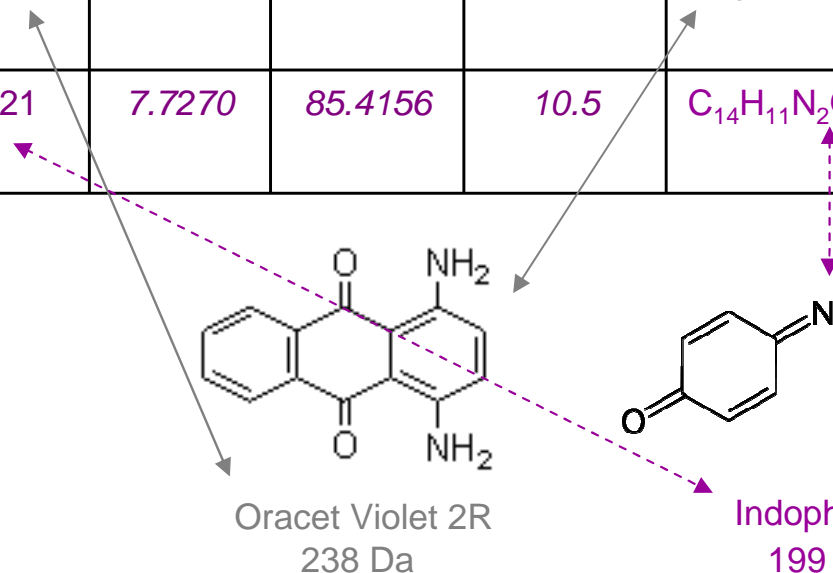
Oracet Red G



Oracet Violet 2R
238 Da



Indophenol
199 Da



Request to supplier for single compounds



→ The elution order was wrongly assigned.

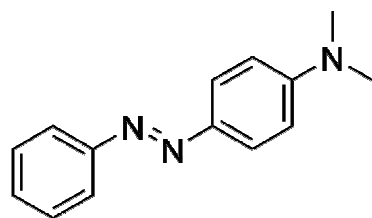


Improved mass assignment

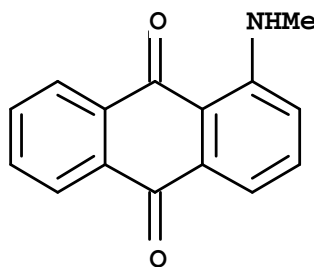
... from a single quadrupole MS → MassWorks software (Cerno)



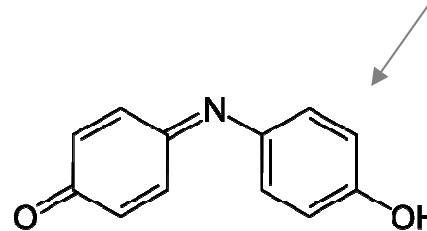
Dye	hR_F -value	Monoisotopic mass measured	Theoretical monoisotopic mass of the proposed ion	Δ (ppm)	Spectral accuracy	Double bond equivalent	Protonated molecular formula	Assigned to
Dimethyl Yellow ✓	65	226.1402	226.1344	-25,5482	98.5341	8.5	$C_{14}H_{16}N_3$	$[M+H]^+$
Oracet Red G ✓	50	238.0893	238.0868	-10.4850	91.1892	10.5	$C_{15}H_{12}NO_2$	$[M+H]^+$
Indophenol ?	17	305.1610	305.1654	14.3803	98.0042	11.5	$C_{20}H_{21}N_2O$	$[M+H]^+$
Oracet Violet 2R ✓	4	239.0839	239.0821	7.7270	85.4156	10.5	$C_{14}H_{11}N_2O_2$	$[M+H]^+$



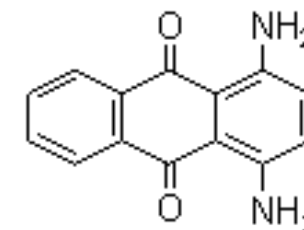
Dimethyl Yellow



Oracet Red G



Indophenol
199 Da



Oracet Violet 2R

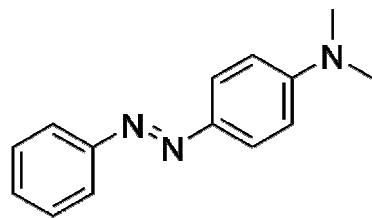


Improved mass assignment

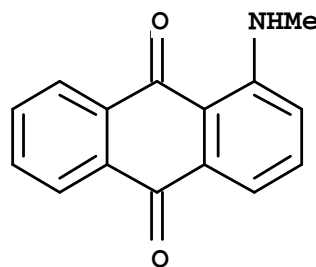
... from a single quadrupole MS → MassWorks software (Cerno)



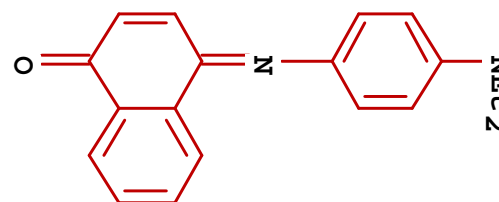
Dye	hR_F -value	Monoisotopic mass measured	Theoretical monoisotopic mass of the proposed ion	Δ (ppm)	Spectral accuracy	Double bond equivalent	Protonated molecular formula	Assigned to
Dimethyl Yellow ✓	65	226.1402	226.1344	-25,5482	98.5341	8.5	$C_{14}H_{16}N_3$	$[M+H]^+$
Oracet Red G ✓	50	238.0893	238.0868	-10.4850	91.1892	10.5	$C_{15}H_{12}NO_2$	$[M+H]^+$
Solvent Blue 22 ✓	17	305.1610	305.1654	14.3803	98.0042	11.5	$C_{20}H_{21}N_2O$	$[M+H]^+$
Oracet Violet 2R ✓	4	239.0839	239.0821	7.7270	85.4156	10.5	$C_{14}H_{11}N_2O_2$	$[M+H]^+$



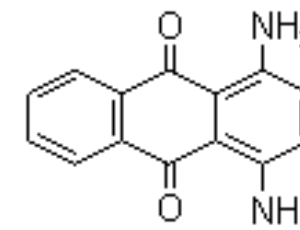
Dimethyl Yellow



Oracet Red G



Chinoneanil



Oracet Violet 2R

Case study: Structure elucidation

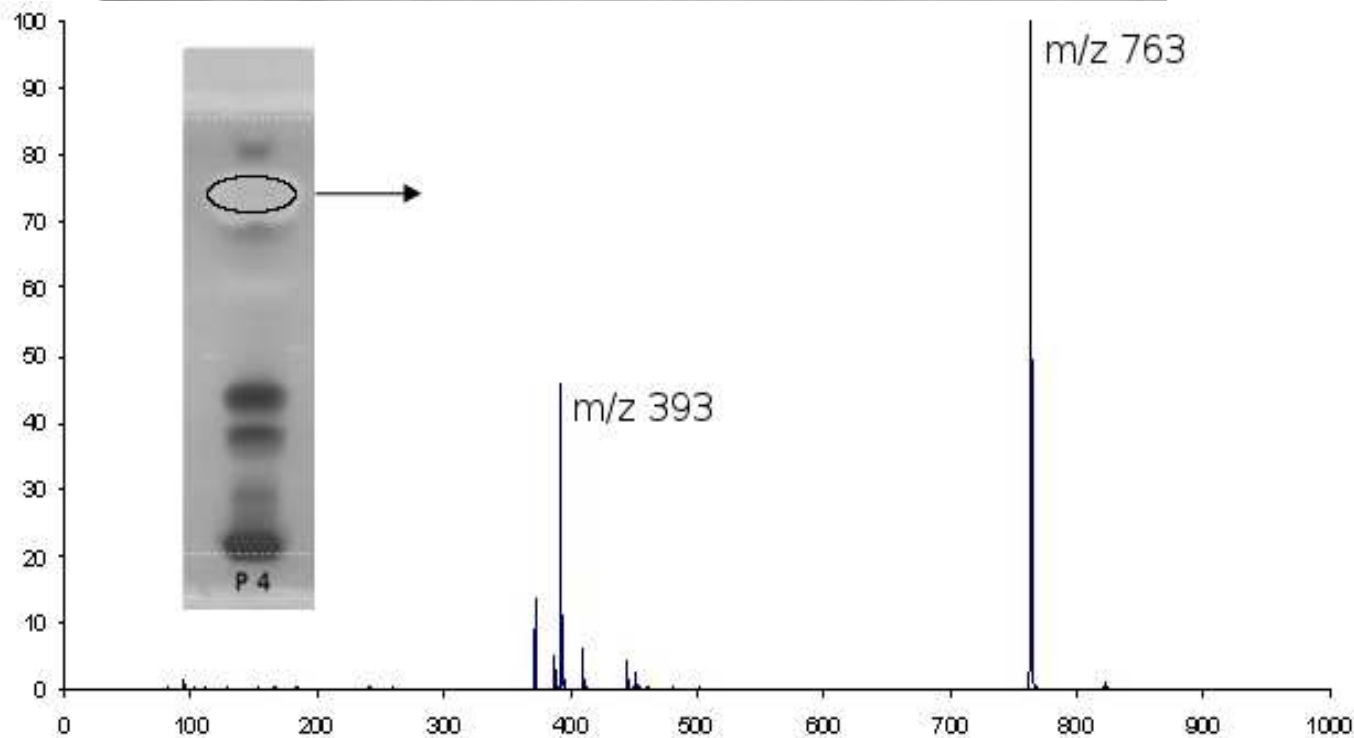
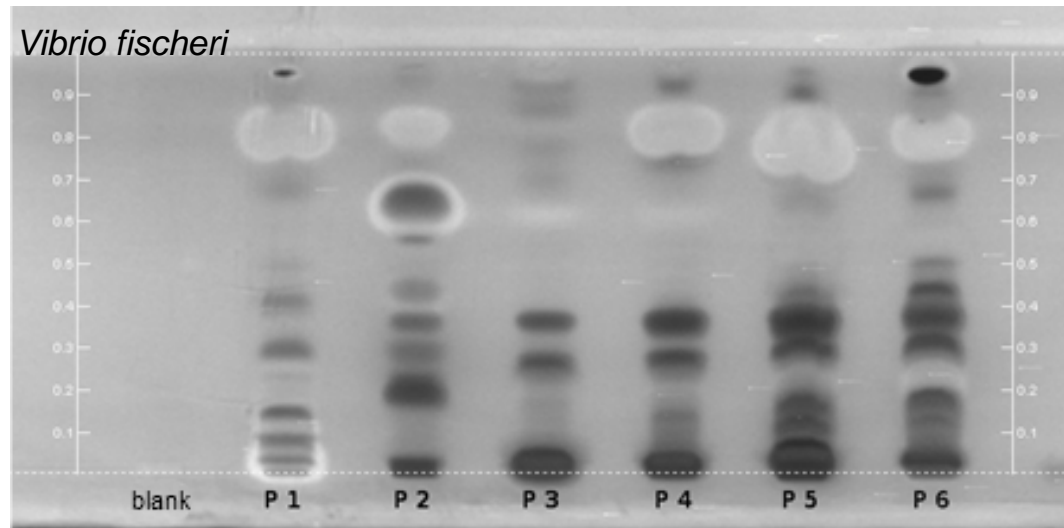
Correct substance assignment of a dye mixture?



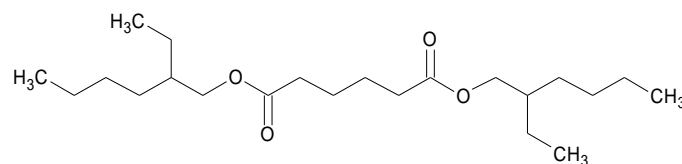
Just partly:

1. Ariabel red is Sudan red G.
2. The elution order was incorrectly assigned.
3. One compound was not the one intended to be: The quinoneanil *Solvent Blue 22* was incorrectly labeled as *indophenol*.
4. Two manufacturers with incorrect label.

Additives in food packaging foils

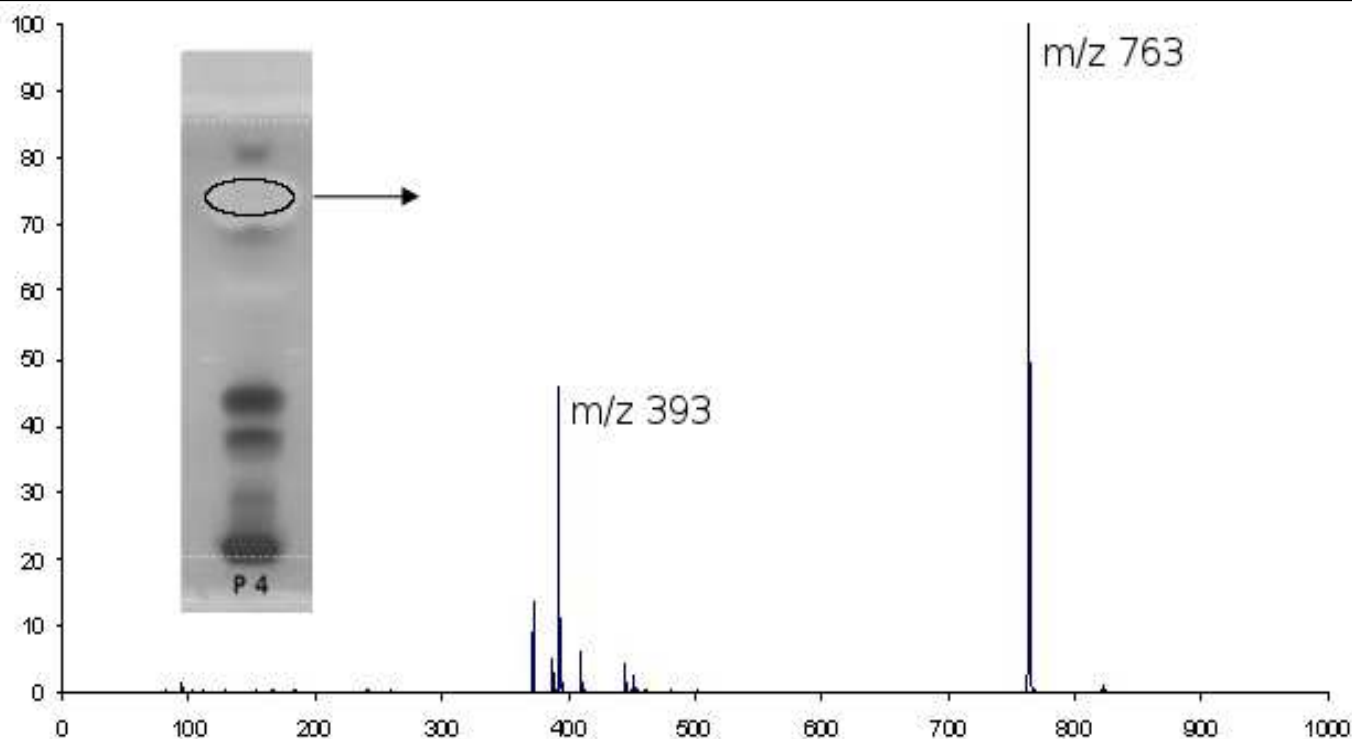


Detection of additives in polymer packaging foils

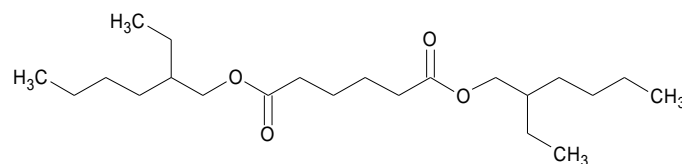


Bis-2-ethylhexyladipate

MS signals of	Mass determined	Mass theoretical	Δ (ppm)	Sum formula	Assignment
HPTLC zone	393,2985	393,2981	-1,0691	$C_{22}H_{42}O_4Na$	$[M+Na]^+$
	763,6077	763,6064	-1,7164	$C_{44}H_{84}O_8Na$	$[2M+Na]^+$

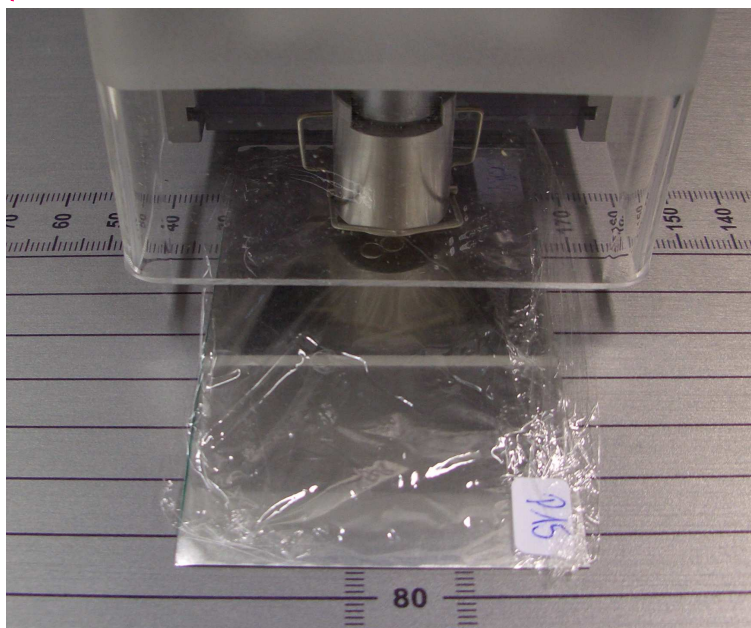


Detection of additives in polymer packaging foils



Bis-2-ethylhexyladipate

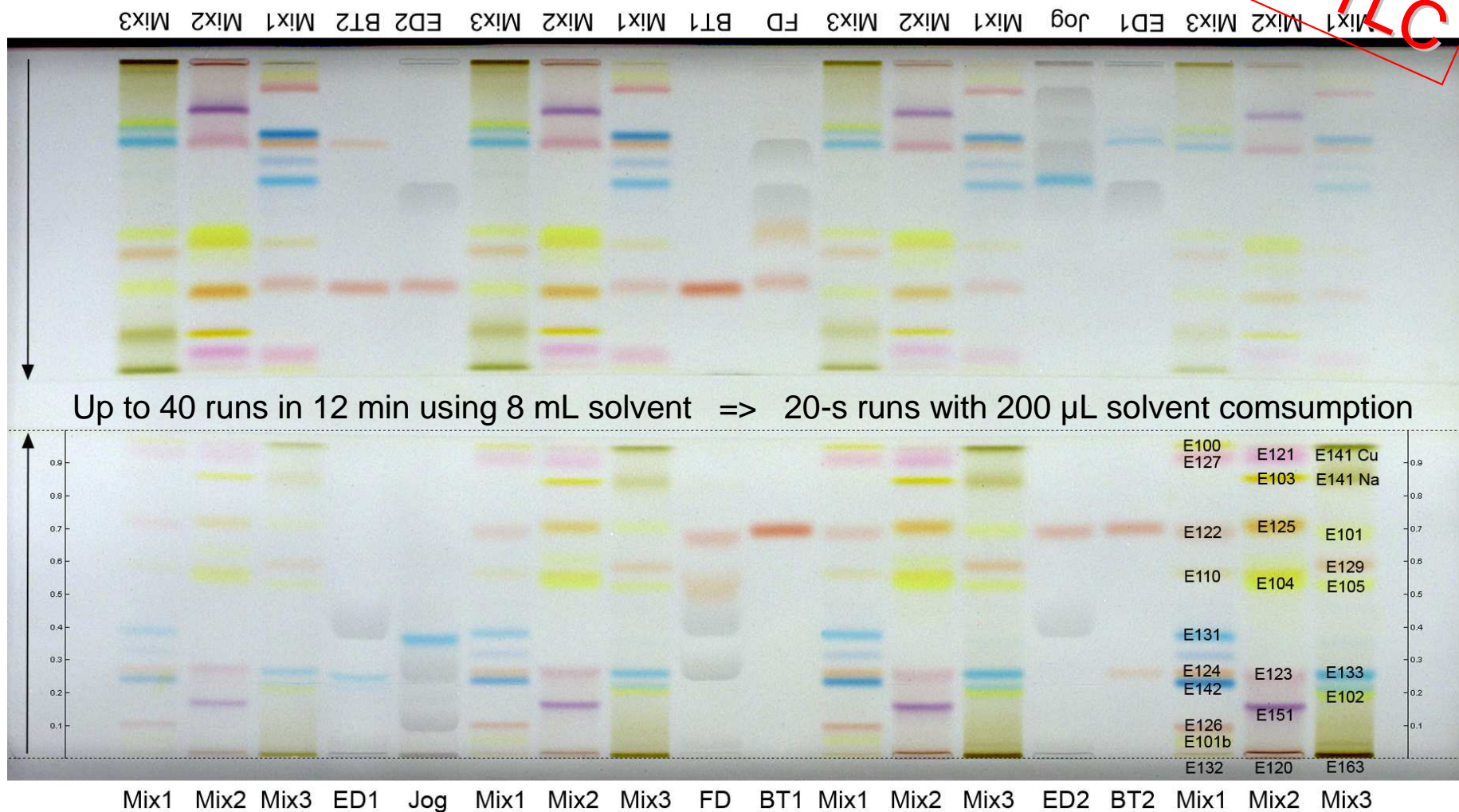
MS signal of	Mass determined	Mass theoretical	Δ (ppm)	Sum formula	Assignment
Plastic foil	371,3174	371,3161	-3,4071	C₂₂H₄₃O₄	[M+H]⁺
HPTLC zone	393,2985	393,2981	-1,0691	C ₂₂ H ₄₂ O ₄ Na	[M+Na] ⁺
	763,6077	763,6064	-1,7164	C ₄₄ H ₈₄ O ₈ Na	[2M+Na] ⁺





Dye analysis

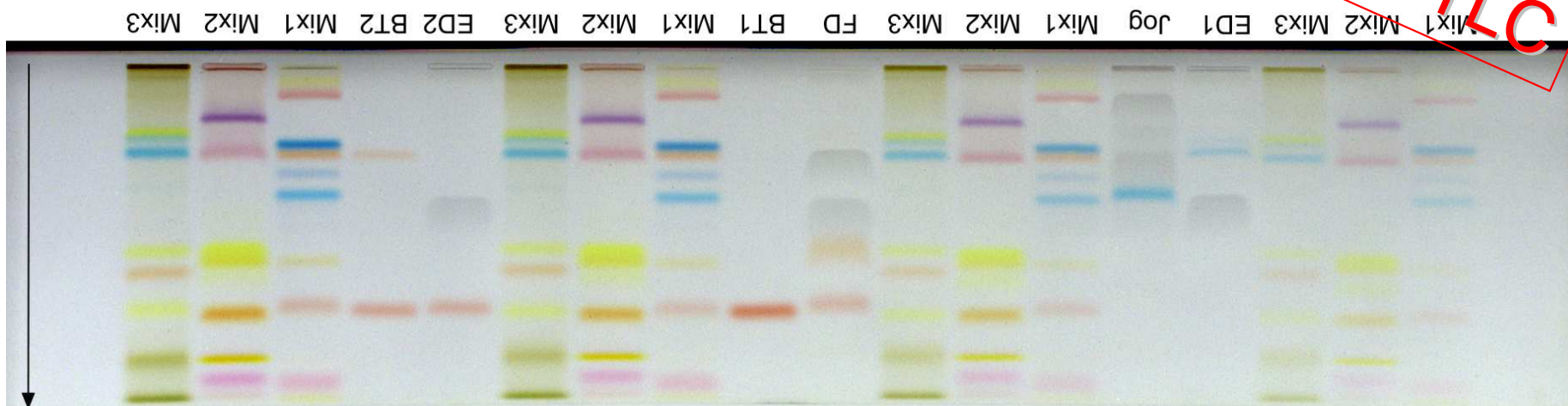
HPTLC



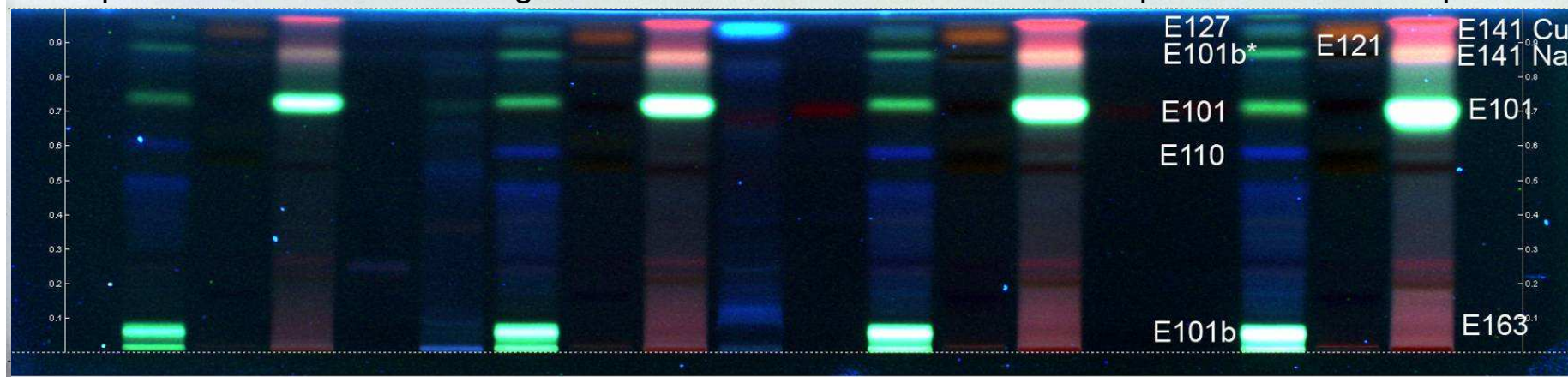


Dye analysis

HPTLC

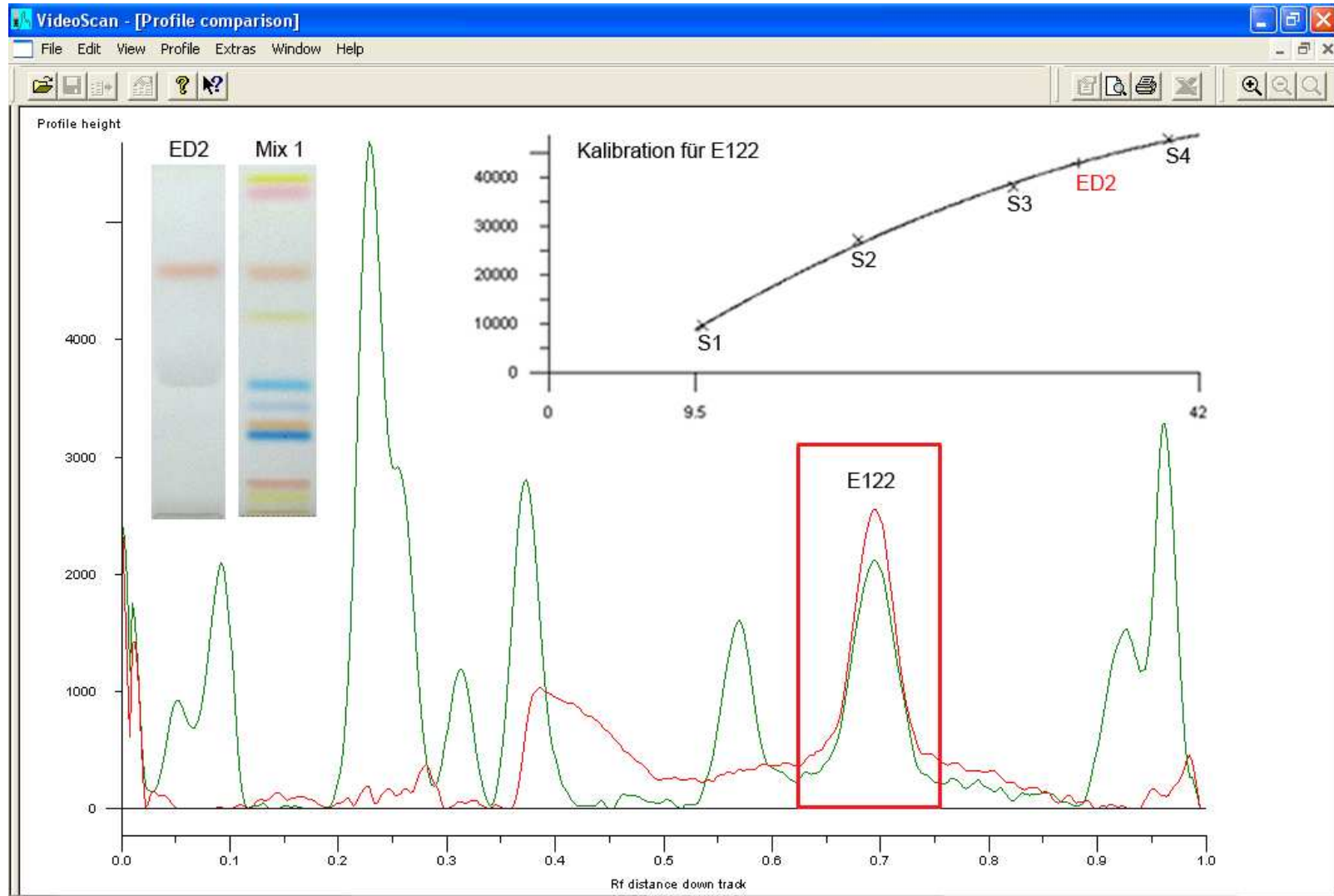


Up to 40 runs in 12 min using 8 mL solvent => 20-s runs with 200 µL solvent consumption

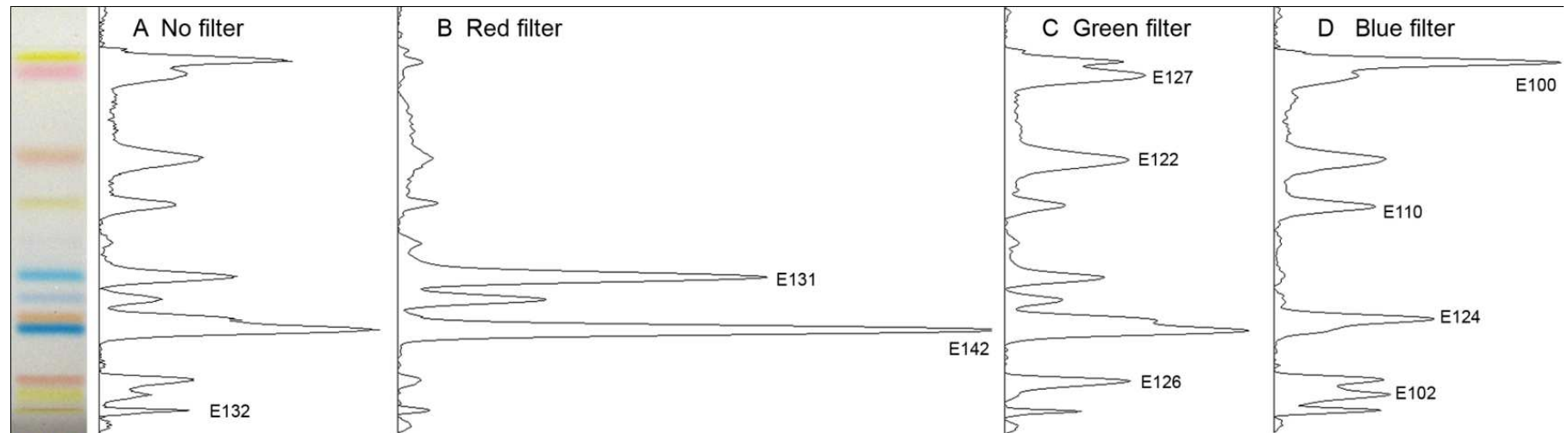


Mix1 Mix2 Mix3 ED1 Jog Mix1 Mix2 Mix3 FD BT1 Mix1 Mix2 Mix3 ED2 BT2 Mix1 Mix2 Mix3

Digital quantification

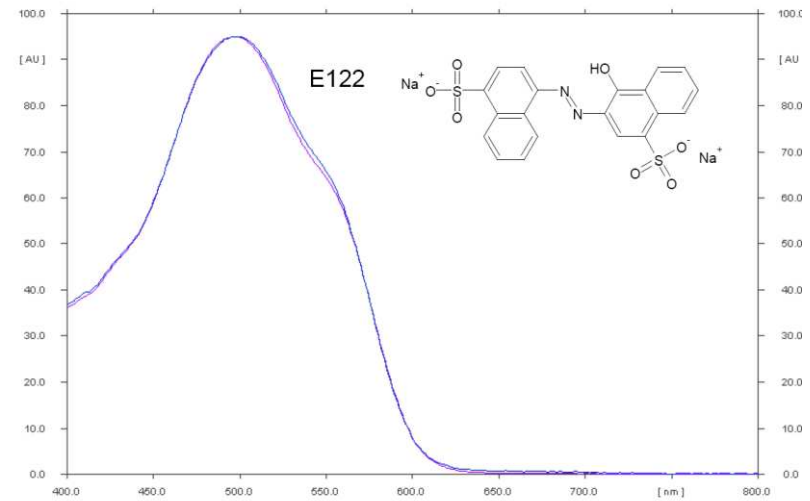
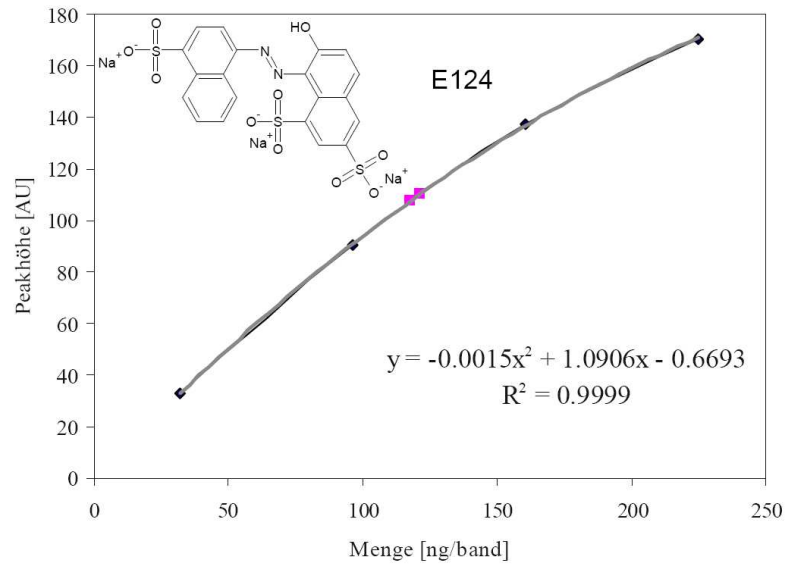
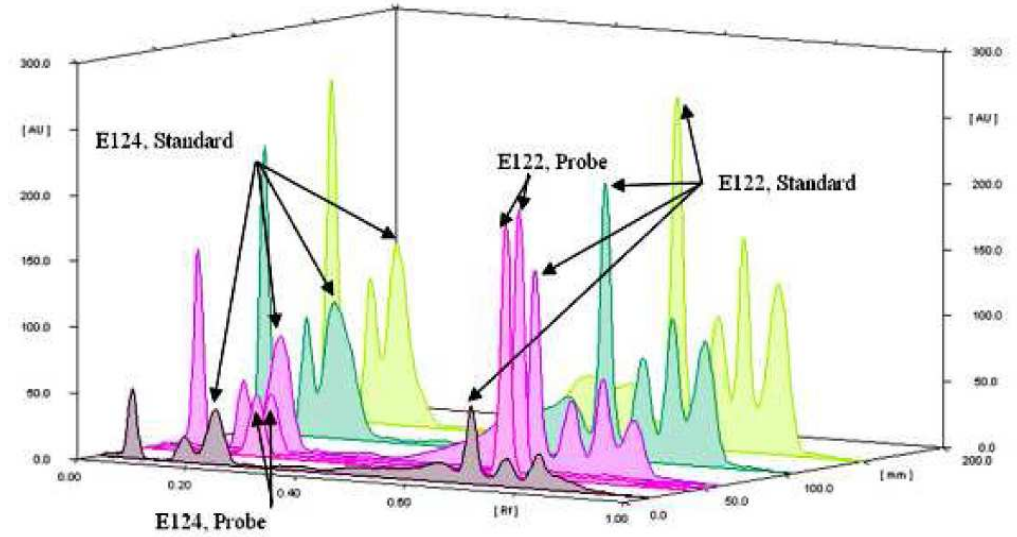
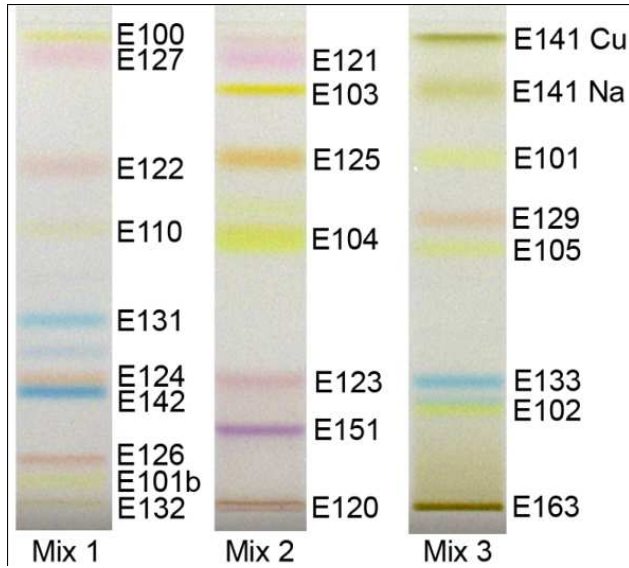


Digital filters



G. Morlock, W. Schwack, Die Aktuelle Wochenschau der GDCh,
Woche 26 (2009), www.aktuelle-wochenschau.de/2009/index09.htm

Dye analysis

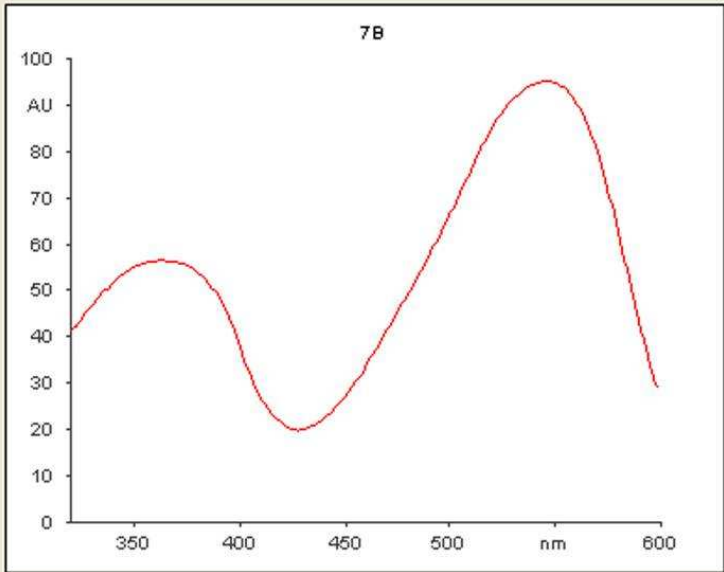




Search in spectra library

Analysis

Spectrum hRfc correction graph



7B

100
AU
80
70
60
50
40
30
20
10
0

350 400 450 500 nm 600

Add this spectrum to library ...

F:\Elodie\Sudan dyes.cna

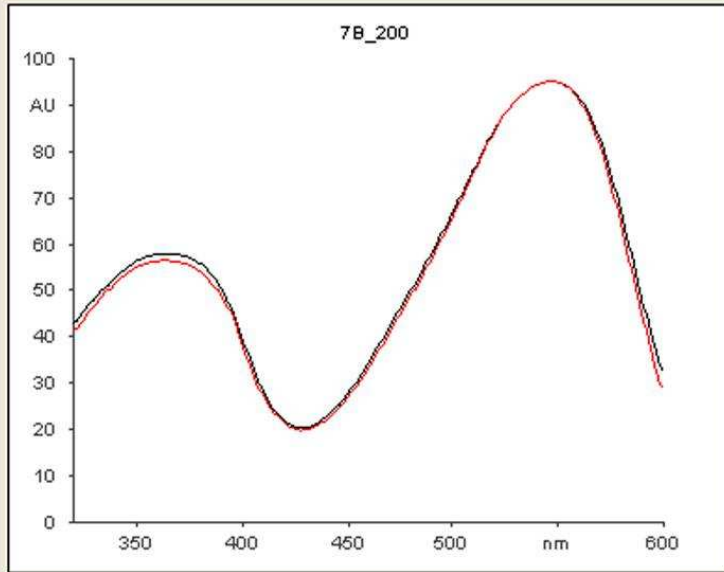
Track : 3

Position : MD Rf hRfc 0,37

Assigned substance : 7B

Library (30 spectra)

Library Overlay Correlation Difference



7B_200

100
AU
80
70
60
50
40
30
20
10
0

350 400 450 500 nm 600

Assign 7B_200 to current peak

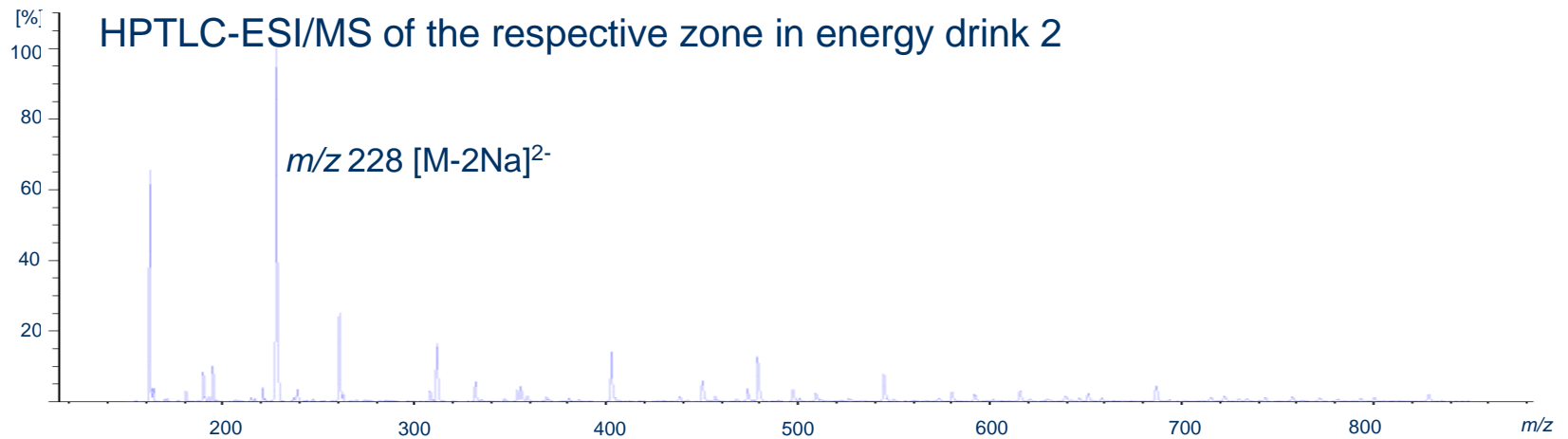
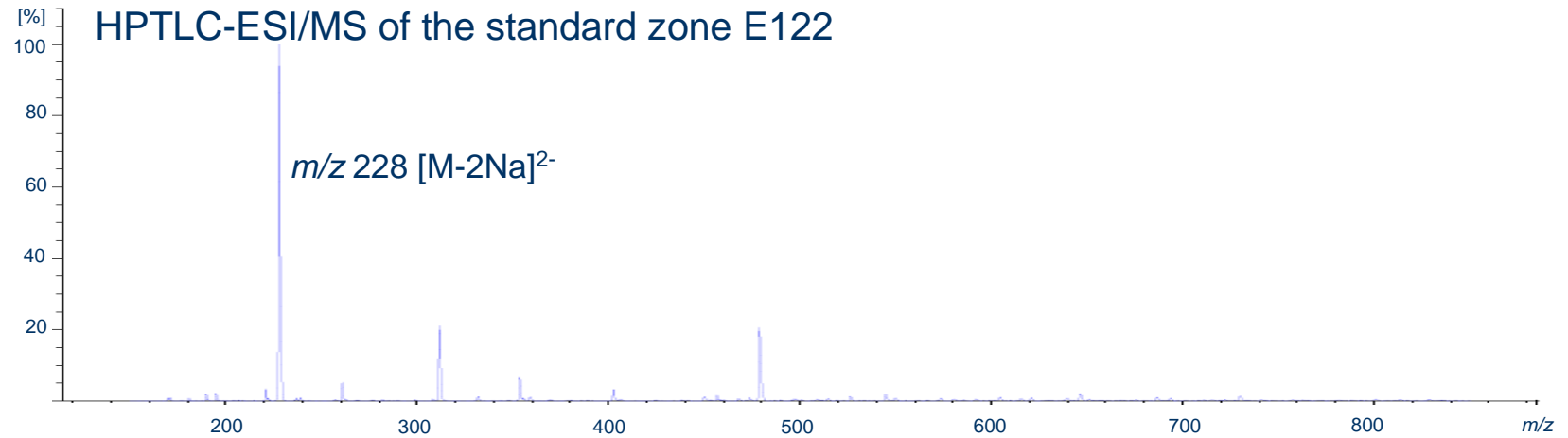
Hit	Substance	corr.	pos.
1	7B_200	0.99937	Rf 0.36
2	7B_80	0.99928	Rf 0.37
3	7B_20	0.98961	Rf 0.36
4	IV_80	0.90488	Rf 0.32
5	IV_200	0.90417	Rf 0.33

◀ ▶ \ Link files \ **Compare analysis - library** \ Edit library \ Compare library - library /





Confirmation by mass spectra





Rare examples for HPTLC

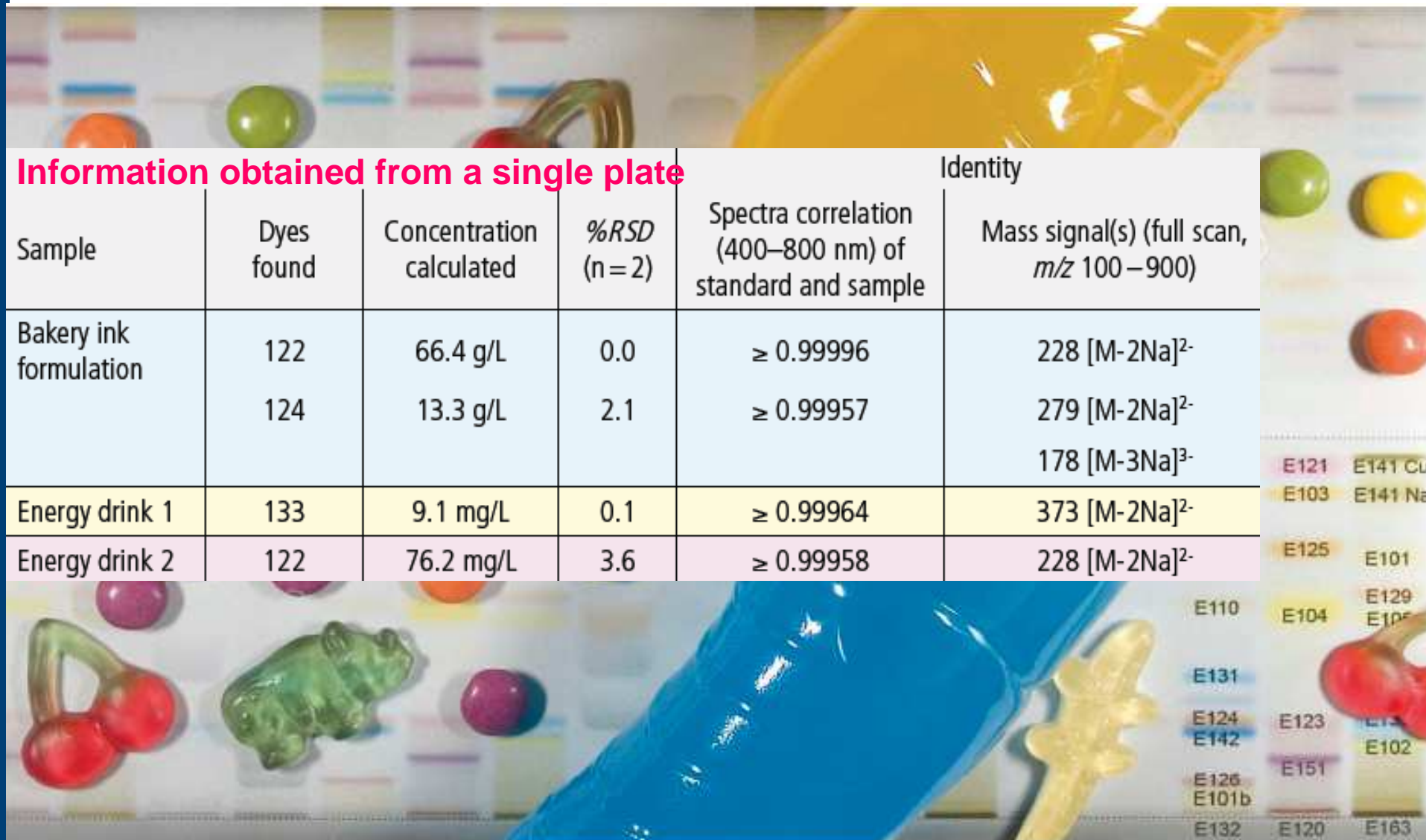
Why stop here?

Sample	Dyes found
Bakery ink formulation	122
	124
Energy drink 1	133
Energy drink 2	122



- E100
- E127
- E121
- E103
- E141 Cu
- E141 Na
- E122
- E125
- E101
- E110
- E104
- E129
- E105
- E131
- E124
- E142
- E123
- E102
- E126
- E101b
- E151
- E132
- E120
- E163

Rare examples for HPTLC



Information obtained from a single plate

Sample	Dyes found	Concentration calculated	%RSD (n=2)	Identity	
				Spectra correlation (400–800 nm) of standard and sample	Mass signal(s) (full scan, <i>m/z</i> 100–900)
Bakery ink formulation	122	66.4 g/L	0.0	≥ 0.99996	228 [M-2Na] ²⁻
	124	13.3 g/L	2.1	≥ 0.99957	279 [M-2Na] ²⁻
					178 [M-3Na] ³⁻
Energy drink 1	133	9.1 mg/L	0.1	≥ 0.99964	373 [M-2Na] ²⁻
Energy drink 2	122	76.2 mg/L	3.6	≥ 0.99958	228 [M-2Na] ²⁻

E121 E141 Cu
E103 E141 Na
E125 E101
E110 E129
E104 E105
E131
E124 E123 E13
E142 E102
E126 E151
E101b
E132 E120 E163



Cost comparison

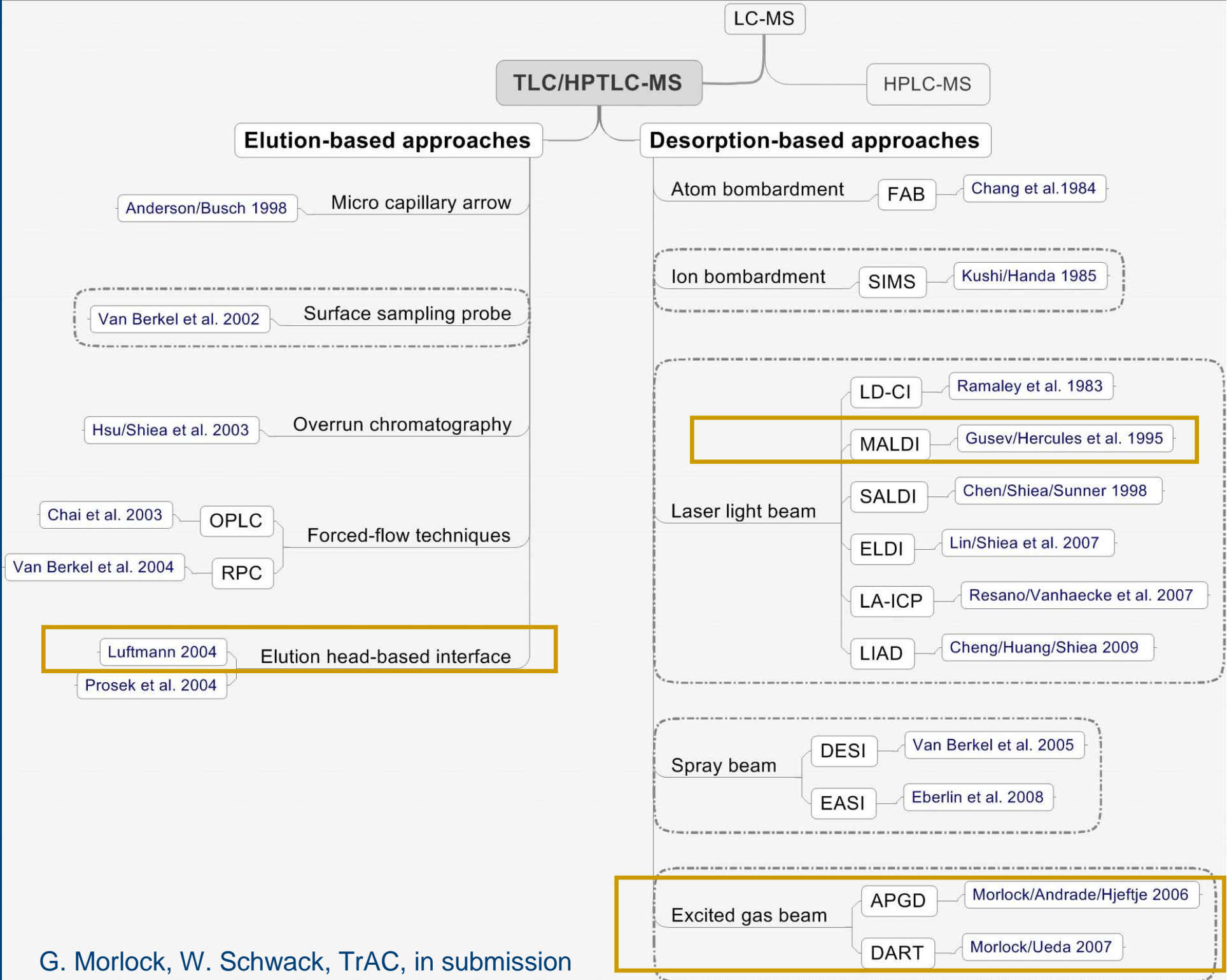
Operating costs/run (€)	HPLC ¹	HPTLC ²
Mobile phase	0,58	0,003
Stationary phase	0,64	0,11
Disposal	0,04	0,0001
Sum	1,26	0,11

=> 11 x lower

Time/run (min)	HPLC	HPTLC
Application/Injection		0,50
Run time	43	0,20
Detection		0,10
Sum	43	0,80

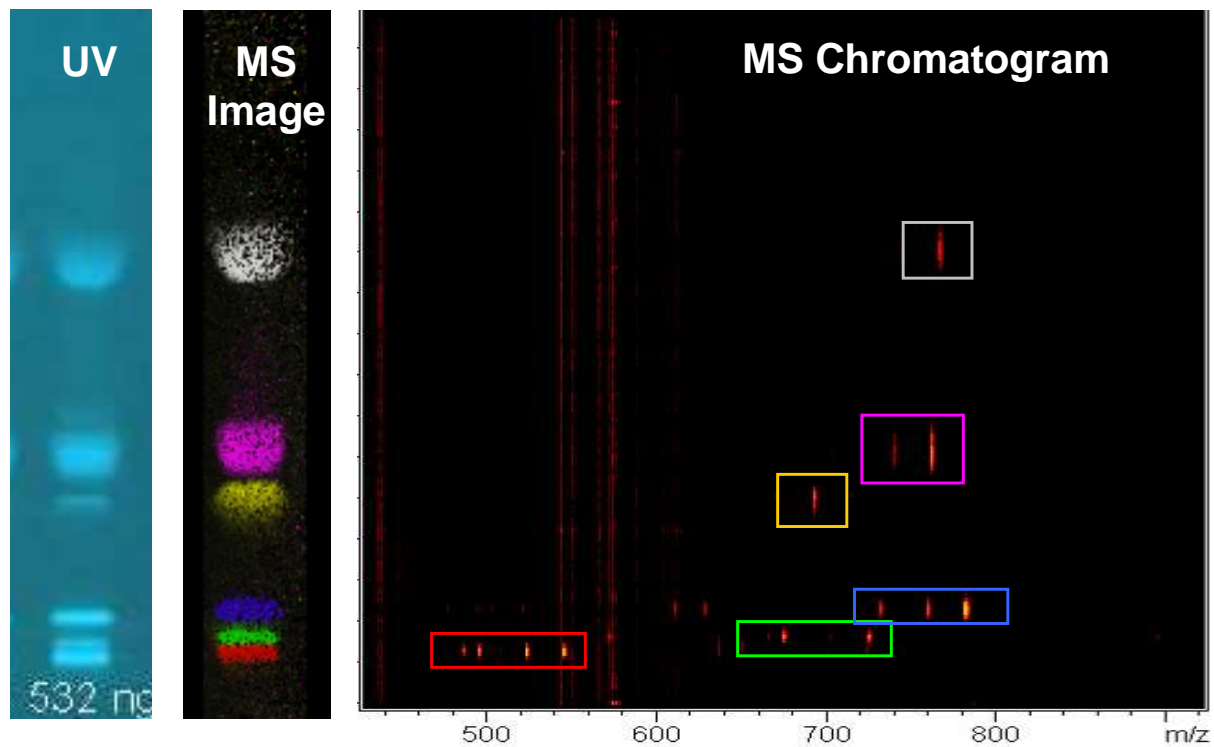
=> 54 x faster

thereof labor time/40 runs	none	5 min
----------------------------	------	-------





HPTLC-FLD-MALDI-TOF MS



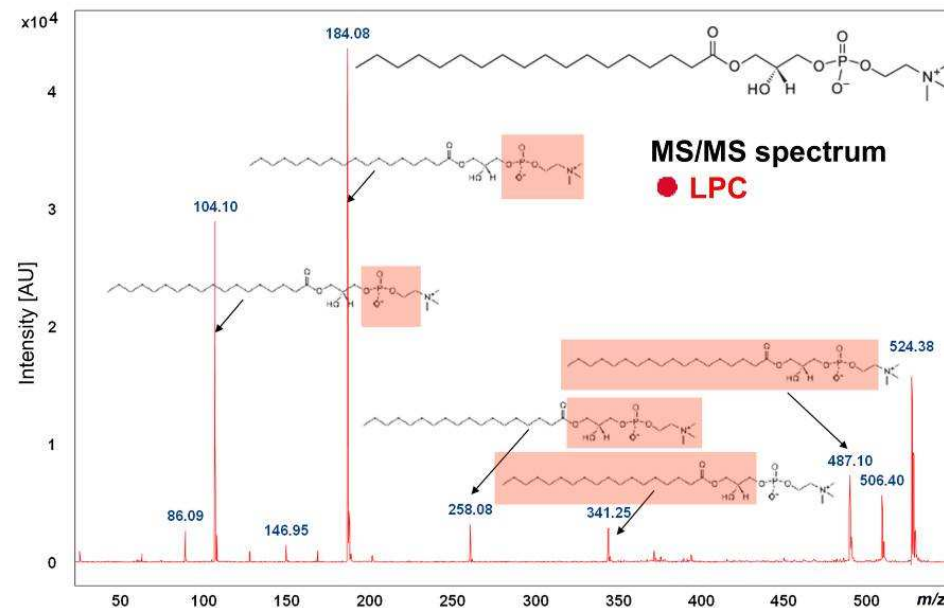
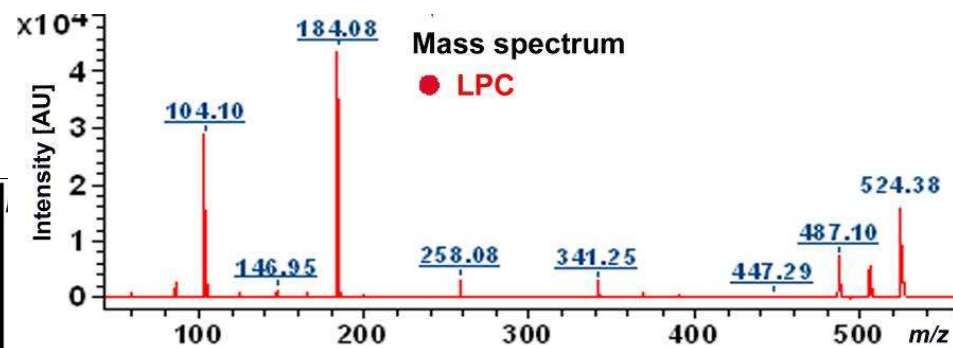
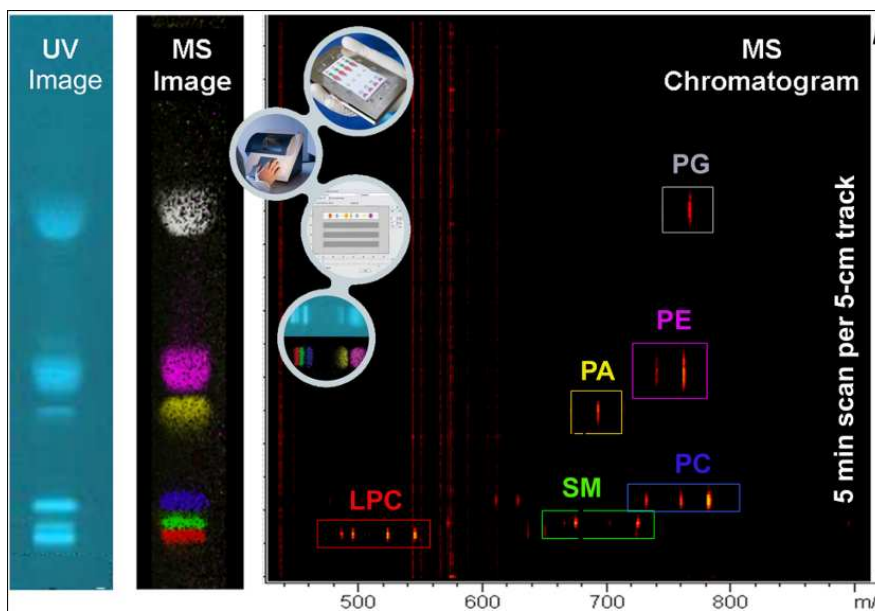
5 min



M. Schuerenberg *et al.*, IMSC 2009, Bremen, Poster PMM 386

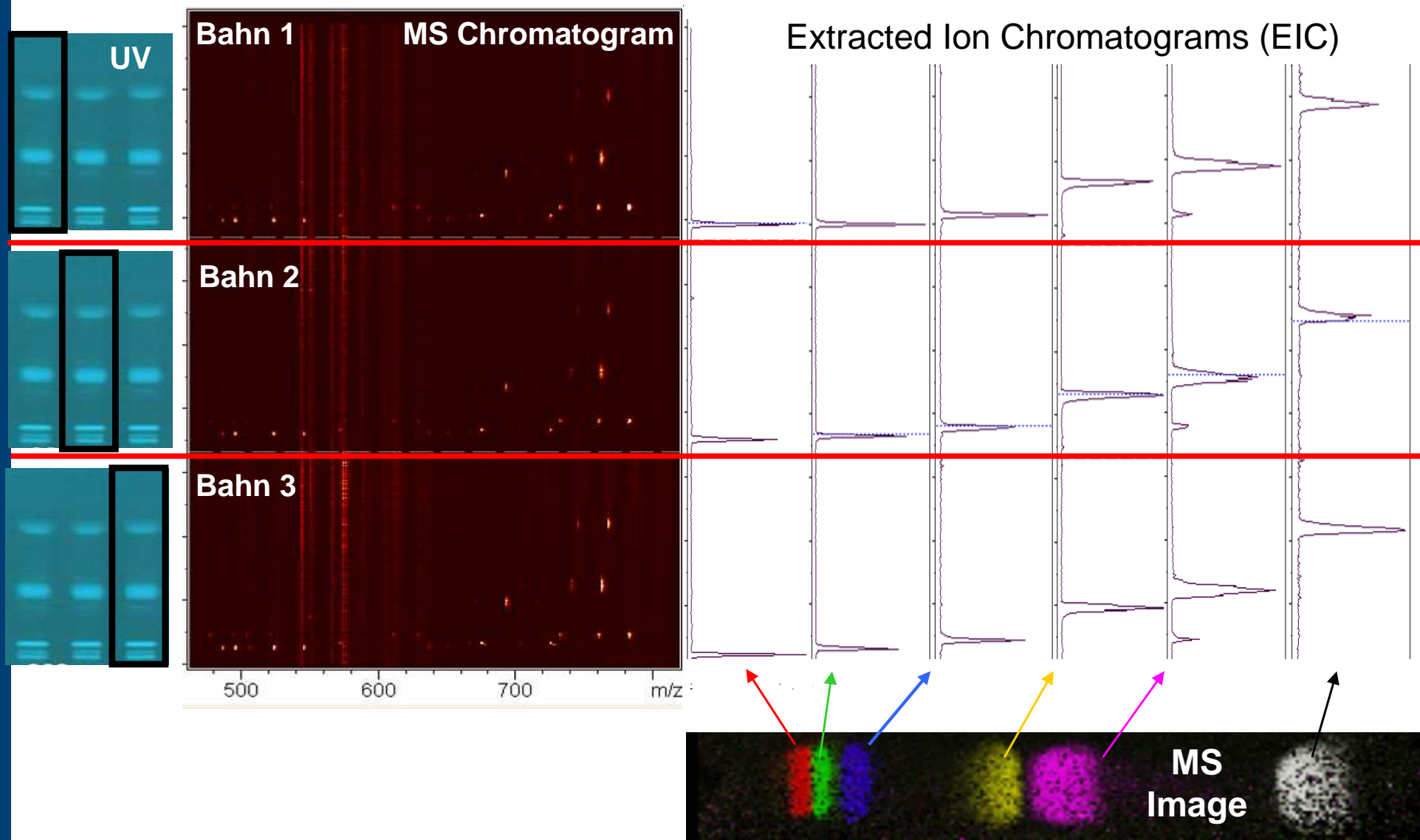


HPTLC-FLD-MALDI-TOF MS



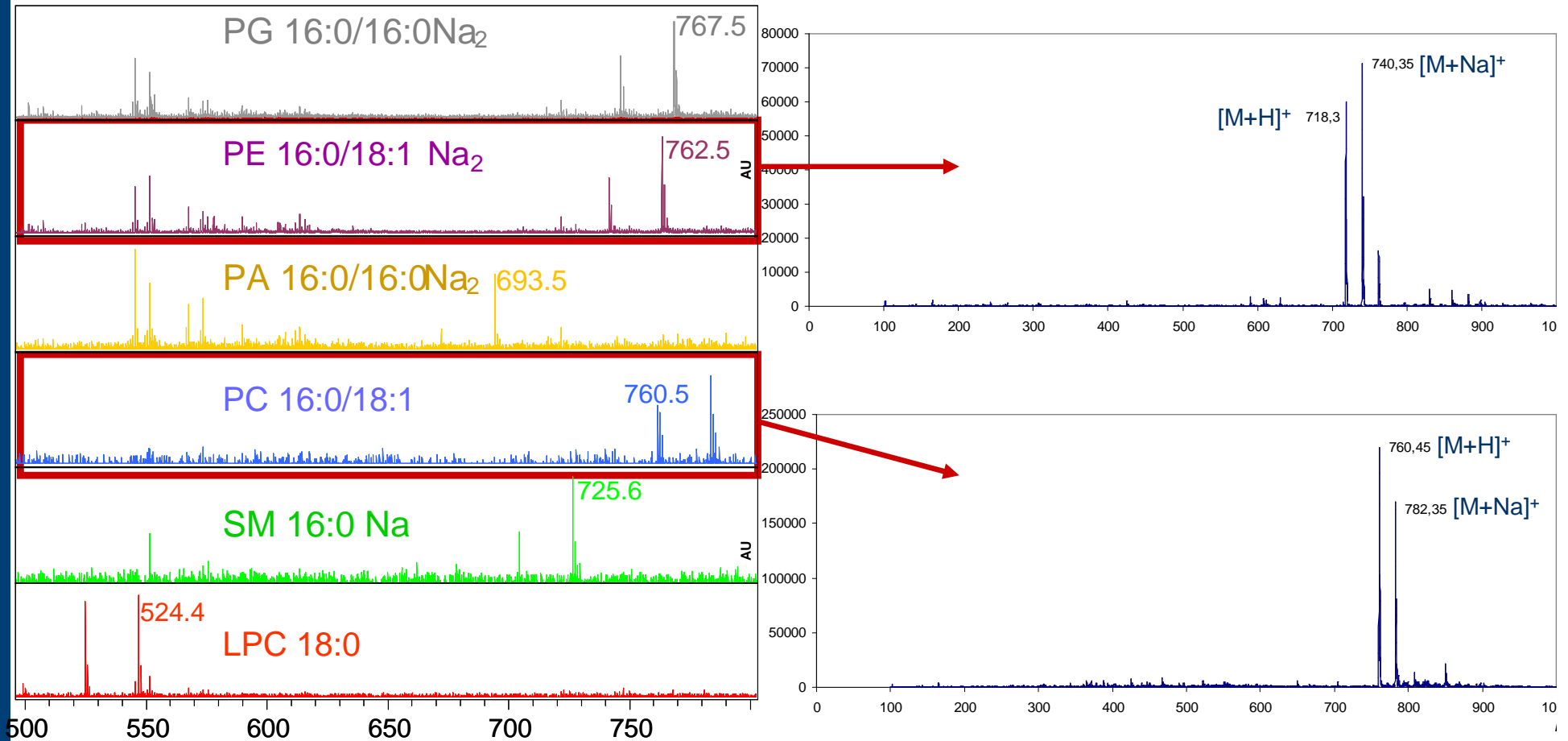


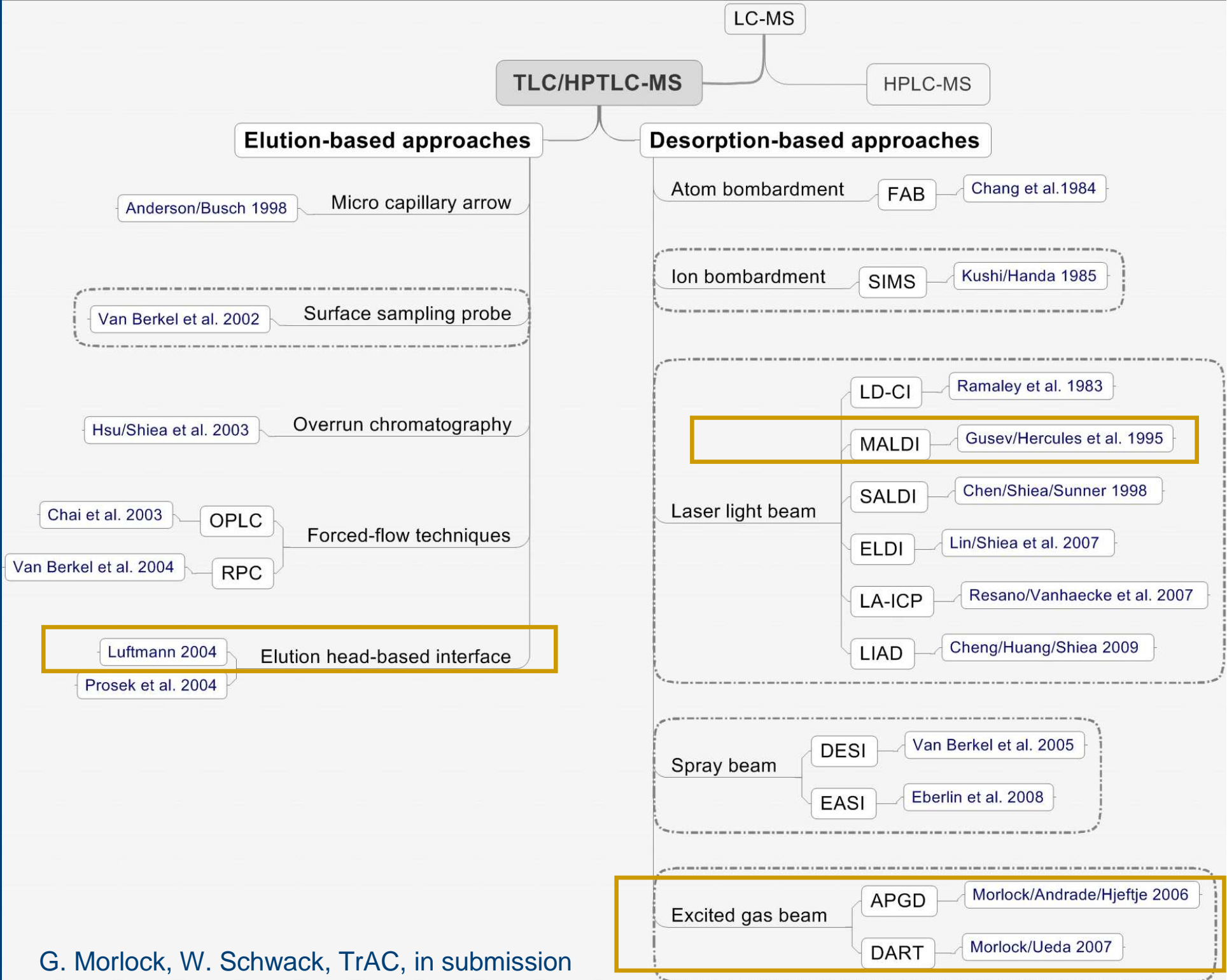
Quantification?



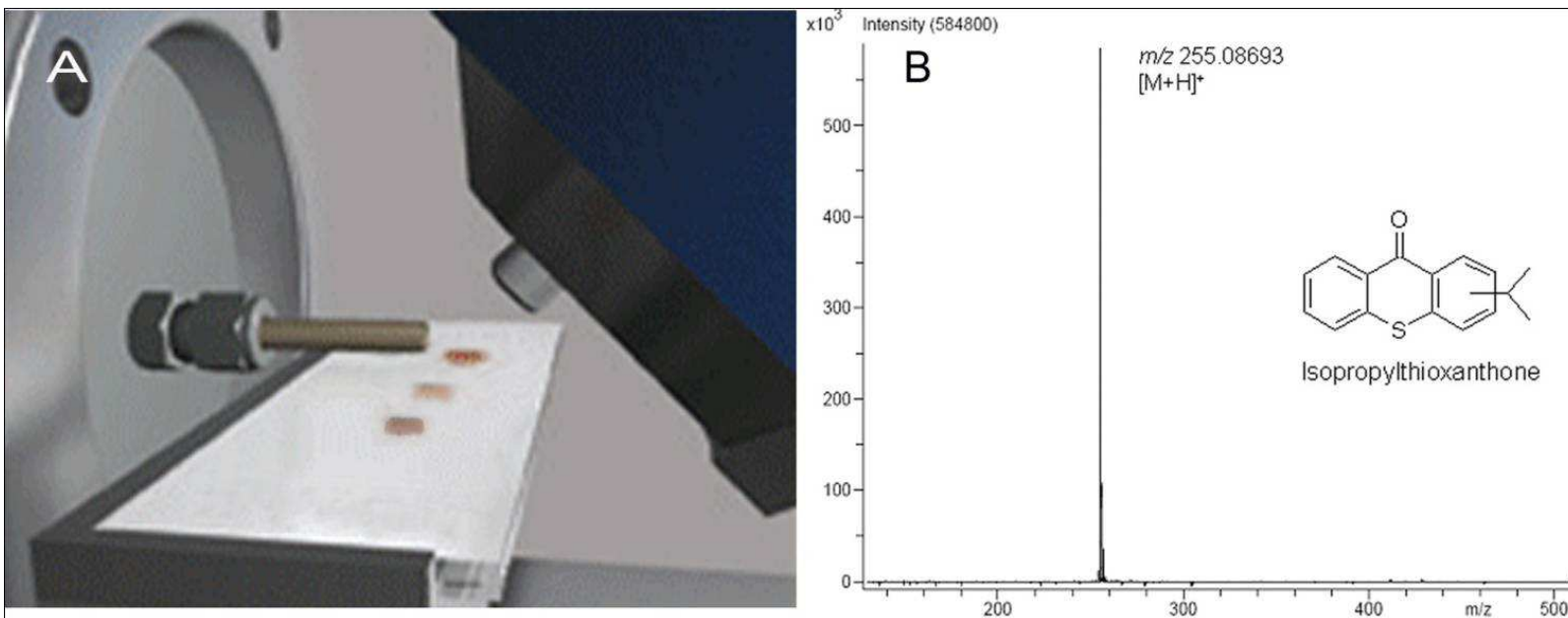


Comparison of mass spectra

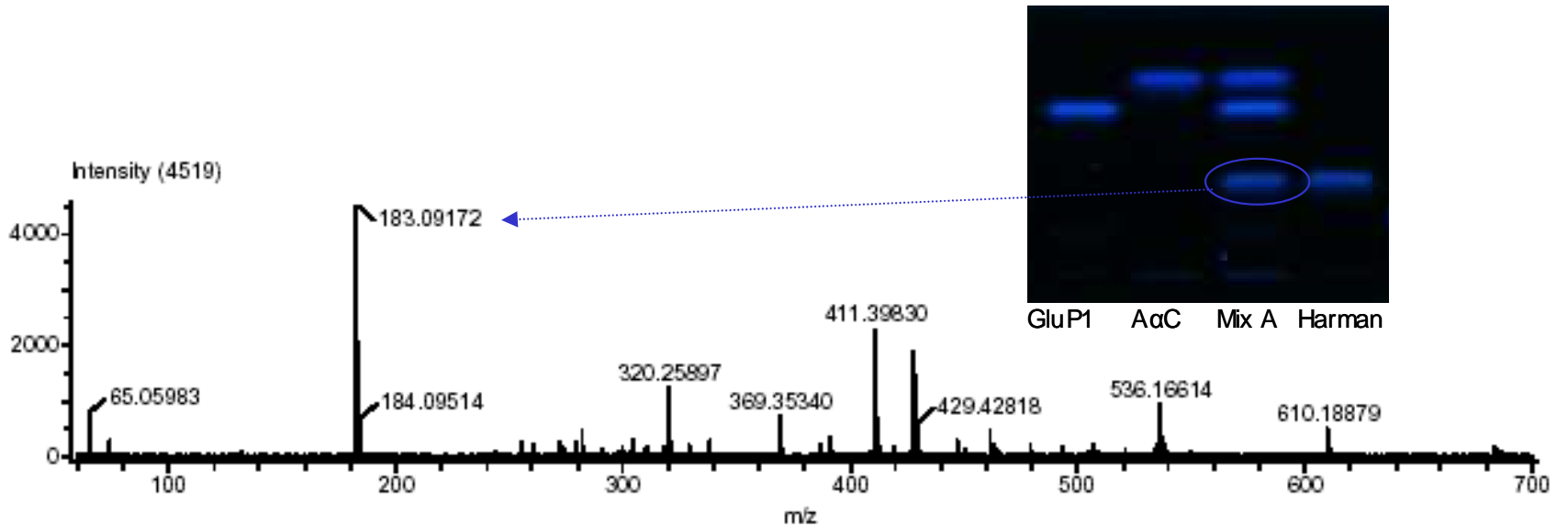
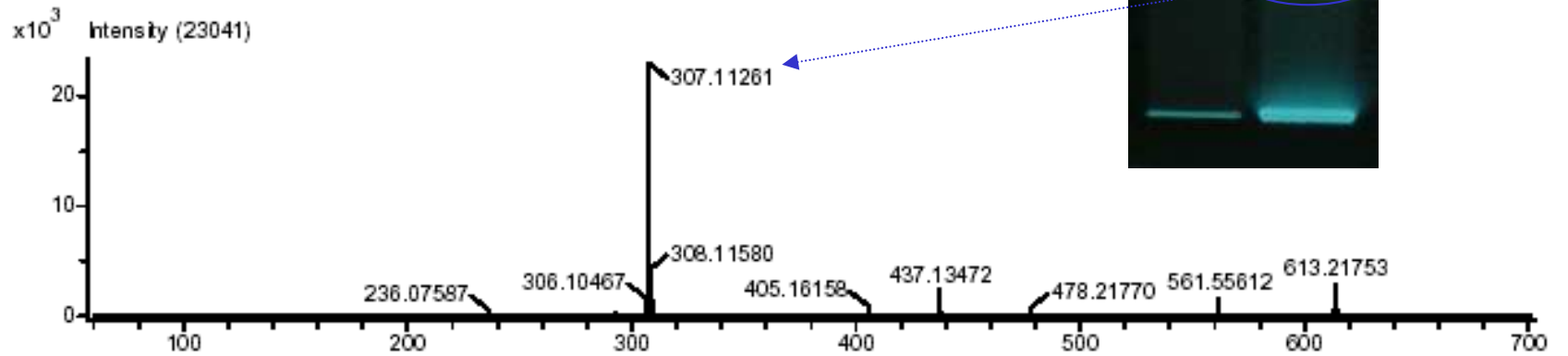




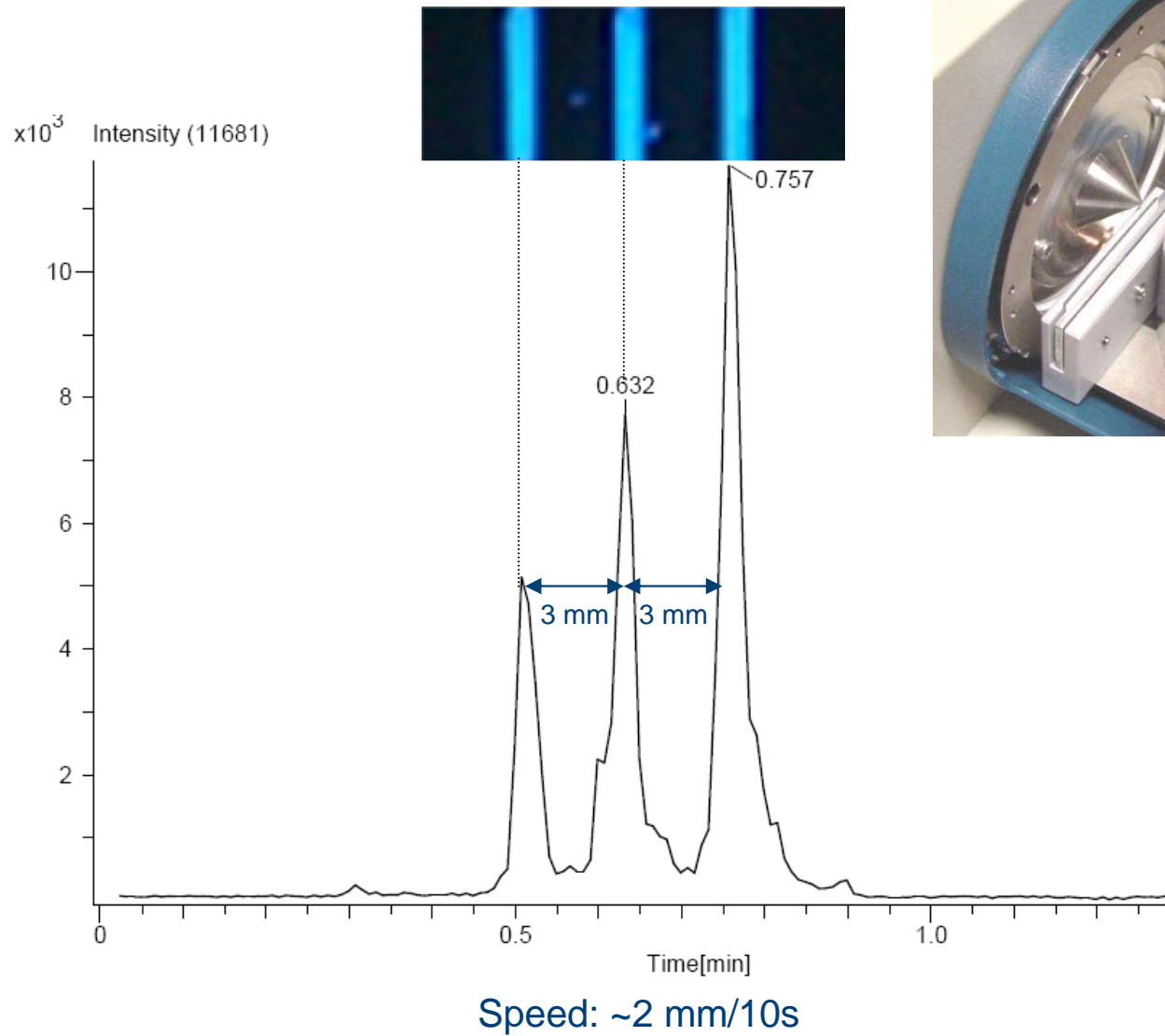
HPTLC-DART-MS



HPTLC/DART-TOF-MS



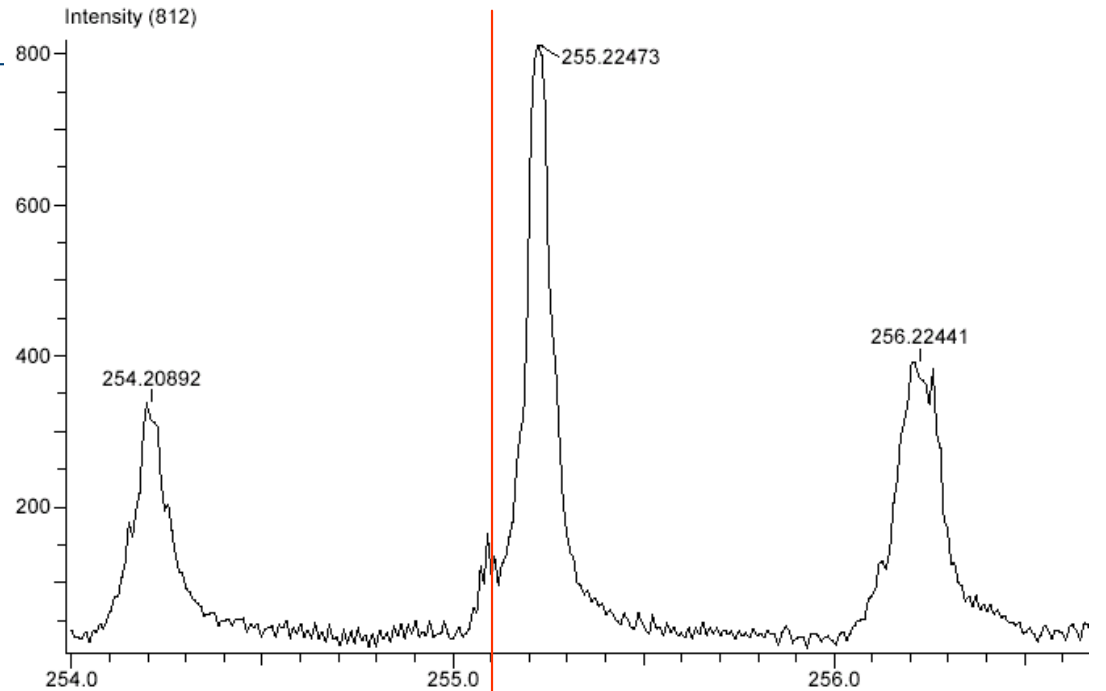
Spatial resolution of DART



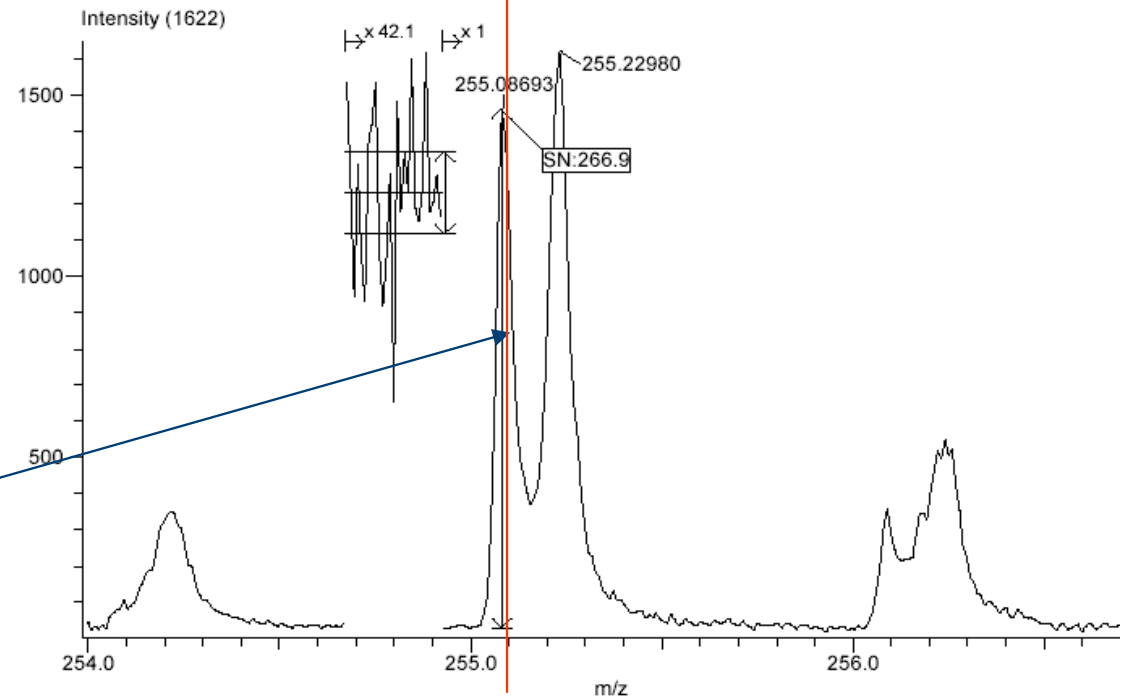


Detectability

Blank
 m/z 255.08693



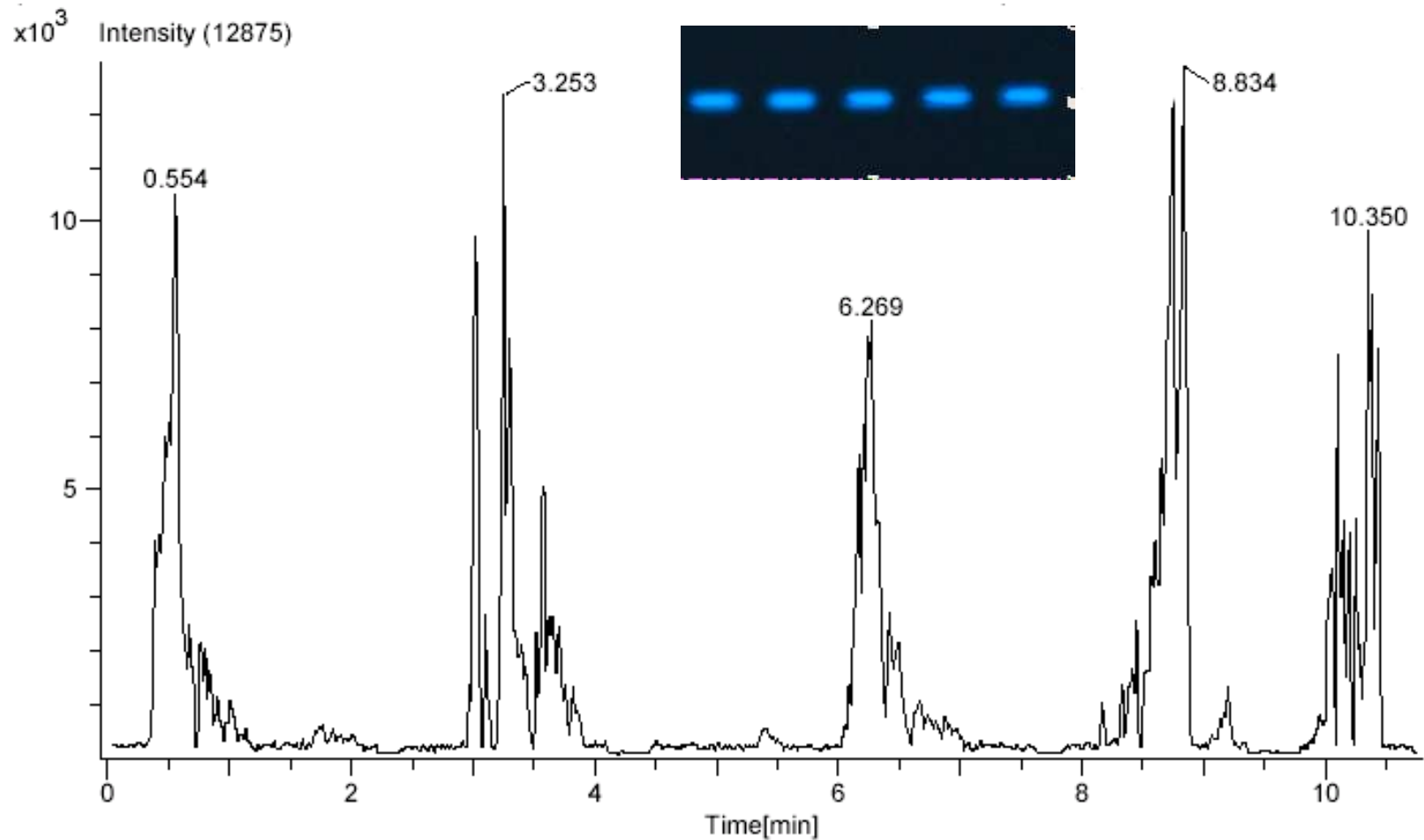
3 ng ITX
 m/z 255.08693
S/N of 267



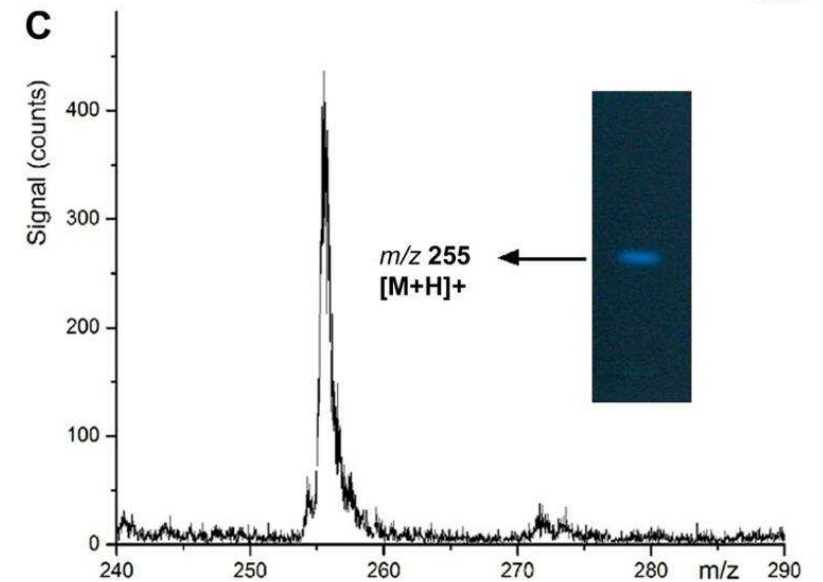
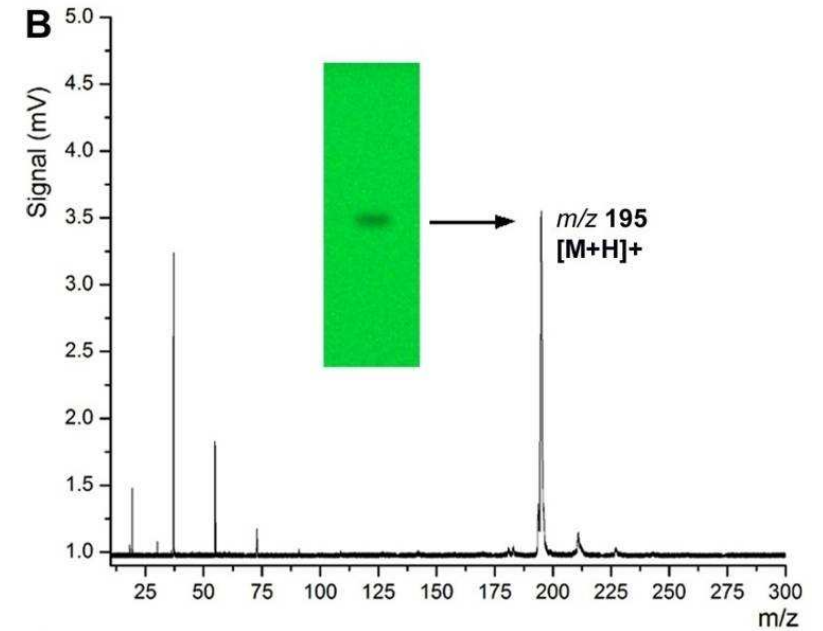
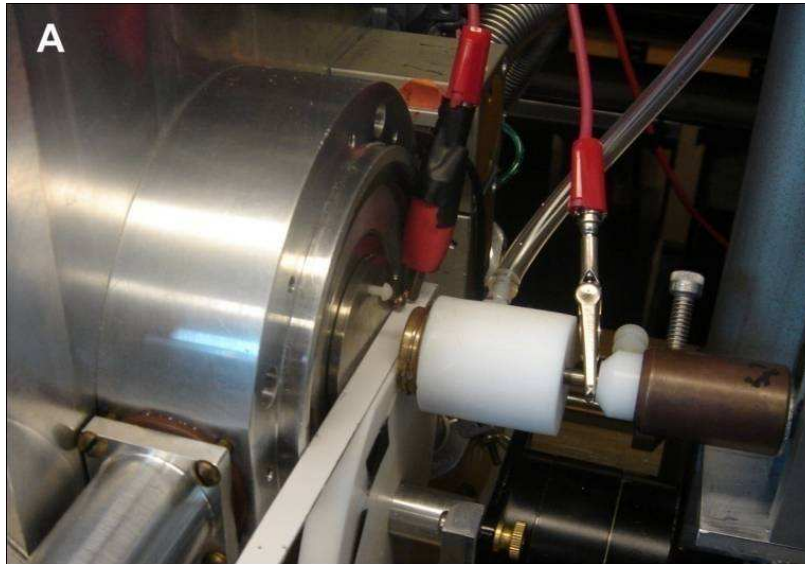
Repeatability

Mass chromatogram of ITX @ m/z 255.087 [M+H]⁺

→ %RSD = 71 % (32 ng/zone, $n = 5$)



APGD-HPTLC-TOF MS



G. Morlock, W. Schwack, TrAC, in submission



Comparison of the approaches

DART/APGD → dry desorption technique ↔ DESI



→ no plate preparation etc. ↔ SALDI, MALDI

→ ambient conditions, no high voltage ↔ micro junction

→ simple spectra ↔ MALDI, SIMS

→ quantitativ *with* internal standard → scan function

MALDI

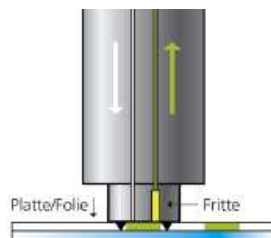


✓ strict protocol for plate preparation

✓ complex spectra

✓ quantitativ *with* internal standard → scan function

Elution-head based Interface



✓ *universally* connectable to any LC-MS system given

✓ plug & play interface (without adjustments or modifications)

✓ whole plate (no cut)

✓ all carriers on mostly all layers ↔ micro junction

✓ whole zone incl. depth profile → high detectabilities

✓ quantitativ *without* internal standard ↔ desorption techniques

✓ targeted recording → cost-effective, but *no* scan function



G Model

CHROMA-351018; No. of Pages 10

ARTICLE IN PRESS

Journal of Chromatography A, xxx (2010) xxx–xxx



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journal homepage: www.elsevier.com/locate/chroma



Review

Hyphenations in planar chromatography

Gertrud Morlock*, Wolfgang Schwack

University of Hohenheim, Institute of Food Chemistry, Garbenstrasse 28, 70599 Stuttgart, Germany

- HPTLC-UV/Vis/FLD-MS [13,14],
- HPTLC-UV/Vis/FLD-bioactivity-HRMS [15],
- HPTLC-UV-FTIR [16,17],
- HPTLC-UV/Vis/FLD-FTIR ATR [18],
- TLC-Vis-SERS [12].

ARTICLE INFO

Article history:
Available online xxx

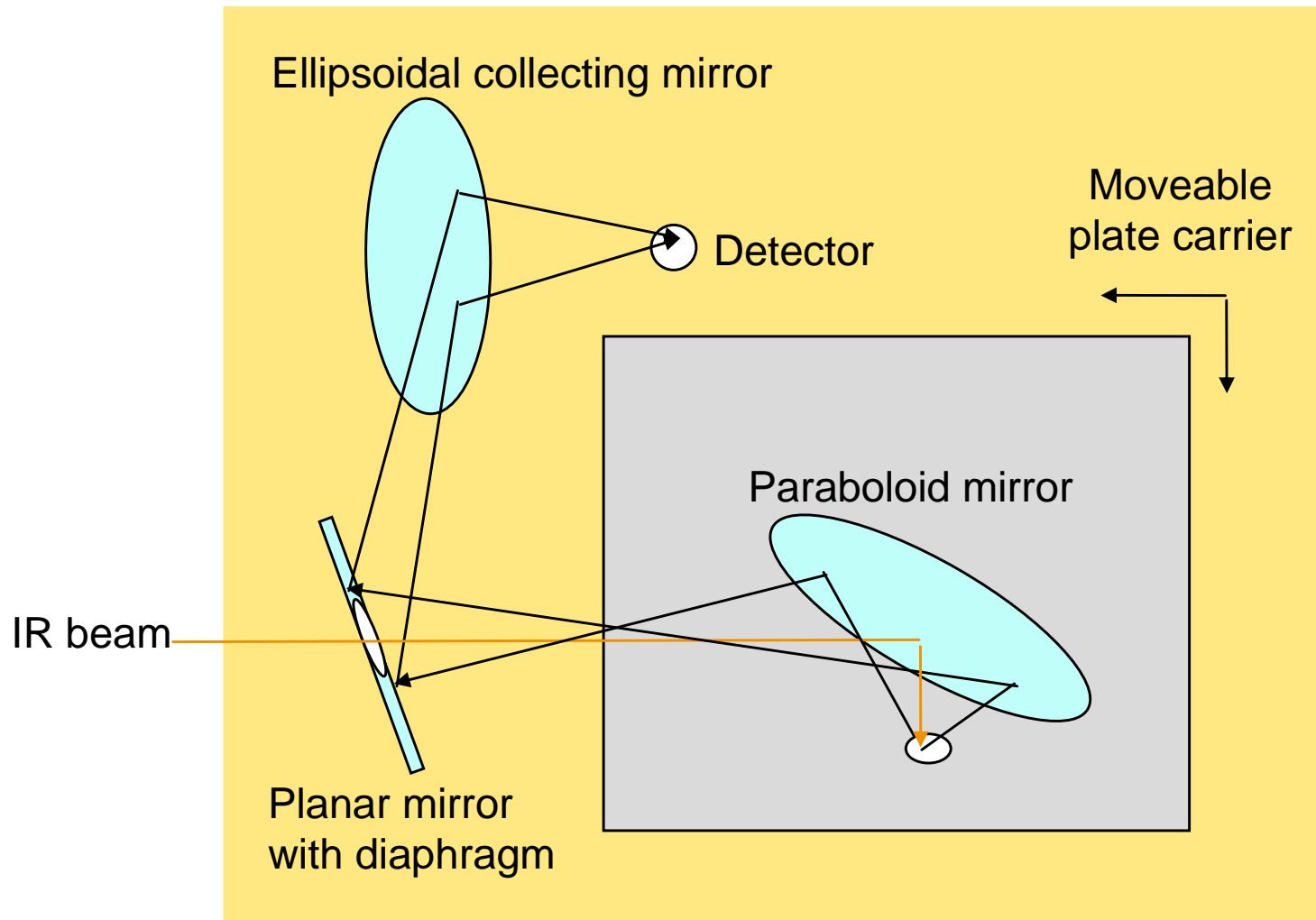
Keywords:
Mass spectrometry
High-performance thin-layer chromatography
Effect-directed analysis
Bioassays
Cost-effective analysis
High-throughput system

ABSTRACT

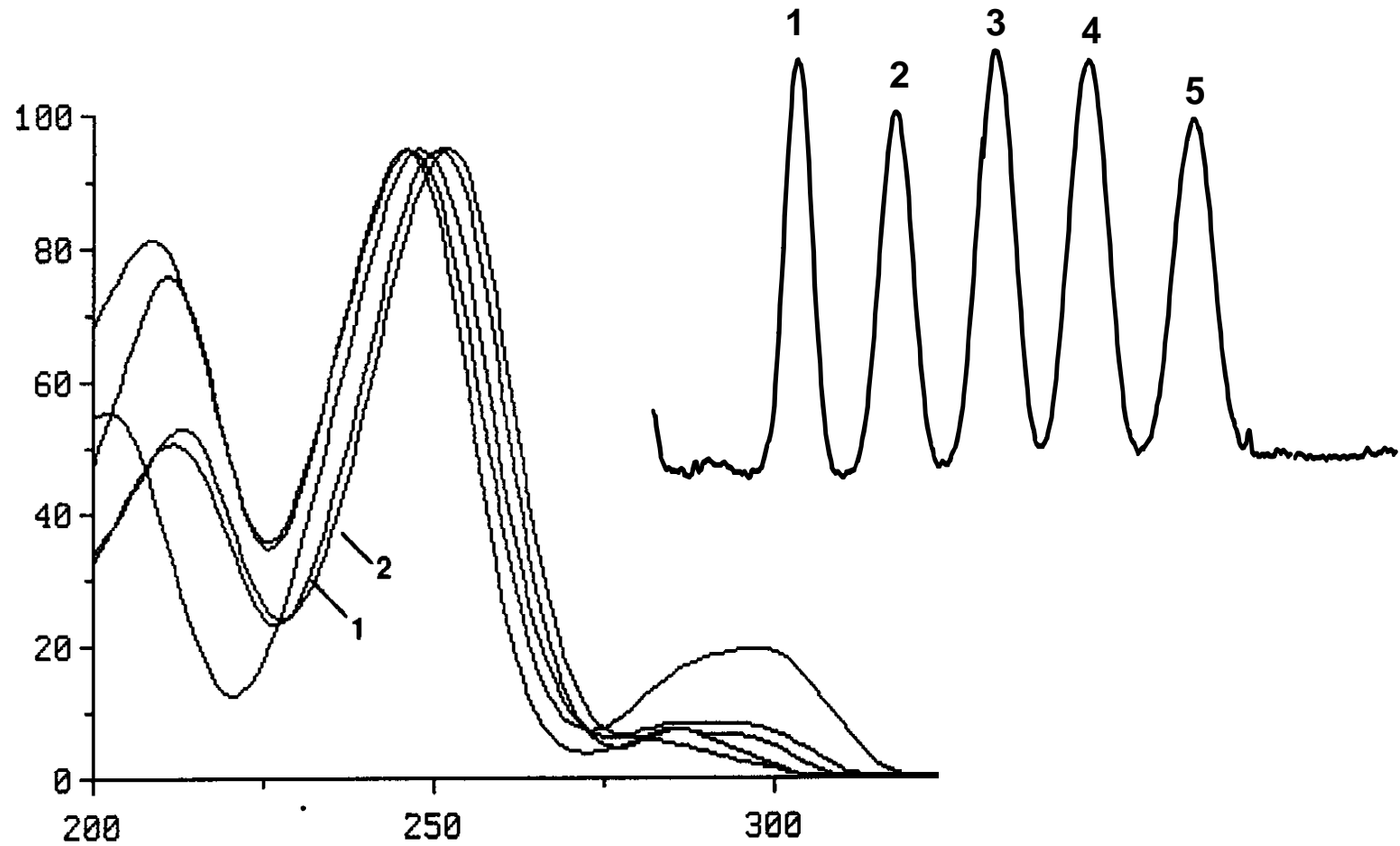
This review is focused on planar chromatography and especially on its most important subcategory high-performance thin-layer chromatography (HPTLC). The image-giving format of the open, planar stationary phase and the post-chromatographic evaporation of the mobile phase ease the performance of various kinds of hyphenations and even super-hyphenations. Examples in the field of natural product search, food and lipid analysis are demonstrated, which point out the hyphenation with effect-directed analysis (EDA) and mass spectrometry and illustrate the efficiency gain. Depending on the task at hand, hyphenations can readily be selected as required to reach the relevant information about the sample, and at the same time, information is obtained for many samples in parallel. The flexibility and the unrivalled features through the planar format valuably assist separation scientists.

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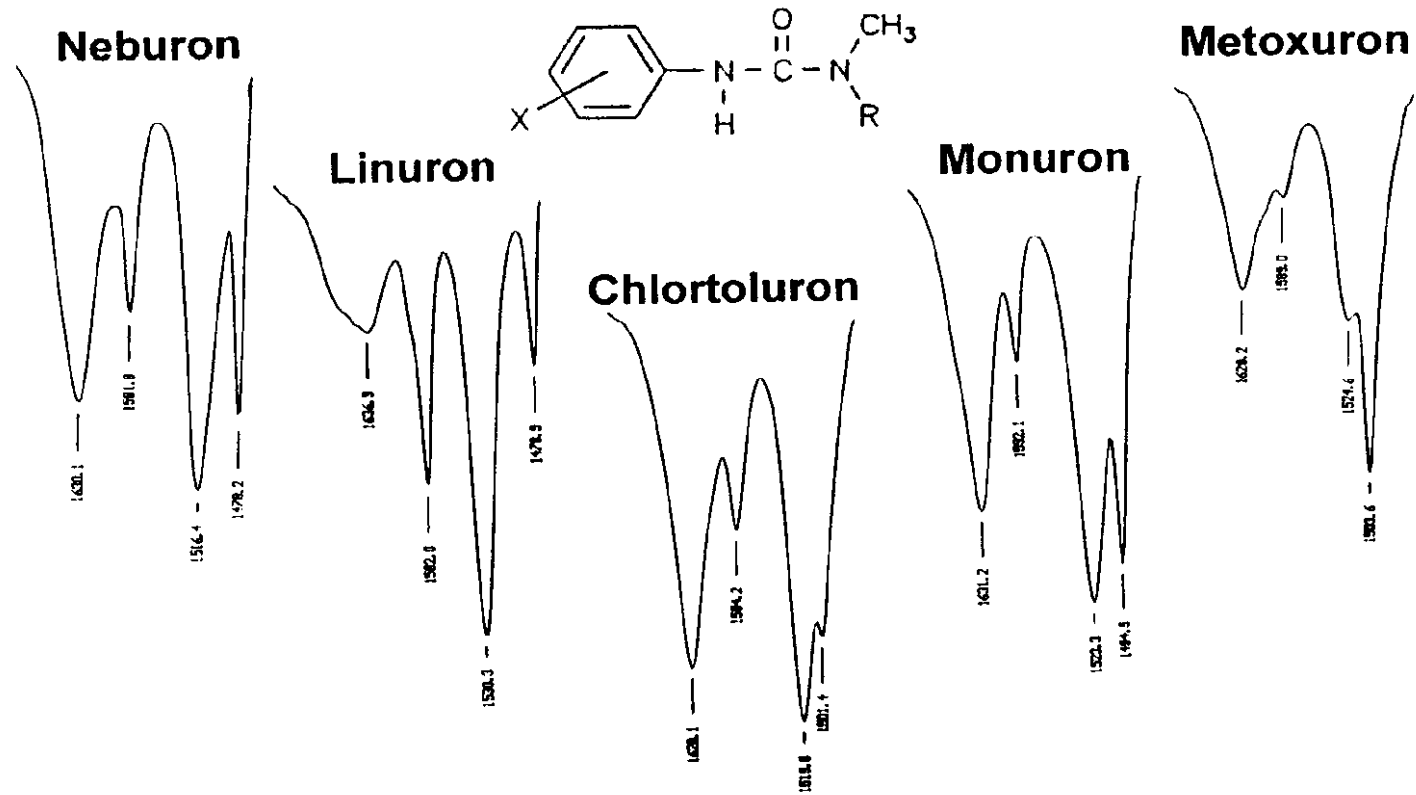
HPTLC-DRIFT



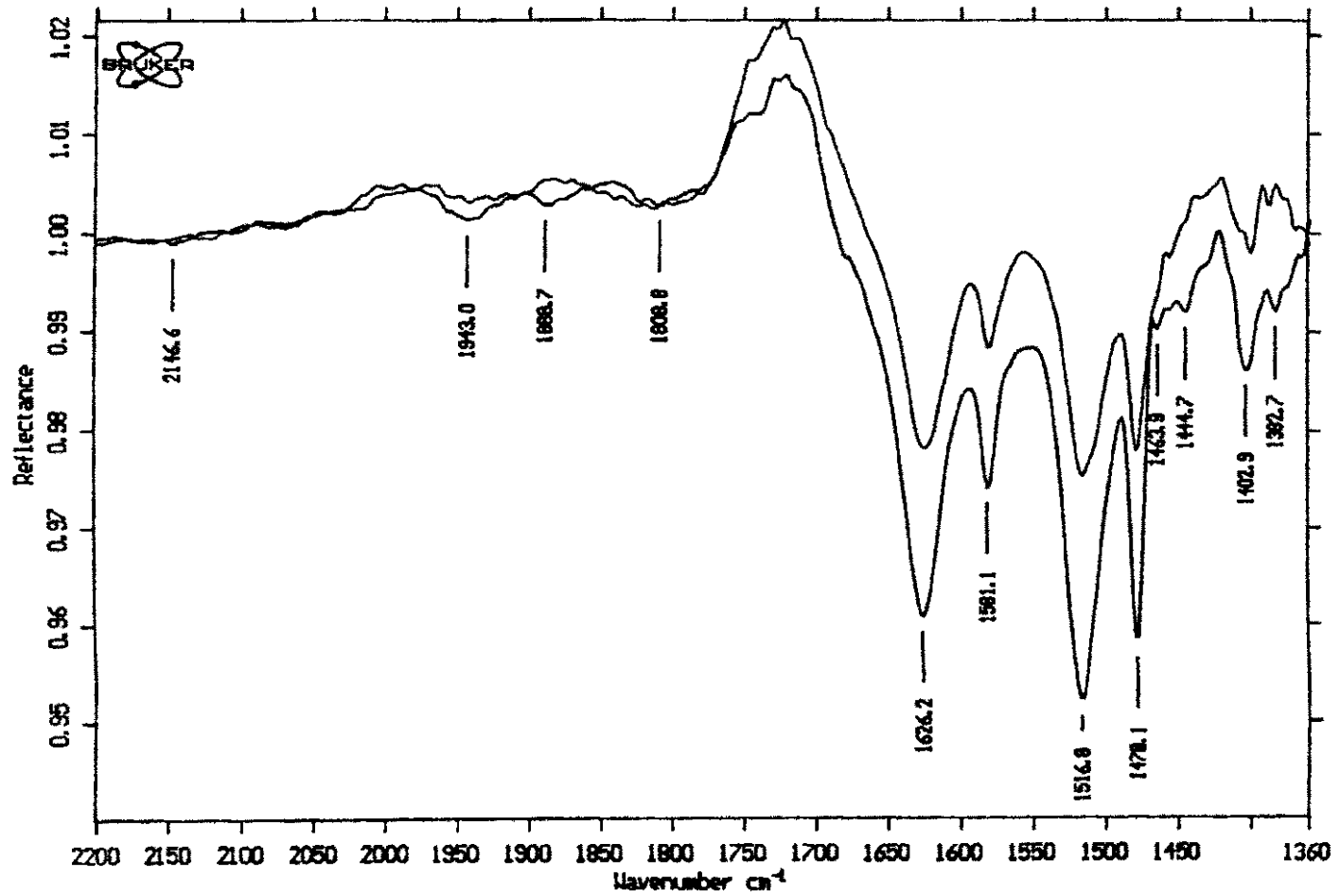
UV-spectra of 5 phenyl urea herbicides



Characteristic FTIR bands



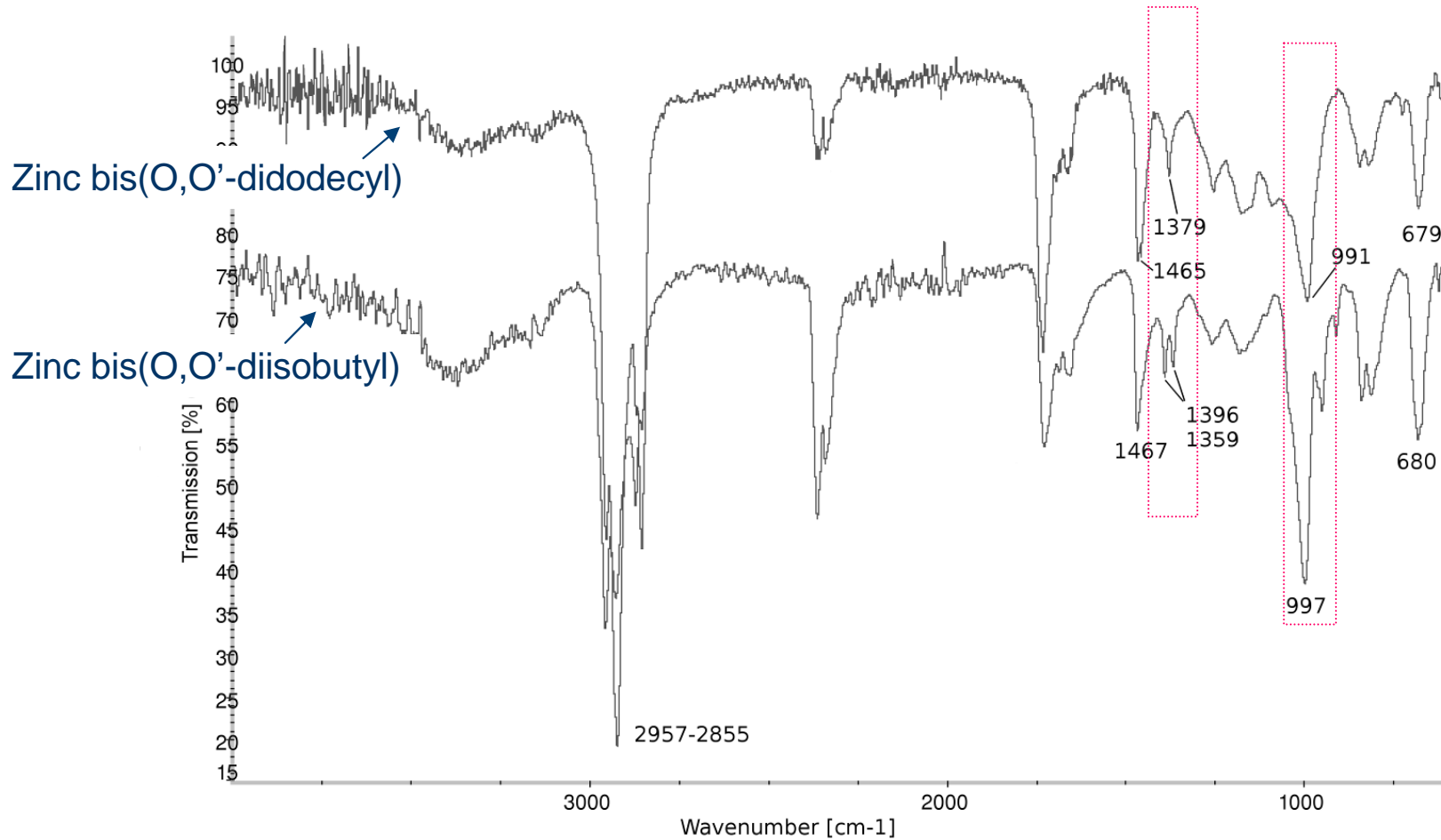
FTIR spectrum of neburon in drinking water





HPTLC/ATR-IR spectra via the interface

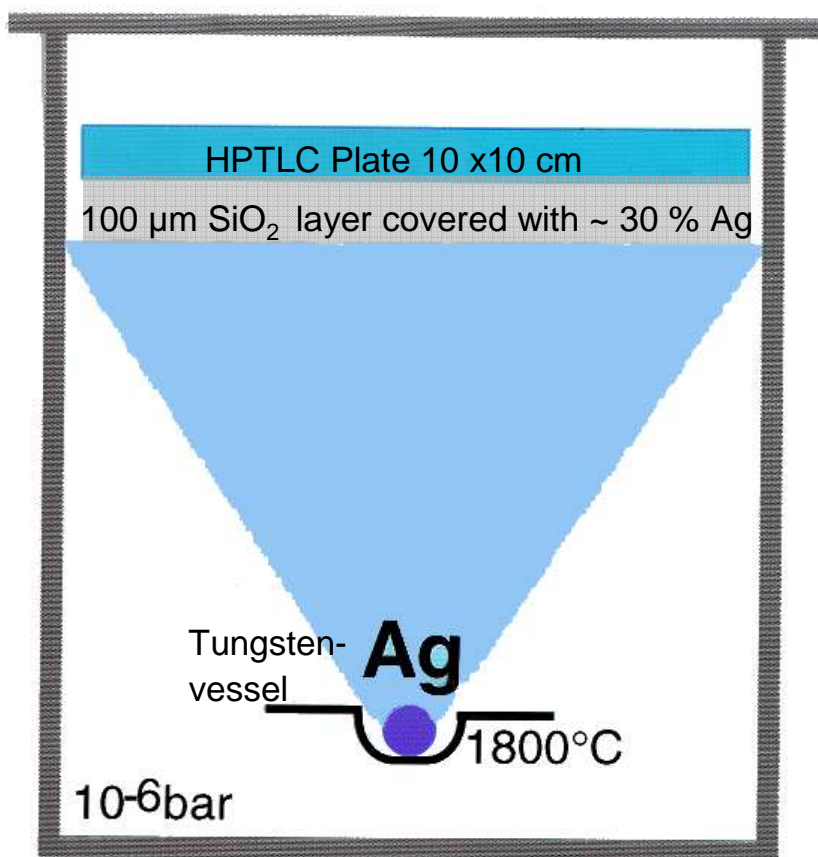
Dithiophosphate additives in mineral oil



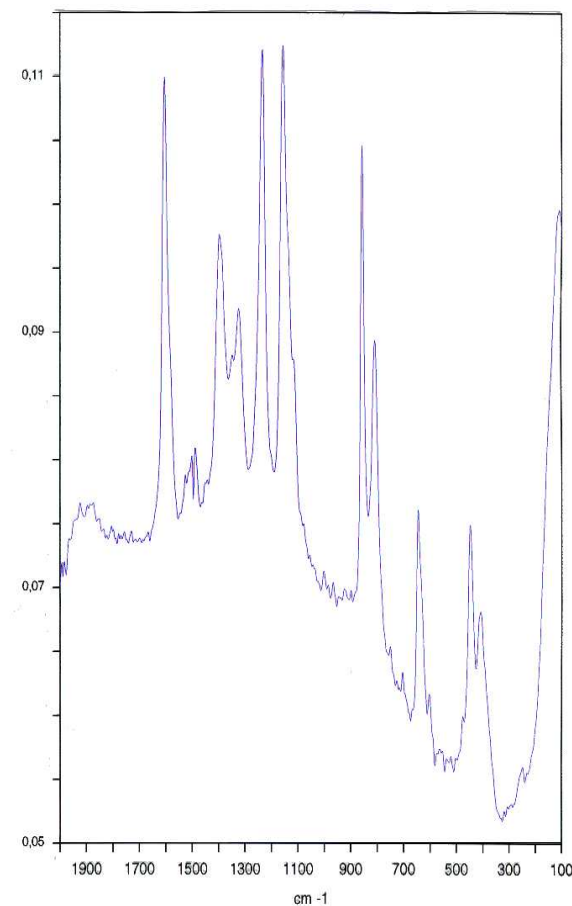
Raman: FT-SERS

→ based on the work of Dr. Klaus Burgert; Bayer Laboratories, Germany

Vacuum transfer



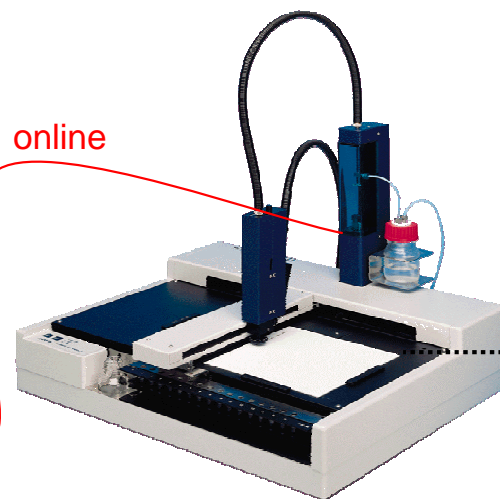
10 ng/zone p-nitrophenol



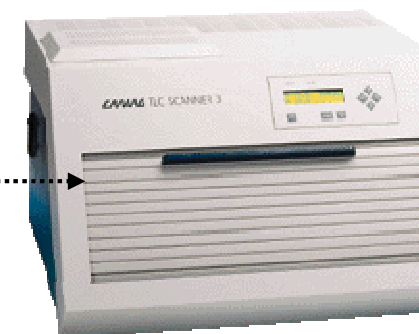
HPLC-HPTLC



Fractions (1-2 min each)
controlled by cutting standards
are applied as area

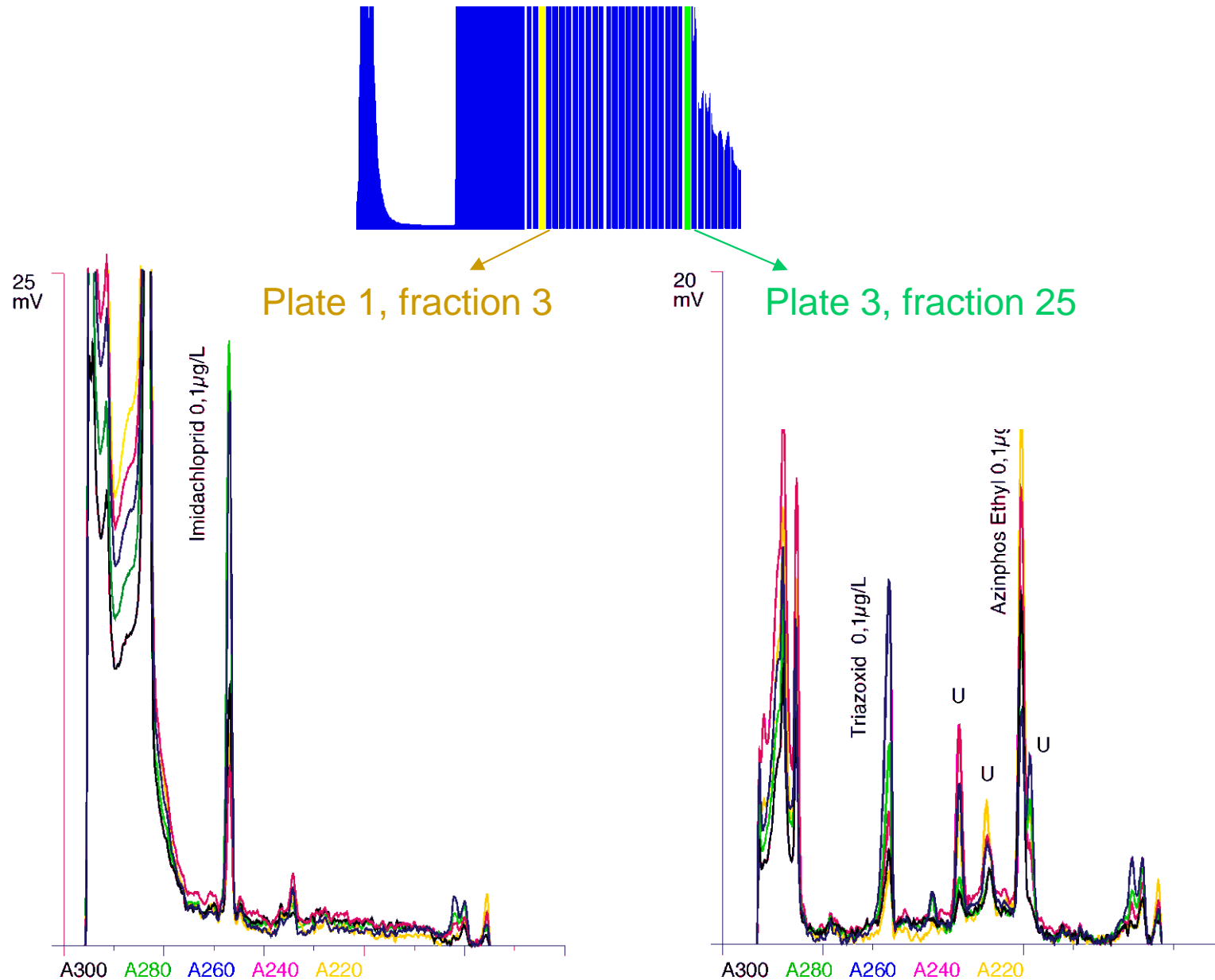


HPTLC analysis of 18 tracks (fractions) per plate



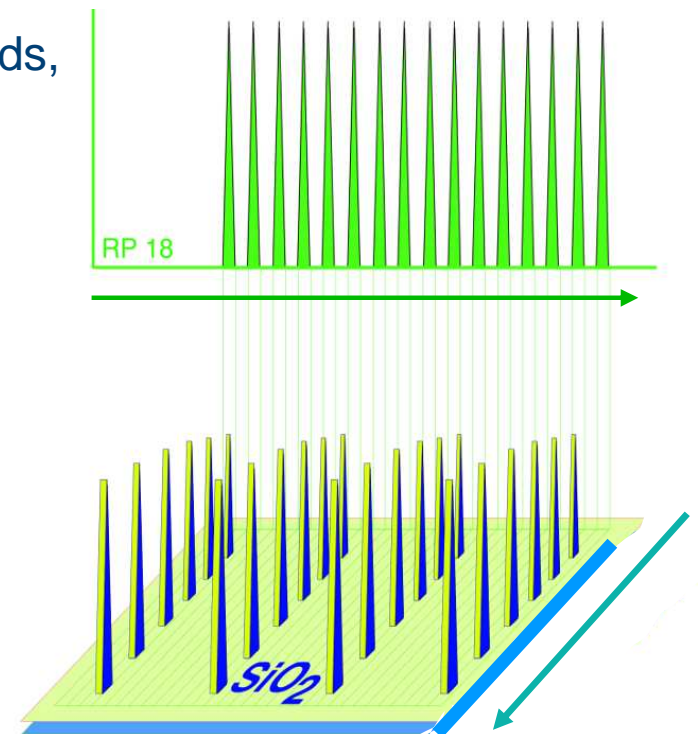
Microbore 2.1 x 100 mm
Flow rate 50-100 μ L/min

Surface water spiked with 50 pesticides

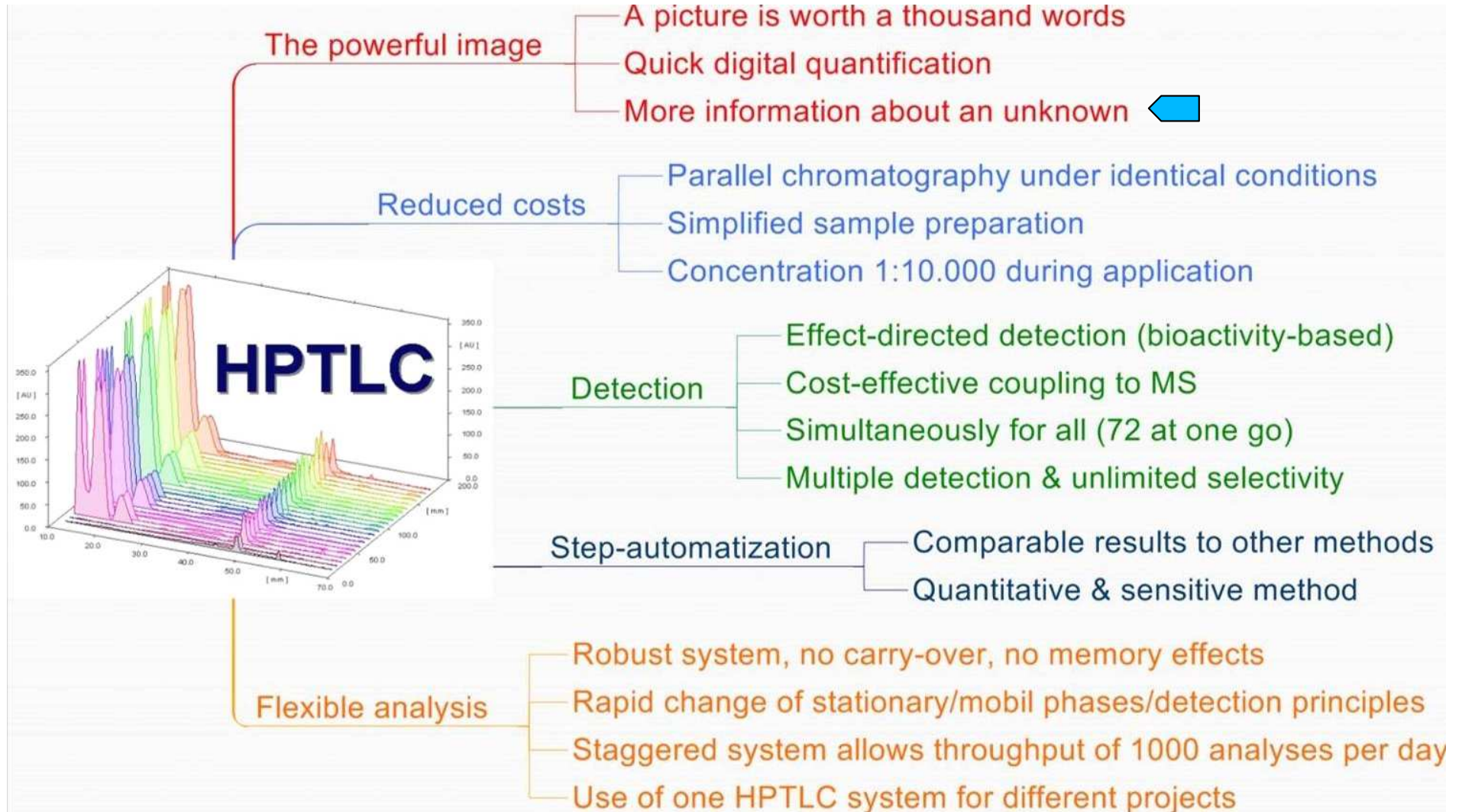


HPLC-HPTLC

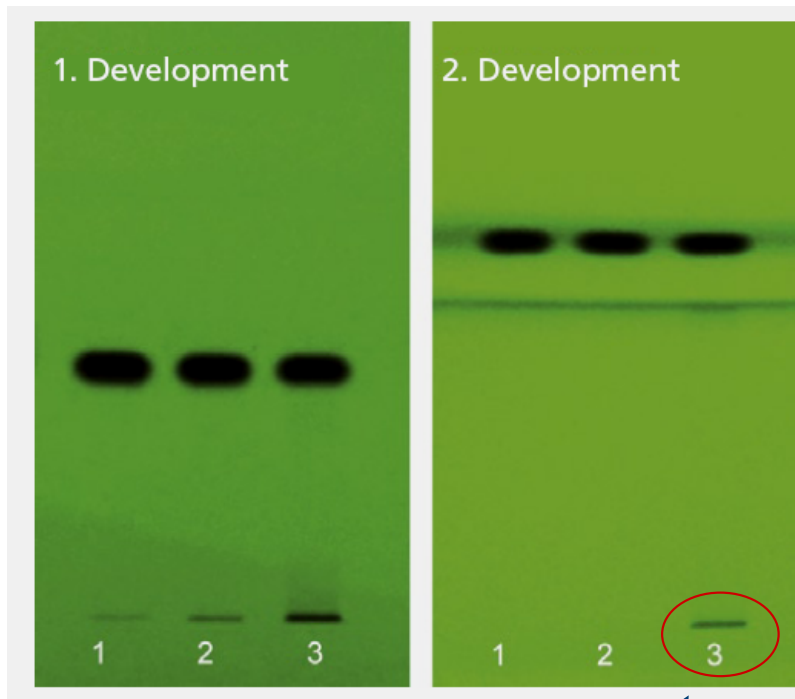
- based on the work of Dr. Klaus Burger†, Bayer Laboratories, Germany
- predestinated for
 - ☺ separation problems due to lack of separation power
 - ☺ samples with varying matrix content (multi-method)
 - ☺ peak purity testing of HPLC peaks
 - ☺ problematic, time-consuming post-chromatographic derivatization in HPLC
- results obtained by two independent methods, i.e. two different separation mechanisms
- gain in analytical quality
- still in use by Andreas Kinast
andreas.kinast@currenta.de
www.currenta.de



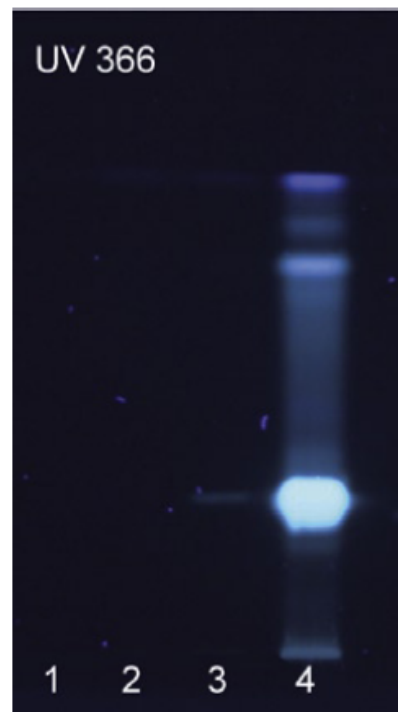
Benefits



More information about unknowns

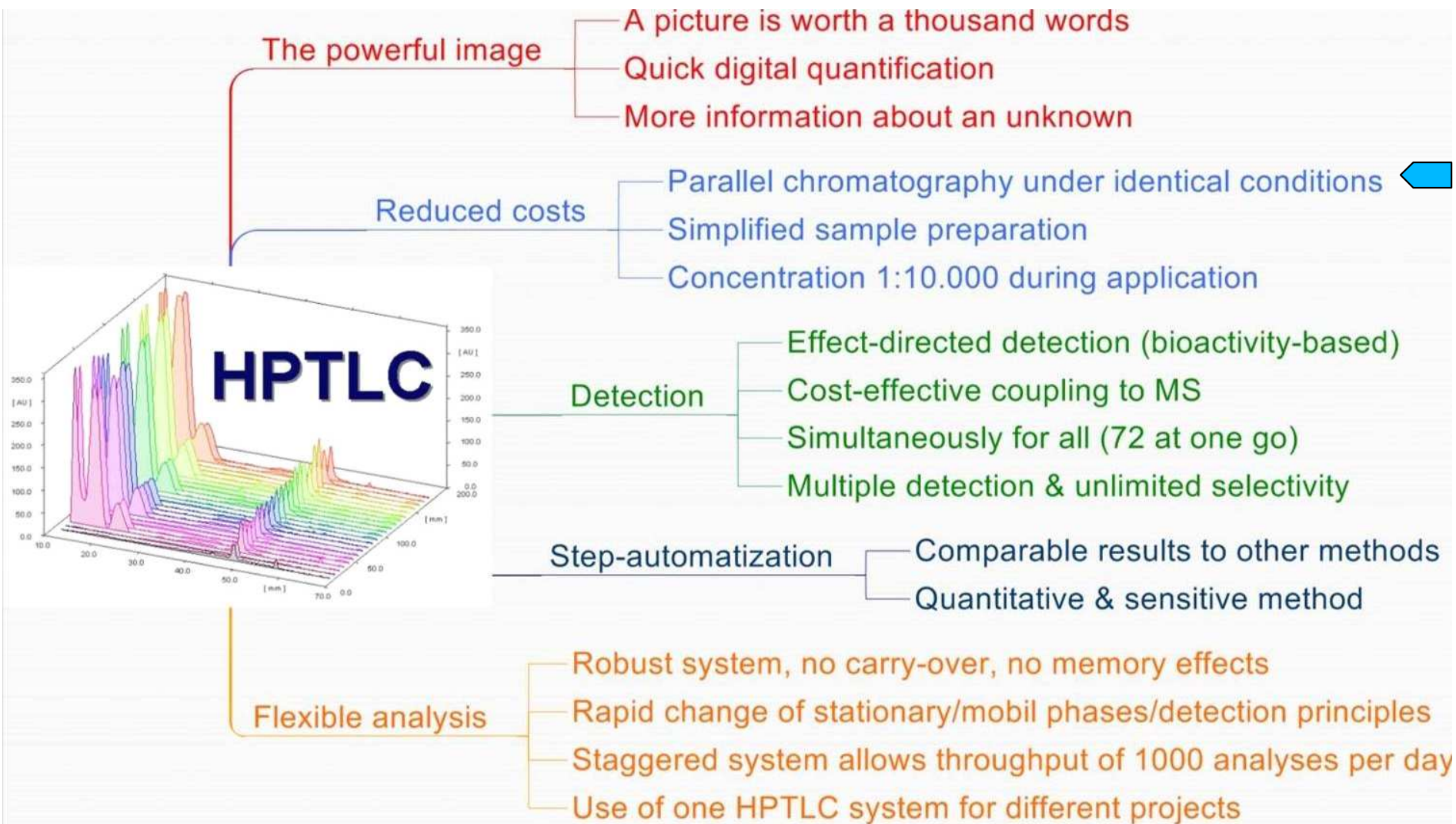


Mass imbalance caused by light degradation product

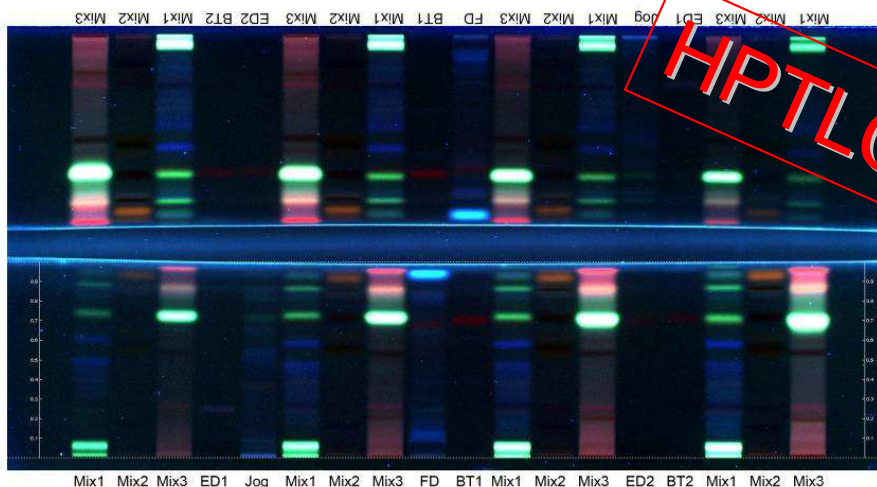
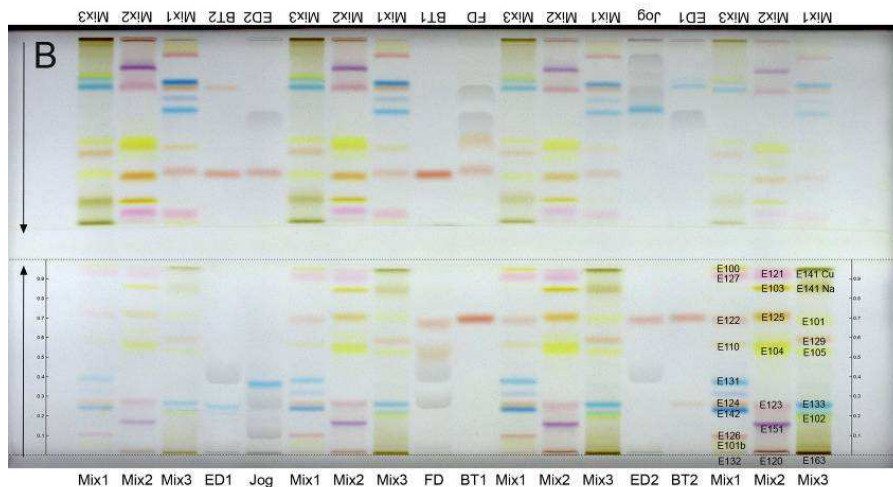
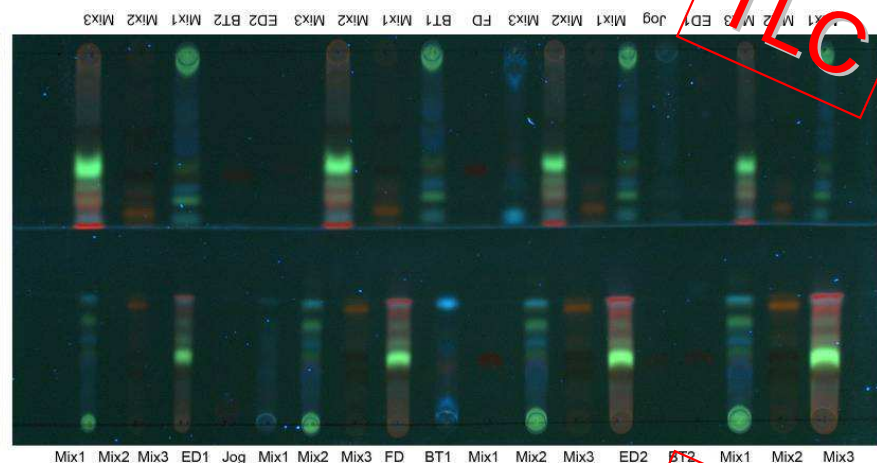
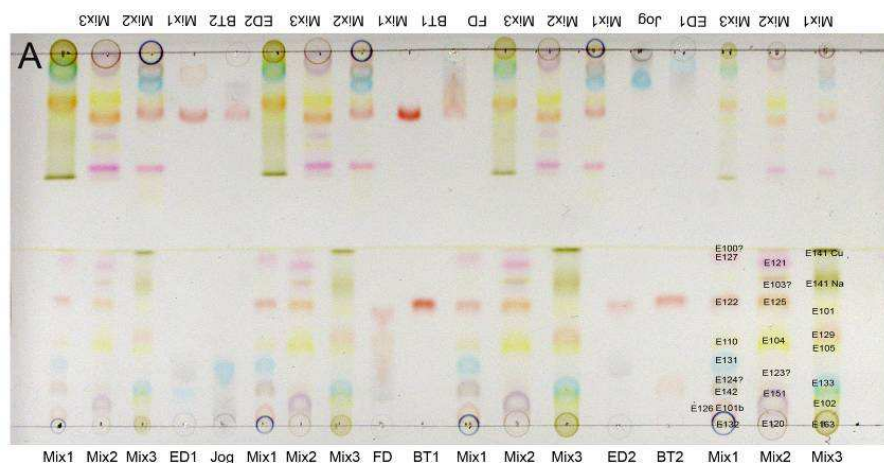


New yellow impurity caused color differences between batches of an API

Benefits

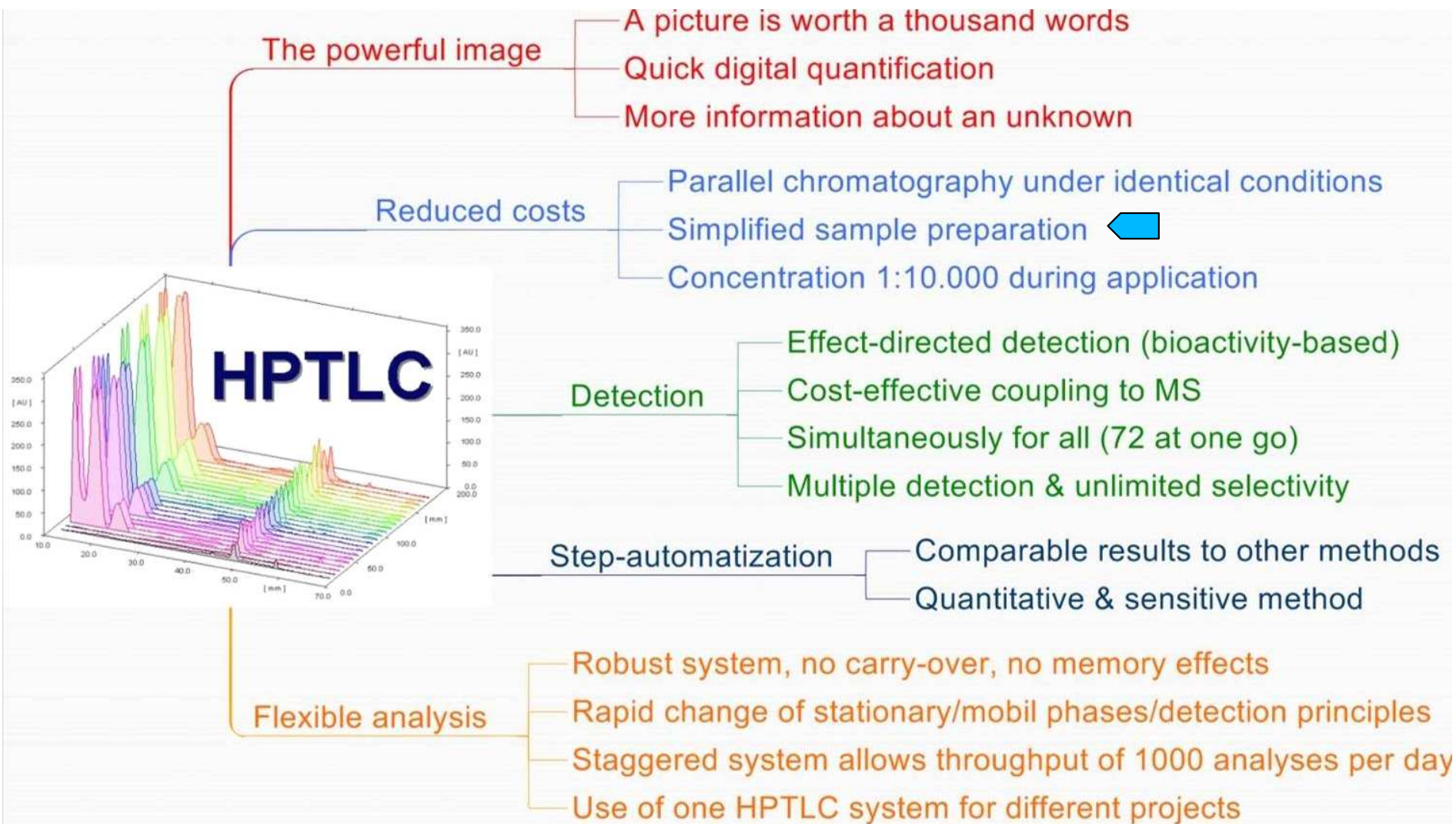


Dye analysis



- No reasonable calibration function was obtained by TLC.
- For quantification, just HPTLC is reliable.

Benefits

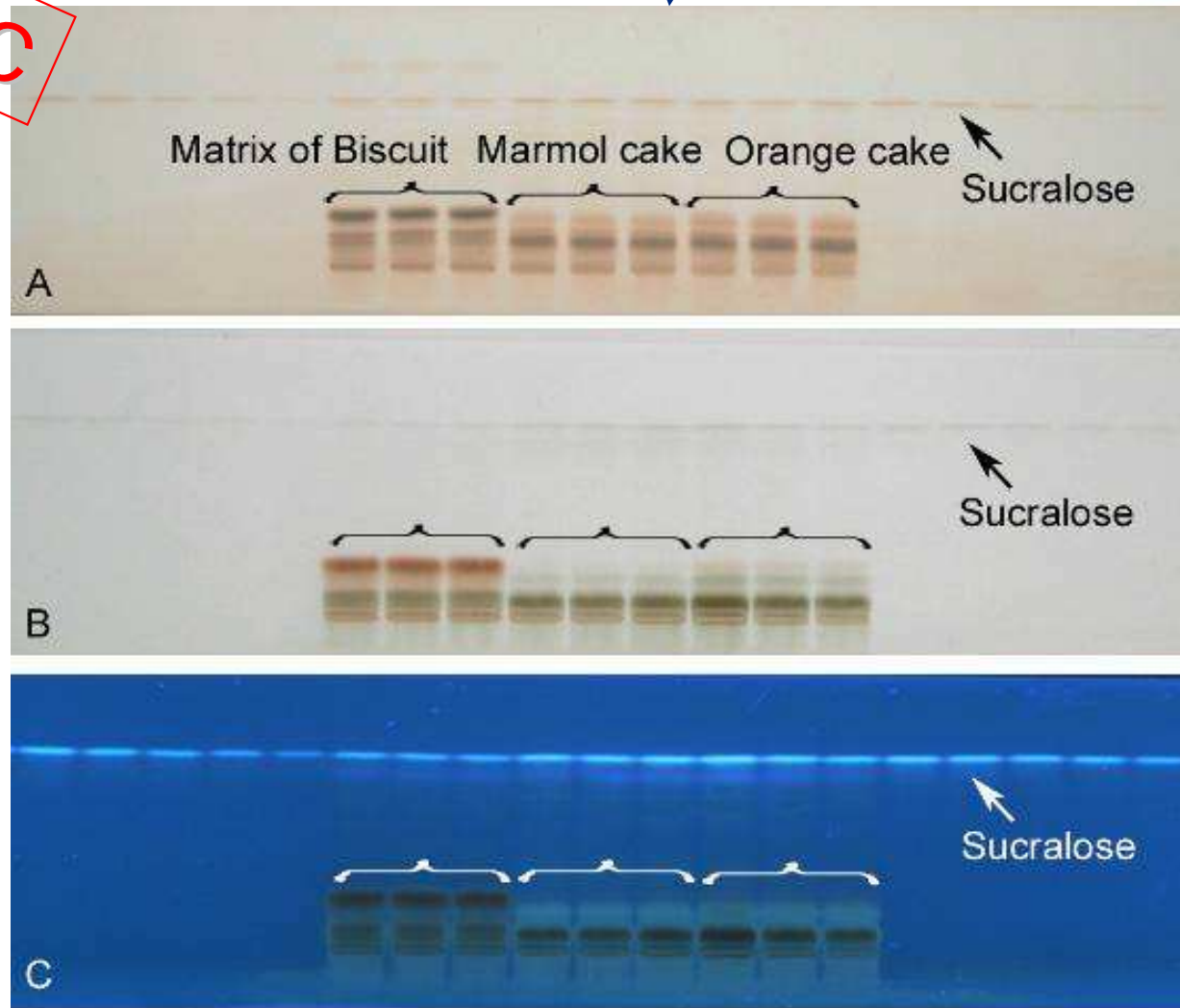




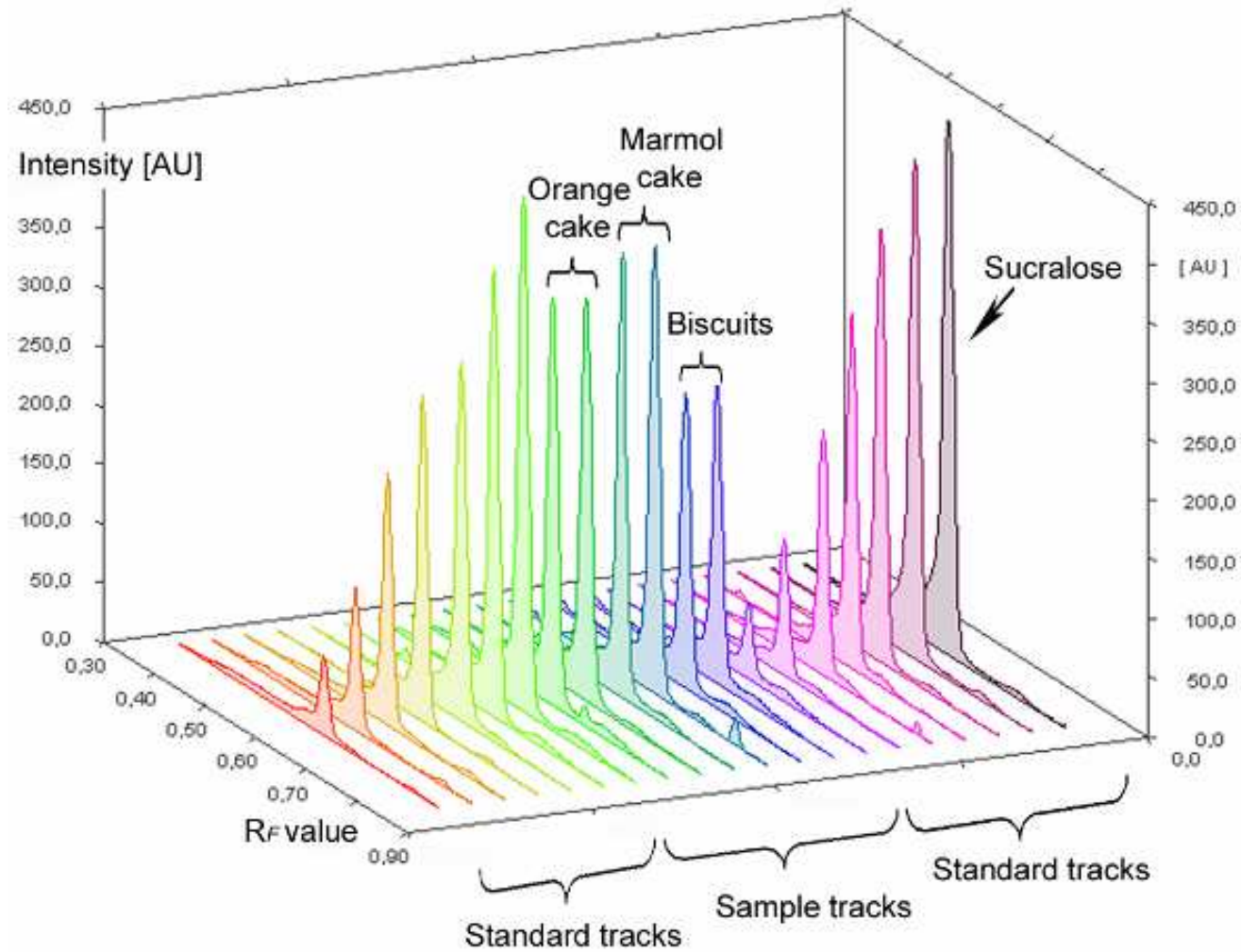
Analysis of sucralose in cakes

HPTLC

extracted
in MeOH, filtered



Quantification of sucralose in cakes





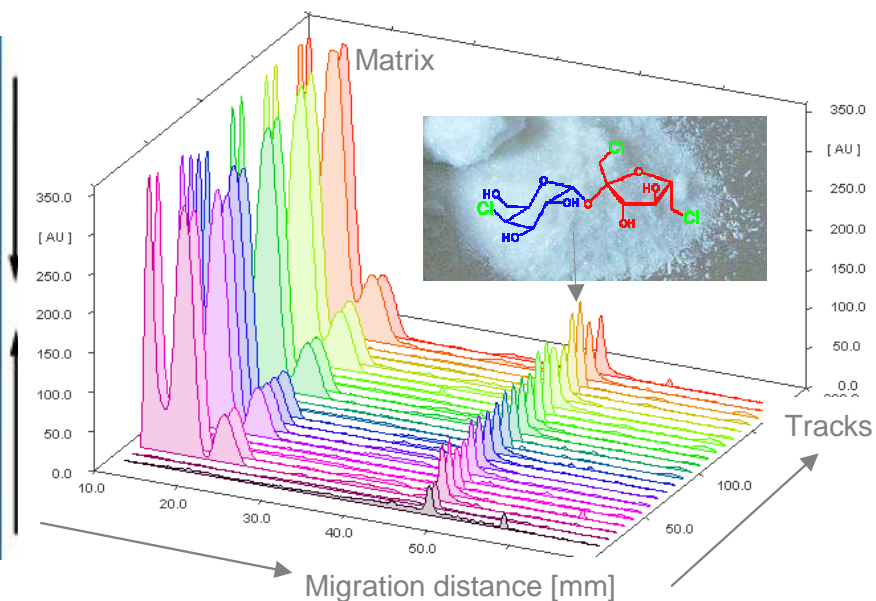
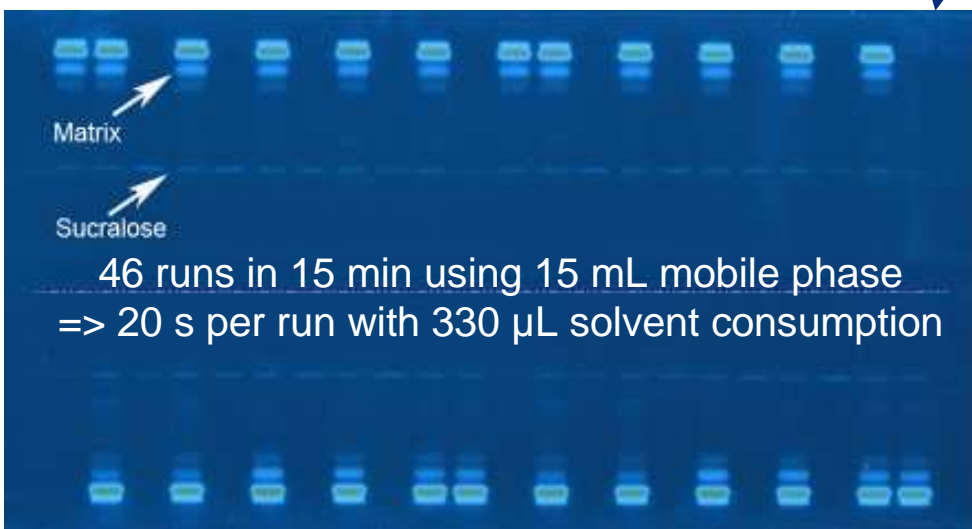
Quantification of sucralose in cakes

Mode A Reagent 1 @ 500 nm				
Samples	hR_F	Sucralose found (mg/100 g)	%RSD $n = 3$	Sucralose labeled (mg/100g)
Biscuits	57	27.7	2.4	24.8
Marmol cake	57	48.0	2.0	45.3
Orange cake	56	43.9	0.6	45.3
Mode B Reagent 2 @ 405 nm				
Biscuits	56	27.9	1.5	24.8
Marmol cake	56	47.4	0.5	45.3
Orange cake	56	44.2	1.6	45.3
Mode C Reagent 2 @ UV 366/>400 nm				
Biscuits	56	27.1	0.9	24.8
Marmol cake	57	44.8	4.2	45.3
Orange cake	56	41.6	3.0	45.3

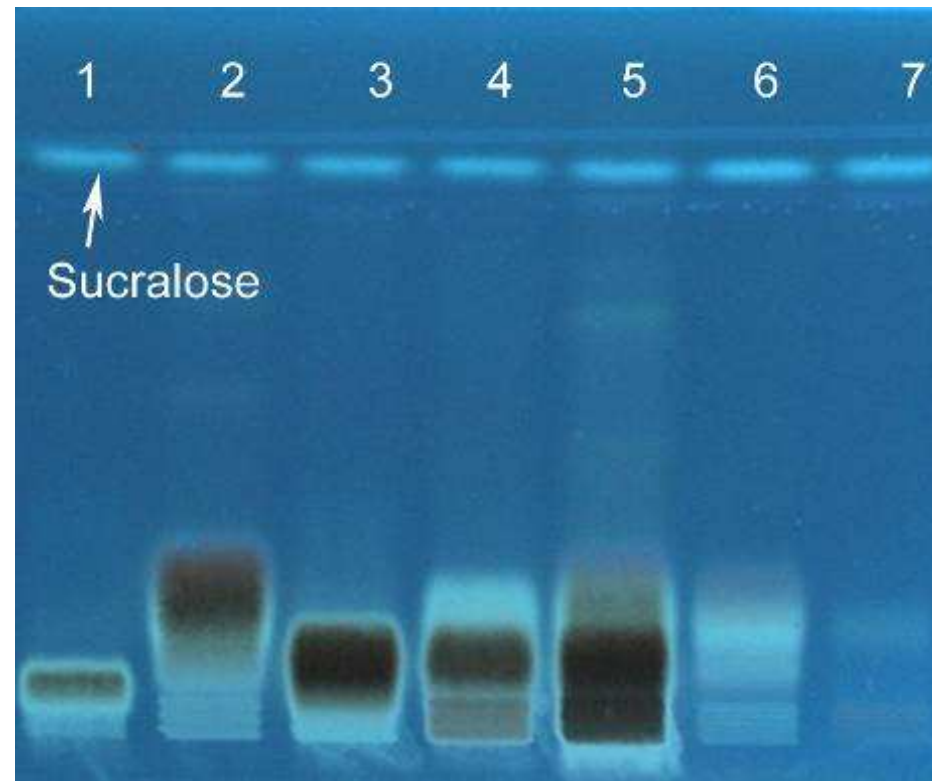
... in milk-based confection (Burfi)



extracted in MeOH, filtered

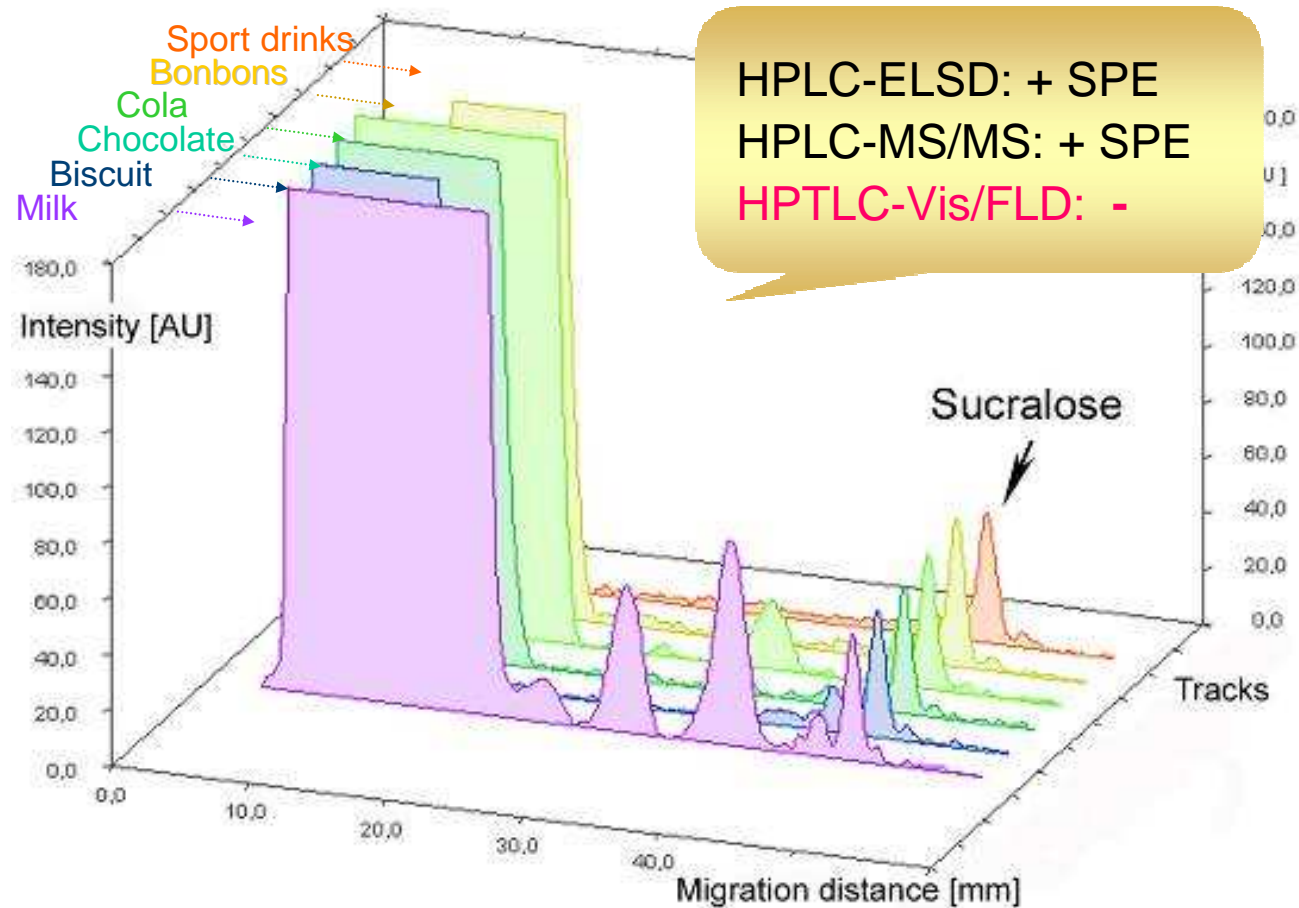


... in further matrices



Milk, biscuit, chocolate, cola, bonbons, energy/sport drinks

Sample preparation and chromatography





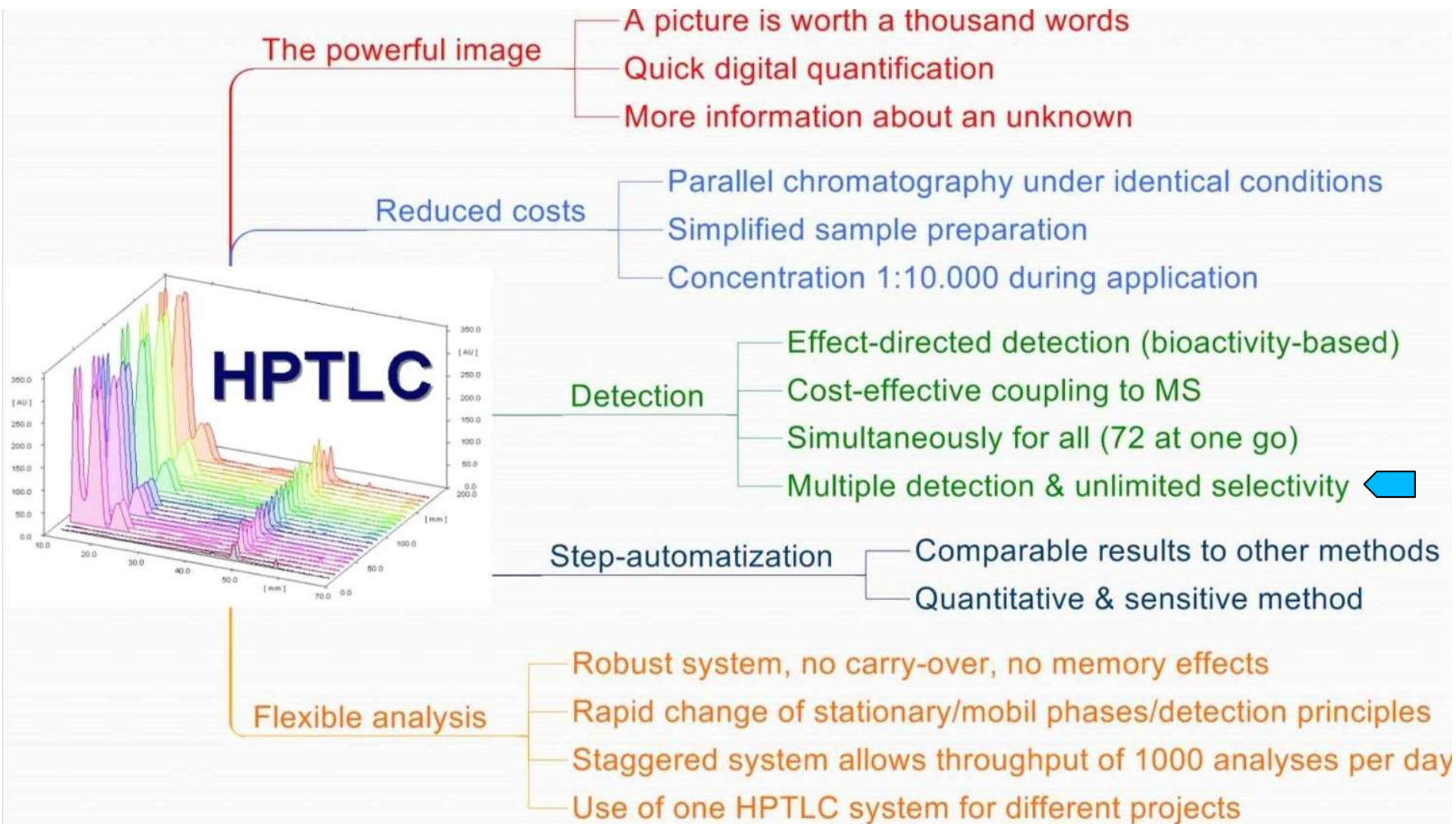
Facts for sucralose analysis

- High throughput (46 runs in 15 min by (anti-)parallel development, 15 min-staggered offline system) → 1000 runs/8h-day
- Resulting in 20-s runs with 330 µL solvent consumption
- Almost no disposal costs < 0.01 Cent/run
- Selective derivatization → compensates low separation power
- Reduced sample preparation: no SPE
- Analysis without acetonitrile!



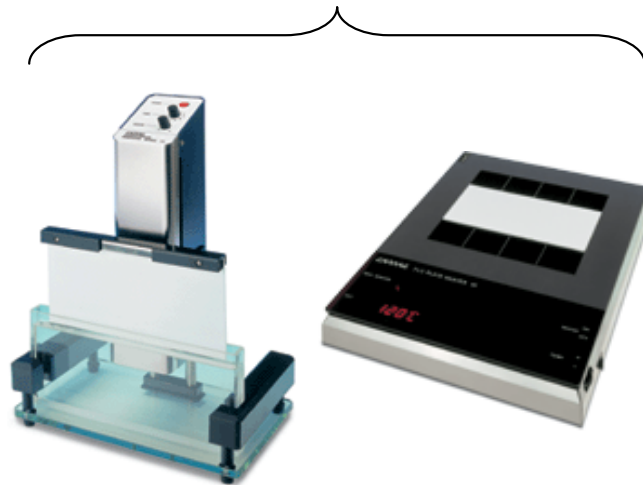
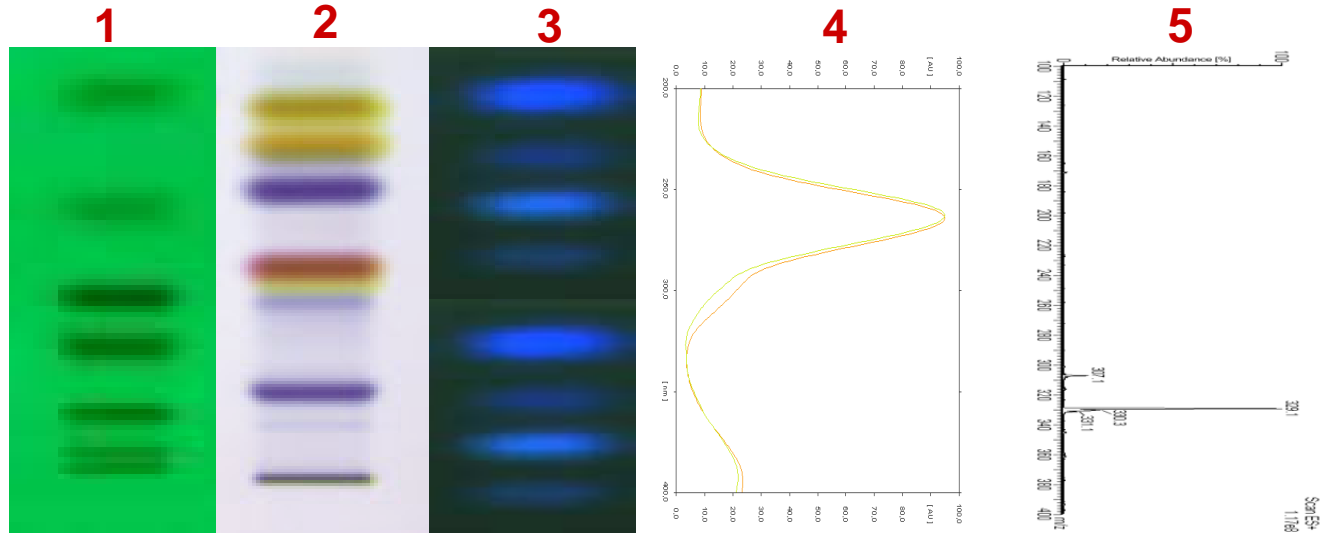
- Ultra-rapid HPLC with 2 min gradient: 720 runs/24-h day
- Sample preparation: Need of SPE for MS or ELSD as detector

Benefits



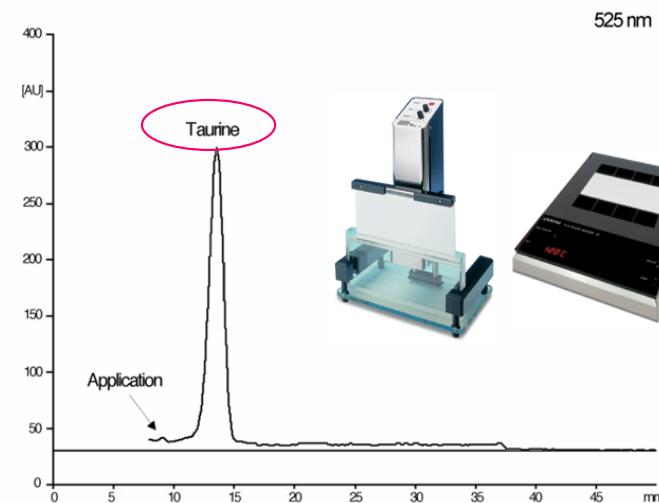
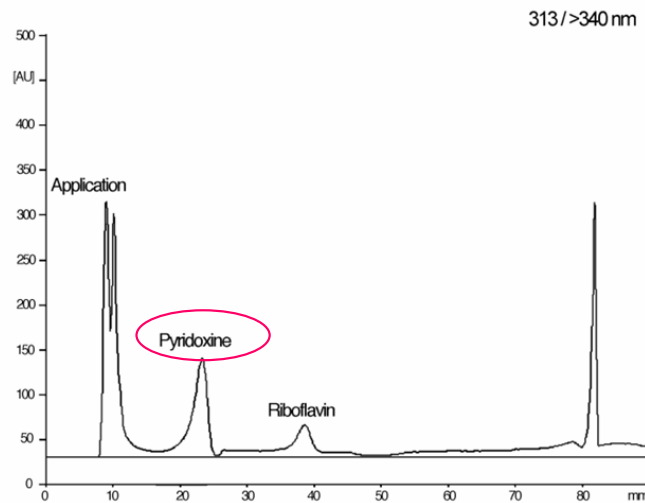
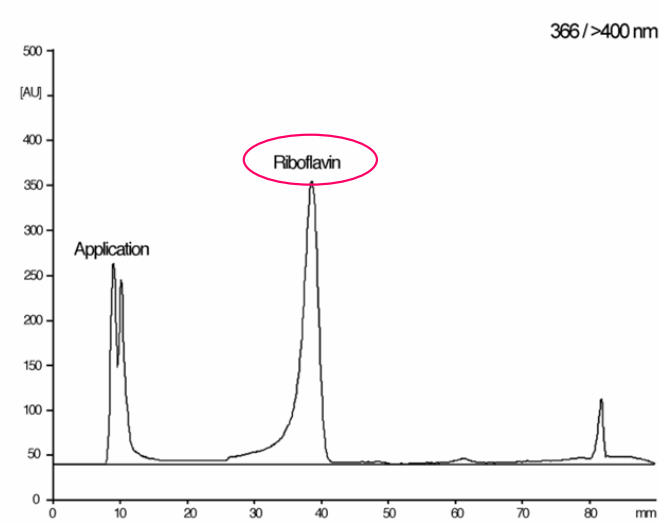
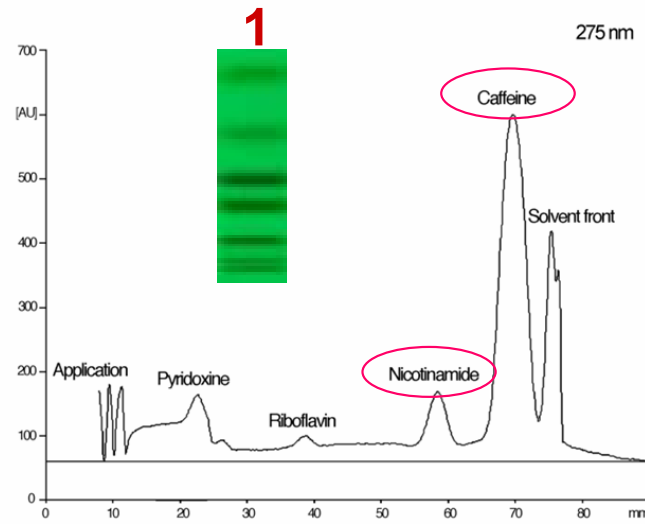
Multiple detection on a **single** track/plate

HPTLC-UV/Vis/FLD-MS



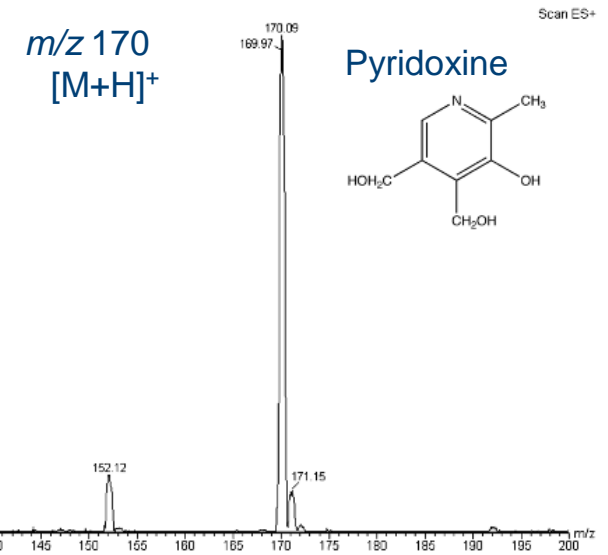
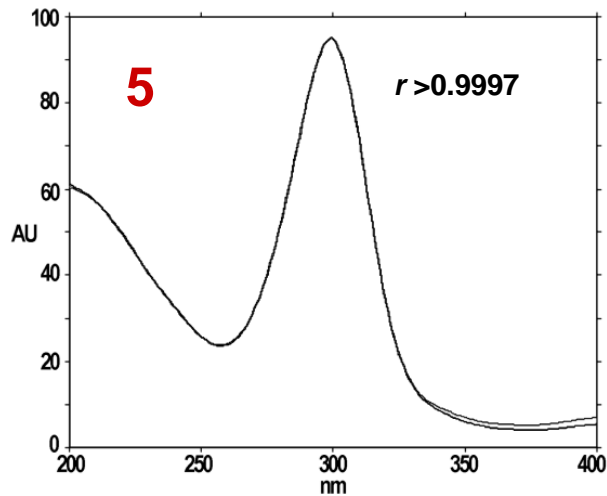
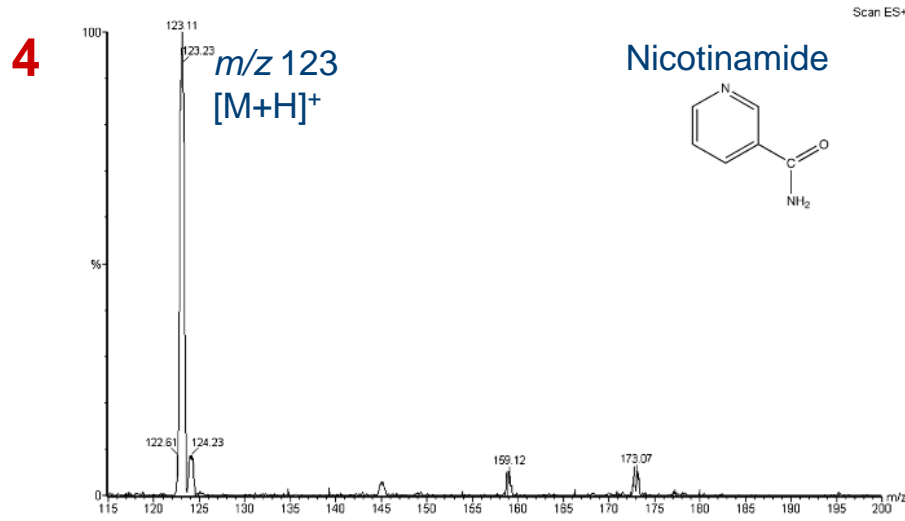
All active ingredients in energy drinks

Simultaneous determination by MWL scan for UV/FLD → derivatization → Vis...

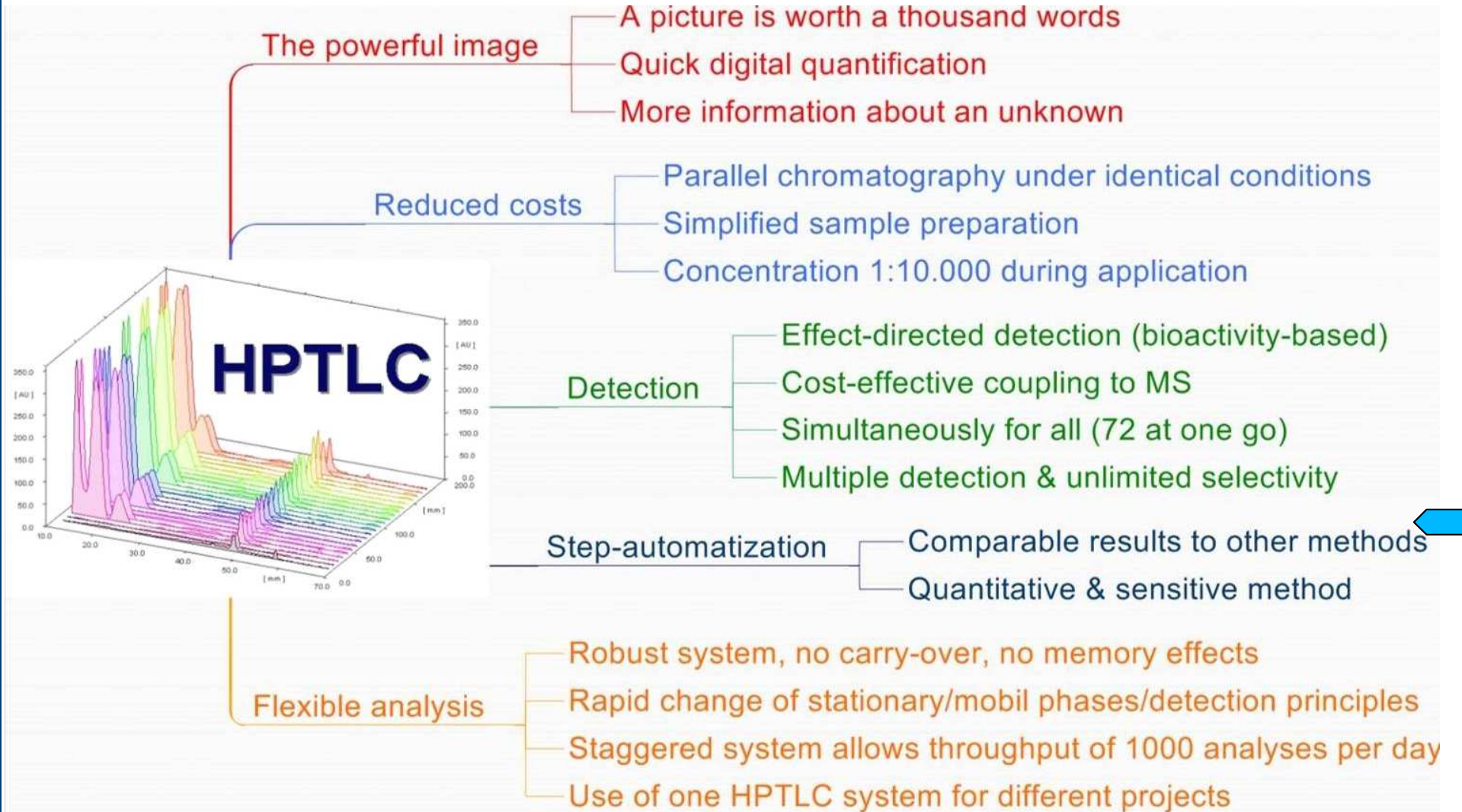


All active ingredients in energy drinks

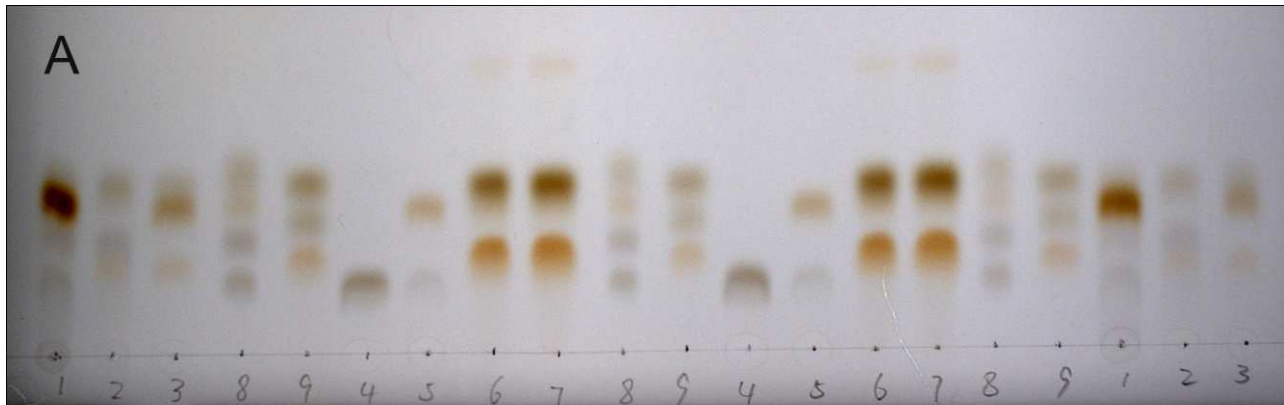
→ Confirmation by MS or UV spectra



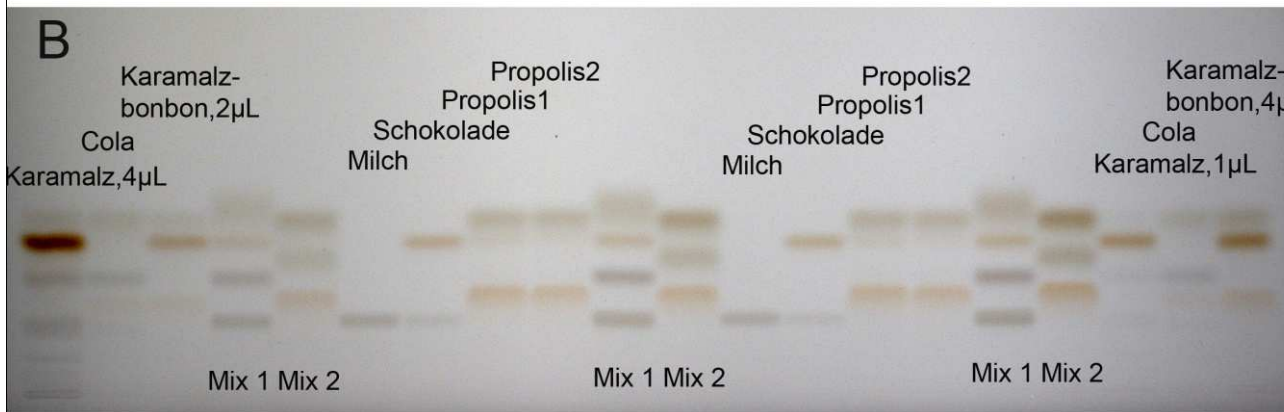
Benefits



Method comparison



Poor quantitative results



Good quantitative results



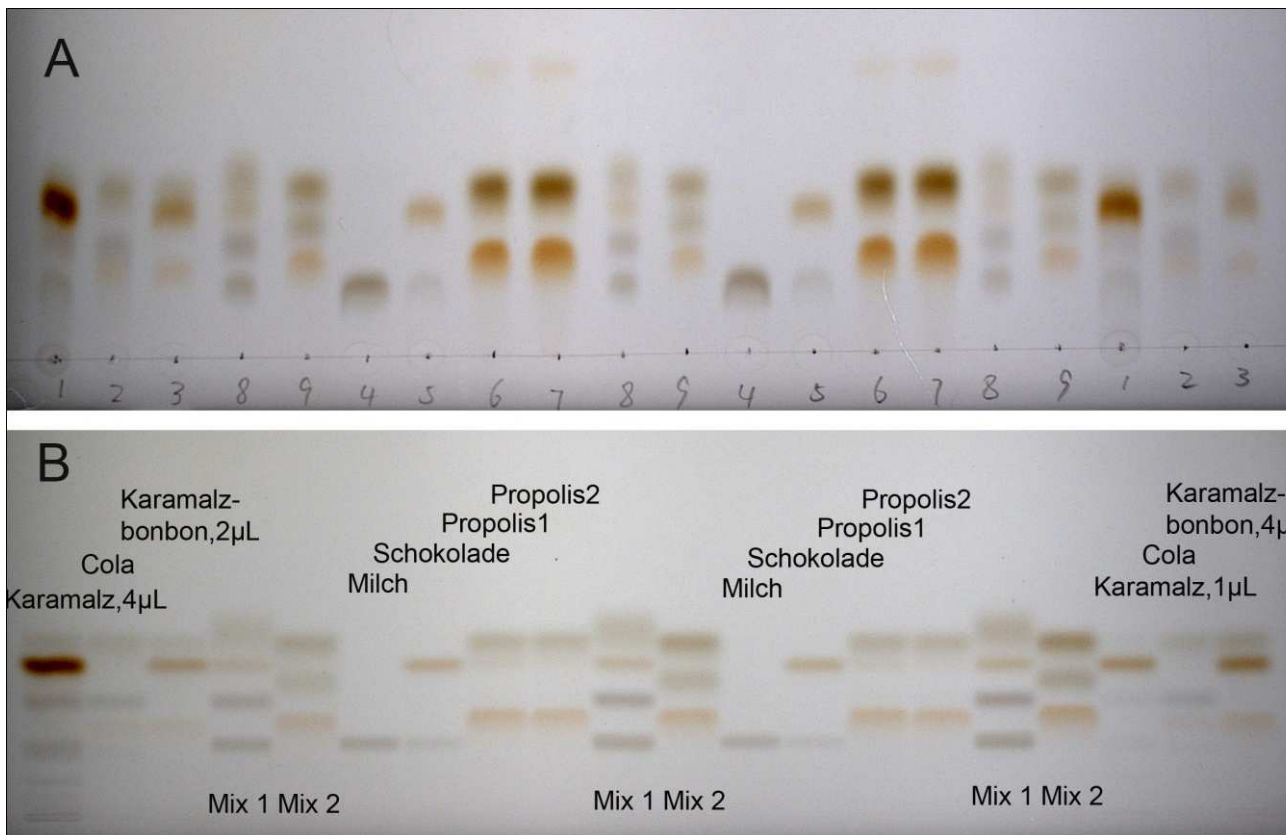
Comparable to HPLC-ELSD?



Method comparison

Sample	Sugar found (n = 2)	HPLC-ELSD		HPTLC-Vis	
		%	%RSD	%	%RSD
Cola	Sucrose	12.0	0.1	12.5	5.3
	Fructose	1.1	0.4	1.1	5.3
	Glucose	1.3	4.5	-	-
Milk	Lactose	8.0*	5.6	5.3	1.8
Chocolate	Sucrose	34.9	0.5	35.9	0.8
	Lactose	6.9	12.9	7.1	10.0
Propolis	Glucose	10.9	9.3	10.7	1.1
	Fructose	17.3	0.0	17.4	6.0
	Sucrose	4.4*	5.0	7.3*	7.0
Karamalt	Glucose	3.3	2.8	4.1	3.9
	Fructose	2.2	0.7	2.0	1.6
	Maltose	2.5	2.5	2.5	6.5
Biscuits	Sucrose	17.9*	1.2	23.9*	1.8

Method comparison



Poor quantitative results

Good quantitative results

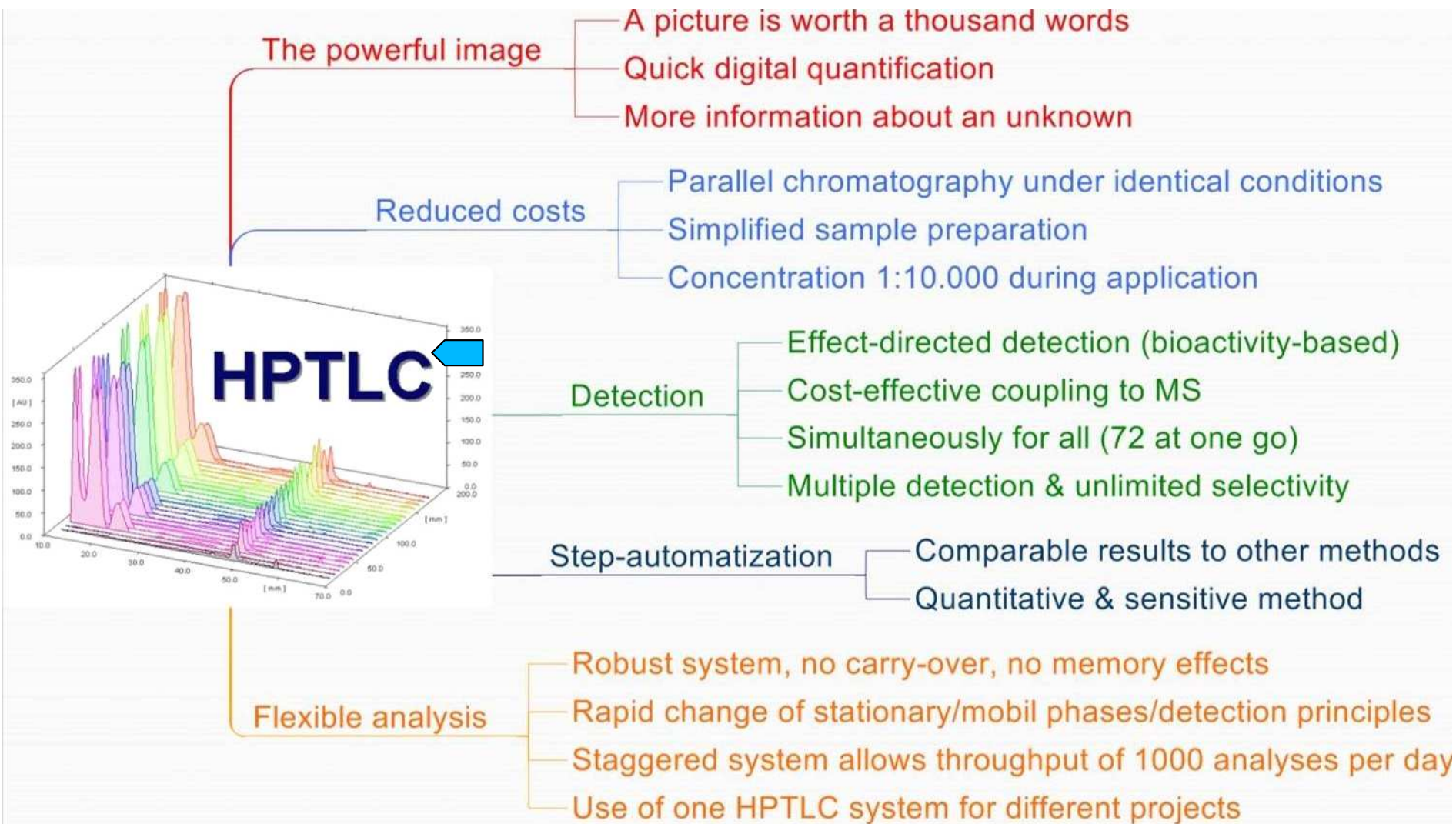


Comparable to HPLC-ELSD? => YES

Analyses time

- HPTLC: 1 h => 3 min per sample
- HPLC: 5.3 h => 16 min per sample

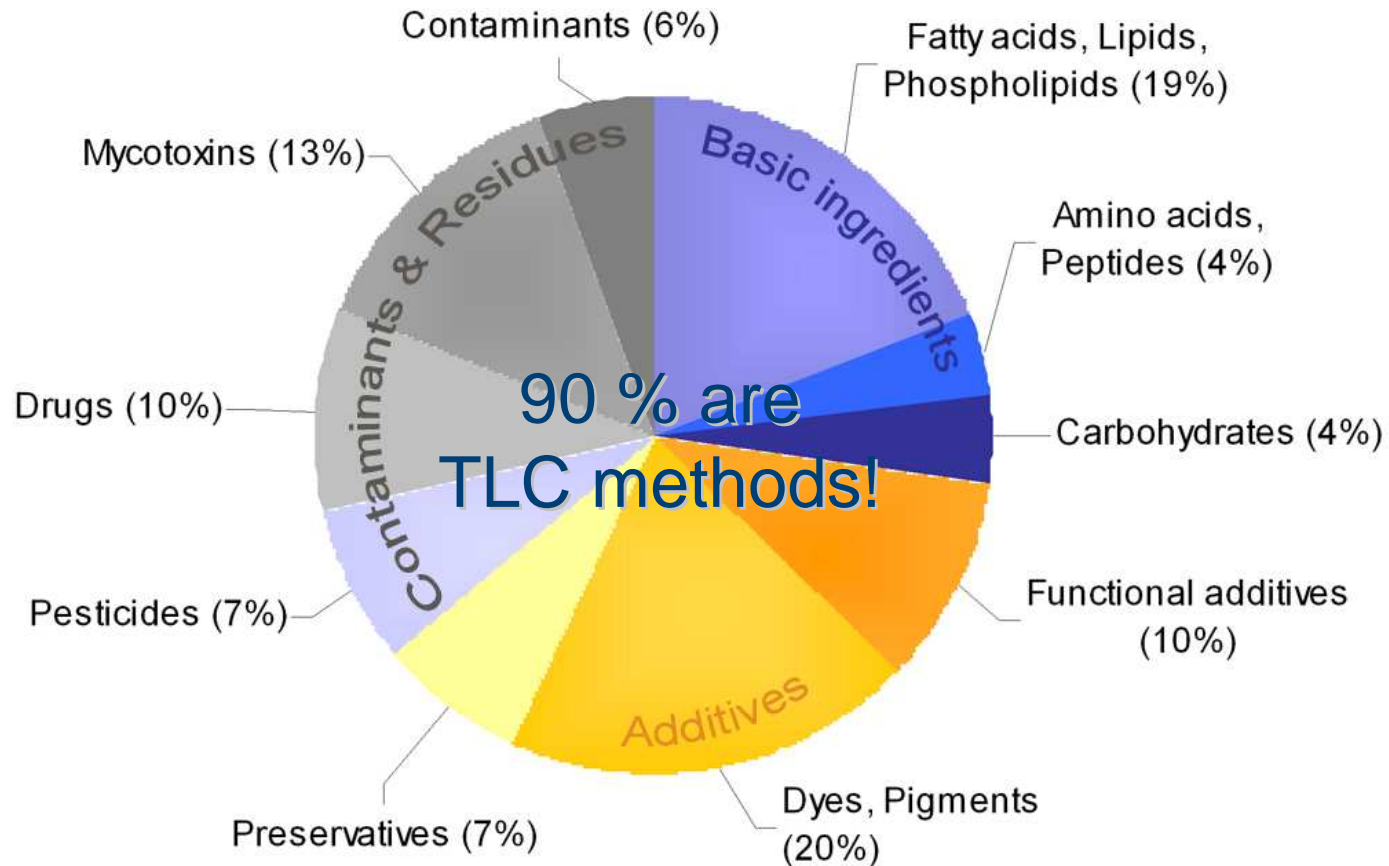
Benefits



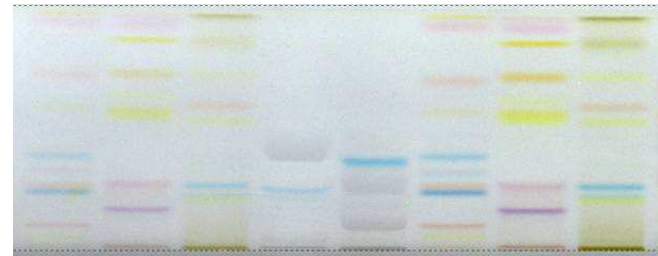
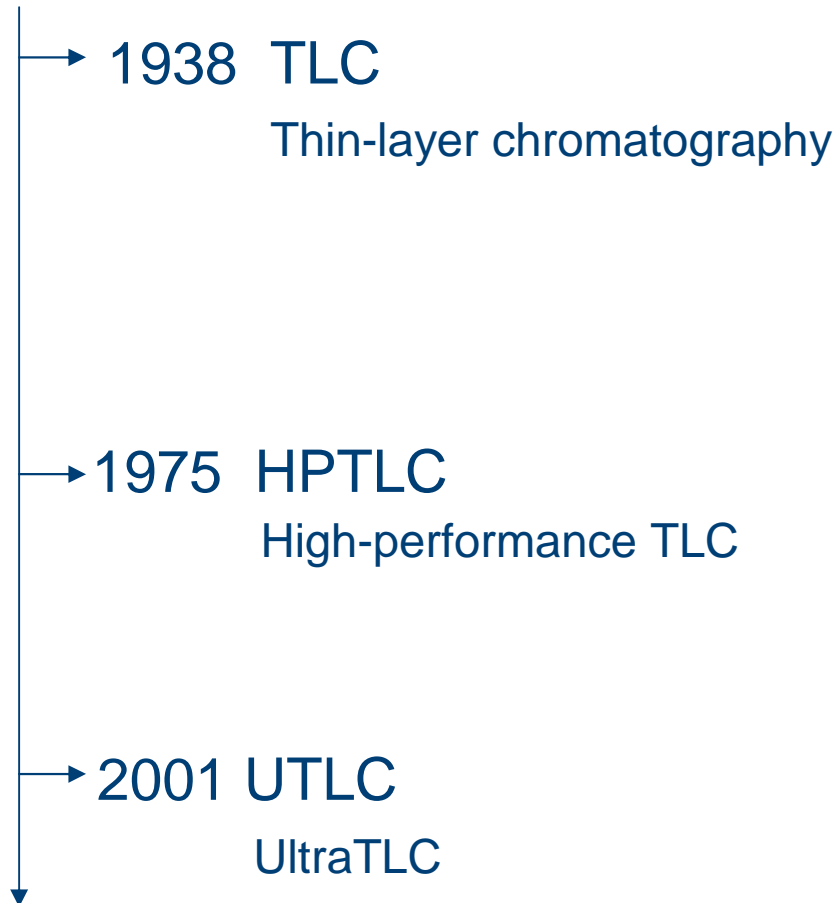


Planar Chromatography

Food analysis 1987-2007

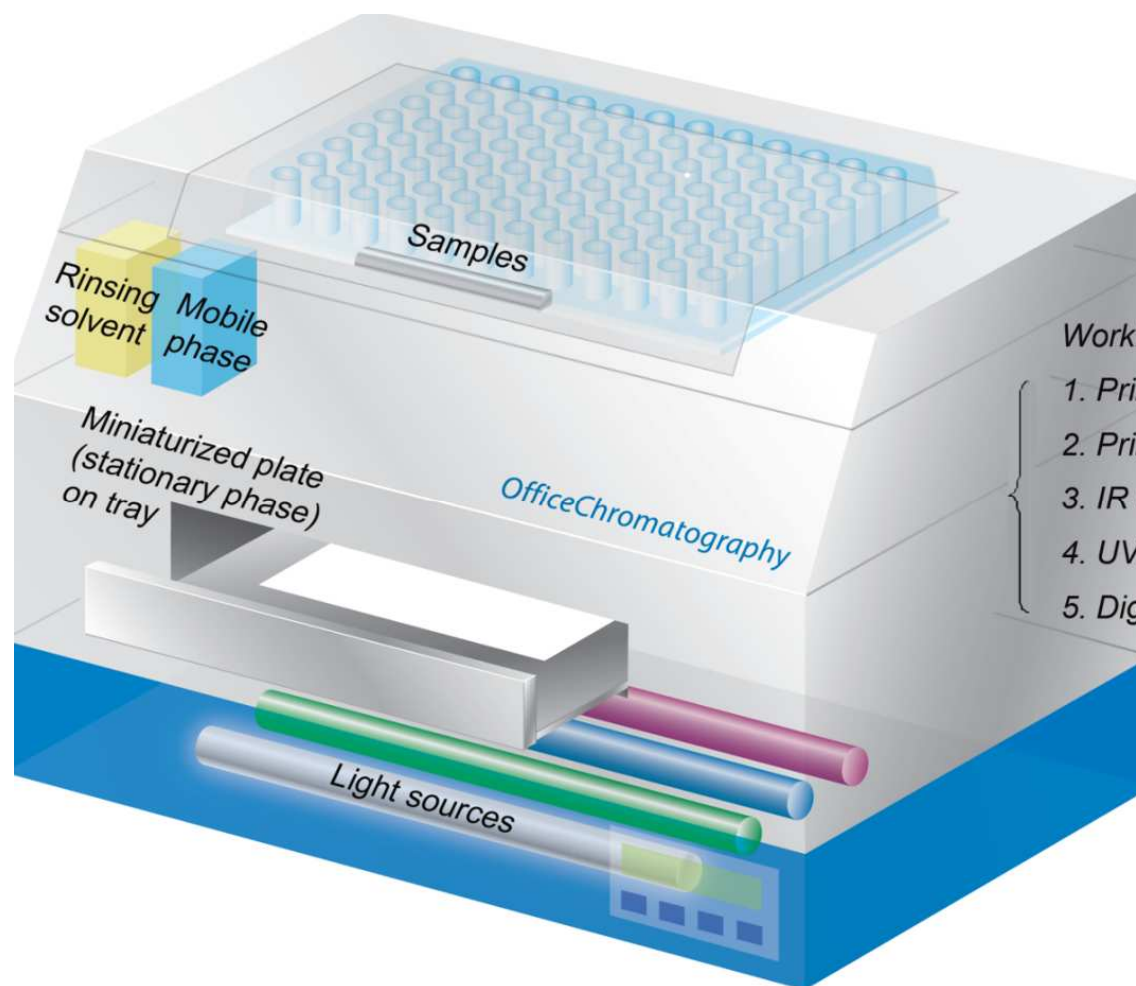


Planar Chromatography



Office Chromatography

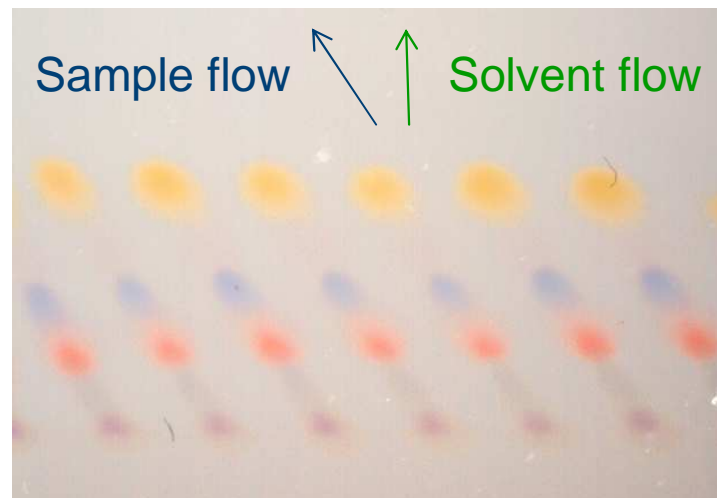
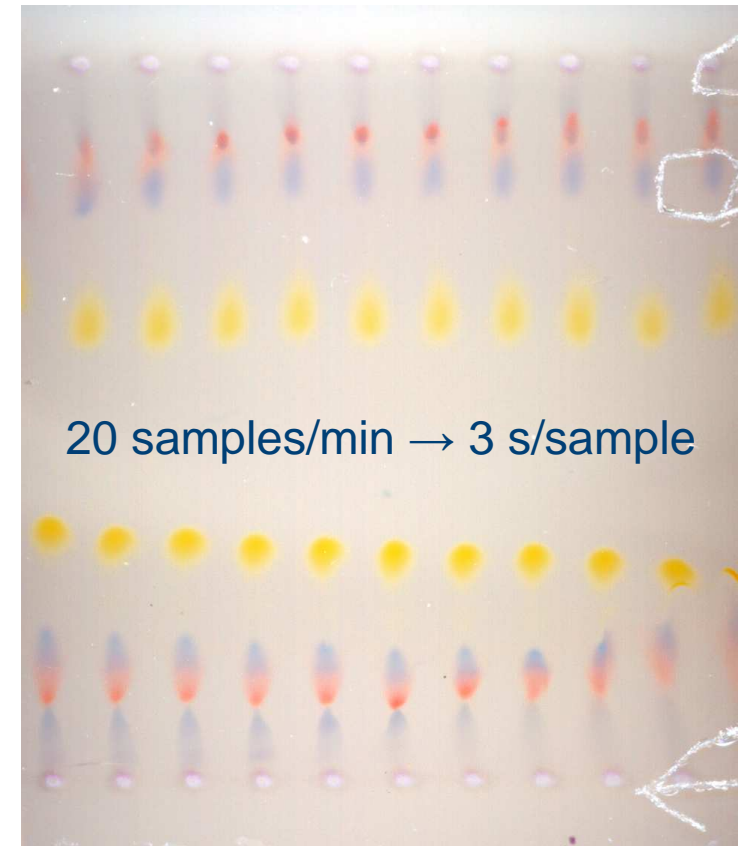
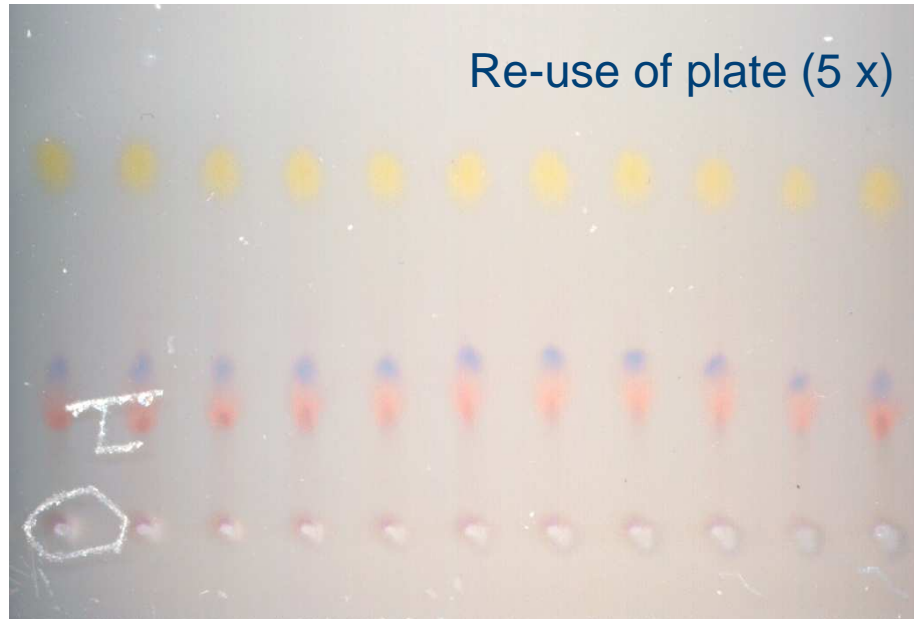
(Miniaturized) Planar chromatography using office peripherals



Workflow:

1. Printing of samples (μL)
2. Printing of mobile phase (μL)
3. IR scan for plate drying
4. UV/Vis scans for detection
5. Digital image evaluation

Nanostructured UTLC plates





Ultrathin plate (UTLC)



S. Jim, M. Taschuk, G. Morlock, L. Bezuidenhout, W. Schwack, M. Brett
Anal. Chem. 82 (2010) in print



Institute of Food Chemistry
University of Hohenheim, Stuttgart



*International Symposium for HPTLC
Basle, Tue 06th - Fri 08th July 2011*

HPTLC 2011, Basel : 6th-8th July 2011 → www.hptlc.com